

How to lose resistance to Aspergillus infections

Jean-Pierre Gangneux, Martin Hoenigl, Nicolas Papon

▶ To cite this version:

Jean-Pierre Gangneux, Martin Hoenigl, Nicolas Papon. How to lose resistance to Aspergillus infections. Trends in Microbiology, 2023, 31 (3), pp.222-224. 10.1016/j.tim.2023.01.008 . hal-03994677

HAL Id: hal-03994677 https://univ-angers.hal.science/hal-03994677v1

Submitted on 17 Apr 2023

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial 4.0 International License

1	SPOTLIGHT
2	Tentative title:
3	How to Lose Resistance to Aspergillus Infections
4	Alternative titles (at your convenience):
5	• How to Lose Resistance to Aspergillus Infections if your Neutrophils are Intact
6	o Unravelling the Mechanism of Aspergillosis in non-Neutropenic ICU Patients
7	
8	Jean-Pierre Gangneux ^{1,2*} , Martin Hoenigl ^{3,4,5} , Nicolas Papon ⁶
9	
10	¹ Inserm, EHESP, IRSET (Institut de Recherche en Santé, Environnement et Travail)-UMR_S 1085, CHU Rennes, Univ Rennes, F-35000
11	Rennes, France.
12 13	² Laboratory of Parasitology and medical mycology, European Confederation of Medical Mycology (ECMM) Excellence Center, Rennes Teaching Hospital, E-35000 Rennes, France
14	³ Division of Infectious Diseases, Department of Internal Medicine, Medical University of Graz, Graz, Austria.
15	⁴ BioTechMed-Graz, Graz, Austria.
16	⁵ Translational Medical Mycology Research Unit, European Confederation of Medical Mycology (ECMM) Excellence Center, Medical
17	University of Graz, Graz, Austria. ⁶ Univ Angers, Univ Brest, IRE, SER ICAT, E-49000 Angers, France
19	
20 21	Correspondence: jean-pierre.gangneux@univ-rennes1.fr (JP. Gangneux)
22	ORCID:
23	
24	JP. Gangneux: <u>https://orcid.org/0000-0002-4974-5607</u>
25	M. Hoenigl: https://orcid.org/0000-0002-1653-2824
26 27	N. Papon: <u>https://orcid.org/0000-0001-6265-7321</u>
28	Twitter:
29	
30 21	JP. Gangneux: @GangneuxJp
31 32	M. Hoenigi: @martinnoenigi N. Papon: @NicolasPapon49
33	
34	COI: MH reports grants and research funding from Astellas, Gilead, MSD, Pfizer, Euroimmun, F2G, Pulmocide, IMMY, Mundipharma and
35	Scynexis. JPG reports grants and research funding from Gilead, Pfizer, and Mundipharma.
36	
3/	
38	
39 40	Abstract
40 41	The distinct risk factors to deadly infections by Aspergillus fumigatus in ICU patients are
42	well known, however, the mechanistic link between these predisposing conditions was
43	unknown so far. Sarden et al. recently unraveled a shared B1a lymphocytes-natural
44	antibodies-neutrophils defense pathway to A. fumigatus, opening new perspectives in
45	diagnostics and therapeutics.
46	

Fungal infections represent a global burden worldwide. While superficial fungal infections account for a large proportion of the overall worldwide prevalence of mycoses and fortunately are rarely fatal, invasive fungal infections are characterized by dramatic high morbidity, mortality, and economic burden. Invasive fungal infections are killing approximately 1.5 million people annually [1]. To tackle the steady increase of invasive fungal infections, the WHO provided recently a first global response to identify and prioritize fungal pathogens according to their impact on global public health for considering the unmet research and development needs [2].

55 Among the list of four fungal species of critical importance topping the WHO Fungal Priority Pathogens List stands Aspergillus fumigatus (Af), one of the most ubiquitous of the airborne 56 57 saprophytic fungi [3]. By filtering more than 10,000 liters of unsterile air daily, our lungs inhale thousands of Af spores. Although mucociliary clearance ensures an efficient mechanical expulsion of 58 a significant amount of these particles, the small spores of *Af* can easily get stuck in alveoli gaps. 59 Yet it is intriguing that most humans do not develop any illness attributable to Af and do not display 60 any evidence of antibody- or cell-mediated acquired immunity to this microorganism in case of 61 62 asymptomatic carriage. This teaches us that, for most healthy individuals, the local and innate immune response is sufficient to clear the organism without requiring intervention of an acquired 63 immunity [4]. 64

65 Risk factors predisposing to Af infections are now well-recognized, in particular defective or deficient neutrophils, underlying the central role of these white cells as first line defense for naturally 66 67 combating *Af.* However, the fact that non-neutropenic intensive care unit (ICU) patients receiving long-term systemic corticosteroid therapy and even immunocompetent people once affected by 68 69 acute respiratory failure driven by influenza A virus (IAV) and acute respiratory syndrome coronavirus 2 (SARS-CoV-2) can also develop life-threatening Af infections suggest that other 70 71 contributors than neutrophils may act to maintaining essential pulmonary host resistance to this mold [5-8]. While the diverse risk factors to losing defenses to *Af* and the essential role of neutrophils 72 in killing this fungal pathogen are well ingrained in the literature, the mechanistic relationship that 73 could link and explain the diverse predisposing conditions to losing resistance to deadly Af infections 74 remains unknown. In such perspective, unprecedented insights into a shared host resistance 75 76 pathway disrupted during viral and steroid-associated aspergillosis were gained recently through a 77 remarkable report by the research group of Bryan G. Yipp published in *Science Translational Medicine* 78 [9].

79

80 In a first series of investigations, Sarden and colleagues demonstrated that sera from 81 individuals with either severe IAV or SARS-CoV-2 infection had decreased concentrations of specific 82 anti-A. fumigatus IgG (anti-Af IgG). By developing an IAV/Af coinfection mouse model, the 83 pathophysiological relevance was further underscored by revealing that these viral infections trigger a decrease in circulating anti-Af IgG and impair resistance to Af in vivo. In addition, by taking 84 advantage of an Affluorescent reporter strain, Sarden et al. provided evidence that lung neutrophils 85 are hampered in their capacity to capture fungal spores while neutrophil recruitment was found 86 unchanged in the context of a IAV/Af co-infection. 87

Because healthy people display innate resistance to *Af* in the absence of prior infection or immunization, the research group of Bryan G. Yipp then postulated that natural antibodies produced by B1 innate lymphocytes may regulate neutrophil-mediated phagocytosis *in vivo*. In this respect, by combining different models of infection in various mice genetic backgrounds, serum transfer 92 experiments, monoclonal antibody administration, and transcriptional profiling of donor samples, 93 investigators elegantly validated this hypothesis. It is noteworthy that decrease in anti-*Af* IgG stands 94 out as a common and conserved mechanism of acquired host susceptibility to invasive aspergillosis 95 since steroid treatment also diminished these specific antibodies in mice. Importantly, further 96 experiments revealed that this decreased serum concentration in anti-*Af* IgG during viral infection 97 (IAV or SARS-CoV-2) or following corticosteroid treatment commonly derived from the depletion of 98 circulating B1 lymphocytes in patients.

99 In a last series of experimentations, Sarden and colleagues aimed at identifying the essential host resistance pathway that is impaired during SARS-CoV-2 for explaining the high prevalence of 100 COVID-19-associated pulmonary aspergillosis (CAPA) [6,7]. Following the observation that SARS-101 CoV-2 spike (S) protein triggers B1a apoptosis *in vivo*, investigators engineered a Vesicular stomatitis 102 virus expressing the SARS-CoV-2 S protein (VSV-GS pseudovirus) for developing a mouse model 103 104 mimicking CAPA. Co-infection of wild-type mice by VSV-GS pseudovirus and Af resulted in high mortality associated to a marked decrease in serum anti-Af IqG levels, similarly to that observed in 105 106 the IAV-aspergillosis model. This added string evidence that the break in *Af* resistance during CAPA is conditioned by a loss in B1a cells and natural antibodies. 107

Overall, all the data compiled in this enlightening article converge toward the unifying evidence that during infection with IAV or SARS-CoV-2 or following corticosteroid treatment, *Af* innate immunity is disrupted and results from the death of B1a lymphocytes leading in turn to depletion of anti-*Af* IgG thereby allowing the fungus to remain concealed from recruited lung neutrophils, leaving the door open for fungal germination and tissue invasion (**Figure 1**).

113

To summarize, Sarden and colleagues findings highlight a new B1a lymphocytes - natural 114 anti-Af IqG - neutrophils axis governing the innate resistance to one of the most critical fungal 115 pathogens (**Figure 1**) [9]. Above all, it is important to applaud the Bryan G. Yipp research group 116 for having focused their investigations toward a putative role of B-cells in the defense toward Af 117 infection. This approach contrasted most of the existing literature on antifungal immunity, focusing 118 primarily on T-cells [10]. The identification of B1a cells and anti-*Af* IgG as essential components of 119 120 the host innate immunity to Af that are lost during IAV and SARS-CoV-2 but also after corticosteroid treatment nicely complement the established knowledge on neutrophils as a key cell population for 121 host defense against *Af* (**Figure 1**). In fact, the relevance of neutrophils for protection against fatal 122 aspergillosis was inferred in early descriptions of invasive pulmonary aspergillosis because of its 123 124 prevalence in individuals with impaired neutrophil numbers or functions such as neutropenic patients, those with inherited immune deficiency or chronic granulomatous disease, and those 125 receiving allogeneic stem cell transplants (Figure 1). The seminal study by Sarden at al. provides 126 127 original mechanistic explanation for resistance breakdown to aspergillosis in at least three distinct clinical contexts. The identification of new actors adding to neutrophils and the discovery of an 128 essential cooperative pathways involved in natural defense against Af now unveils the functional 129 relationship that could link and explain a multitude of diverse risk factors predisposing to this life-130 threatening fungal infection. Finally, this groundbreaking work not only opens exciting perspectives 131 132 for diagnostics considering the possibility to stratify the risk of fatal aspergillosis by determining the concentration of anti-AfIgG in the serum of patients but also propels future hope in therapeutics by 133 using pooled sera from healthy donors or biotechnologically-produced anti-Af IgG. 134 135

References 137

- 138
- 139 1. Bongomin F. et al. (2017) Global and Multi-National Prevalence of Fungal Diseases-Estimate Precision. J. Fungi 3, 57
- 140 2. WHO. (2022) WHO fungal priority pathogens list to guide research, development and public health action. https://www.who.int/publications/i/item/9789240060241. 141
- 142 3. Wassano, N.S., Goldman, G.H., Damasio, A. (2020) Aspergillus fumigatus. Trends Microbiol. 28, 594-595
- 143 4. Latgé J.P., Chamilos G. (2019) Aspergillus fumigatus and Aspergillosis in 2019. Clin. Microbiol. Rev. 33, e00140-18.
- 144 5. Feys S. et al., (2022) Lung epithelial and myeloid innate immunity in influenza-associated or COVID-19-associated pulmonary 145 aspergillosis: an observational study. Lancet Respir. Med. 10, 1147-1159
- 146 6. Gangneux J.P. et al., (2022) Fungal infections in mechanically ventilated patients with COVID-19 during the first wave: the French multicentre MYCOVID study. Lancet Respir. Med. 10, 180-190. 147
- 148 7. Hoenigl M., et al., (2022) COVID-19-associated fungal infections. Nat. Microbiol. 7: 1127-1140
- 149 8. Salazar F., et al., (2022) Pathogenesis of Respiratory Viral and Fungal Coinfections. Clin. Microbiol. Rev. 35, e0009421.
- 150 9. Sarden N. et al., (2022) A B1a-natural IgG-neutrophil axis is impaired in viral- and steroid-associated aspergillosis. Sci. Transl. Med. 151 14, eabq6682
- 152 10. Lionakis M.S., Drummond R.A., Hohl T.M. (2023) Immune responses to human fungal pathogens and therapeutic prospects. Nat. 153 Rev. Immunol. doi: 10.1038/s41577-022-00826-w.
- 154



156

Figure 1. A new core resistance pathway to Aspergillus fumigatus (Af) infections. Lungs inhale thousands of Af 157 158 spores daily. However, healthy humans do not develop any illness attributable to Af. Risk factors predisposing to Af infections are now well-categorized and include primarily immunosuppressed patients such as those for instance with 159 160 neutropenia, undergoing allogeneic stem cell transplantation (SCT), or with chronic granulomatous disease (CGD) (right upper panel). T-cell suppression may also play a role in other patient groups (not shown). Immunocompetent patients 161 162 receiving long-term systemic corticosteroid therapy or affected by pneumonia driven by SARS- IAV and CoV-2 (left upper 163 panel) can also develop life-threatening Af infections. In their new report, Sarden and colleagues detailed the mechanistic 164 relationship that explain the diverse predisposing conditions to losing resistance to deadly Af infections by highlighting a 165 B1a lymphocytes - natural anti-Af IgG - neutrophils axis (lower panel) governing the innate resistance to this prominent 166 fungal pathogen.