



HAL
open science

Assessment of SARS-CoV-2 serological tests for the diagnosis of COVID-19 through the evaluation of three immunoassays: Two automated immunoassays (Euroimmun and Abbott) and one rapid lateral flow immunoassay (NG Biotech)

Thomas Nicol, Caroline Lefevre, Oriane Serri, Adeline Pivert, Françoise Joubaud, Vincent Dubée, Achille Kouatchet, Alexandra Ducancelle, Françoise Lunel-Fabiani, Hélène Le Guillou-Guillemette

► To cite this version:

Thomas Nicol, Caroline Lefevre, Oriane Serri, Adeline Pivert, Françoise Joubaud, et al.. Assessment of SARS-CoV-2 serological tests for the diagnosis of COVID-19 through the evaluation of three immunoassays: Two automated immunoassays (Euroimmun and Abbott) and one rapid lateral flow immunoassay (NG Biotech). *Journal of Clinical Virology*, 2020, 129, pp.104511. 10.1016/j.jcv.2020.104511 . hal-03443223

HAL Id: hal-03443223

<https://univ-angers.hal.science/hal-03443223>

Submitted on 22 Aug 2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial 4.0 International License

1 **Title page**

2 **Title:** Assessment of SARS-CoV-2 serological tests for the diagnosis of COVID-19 through
3 the evaluation of three immunoassays: two automated immunoassays (Euroimmun and
4 Abbott) and one rapid lateral flow immunoassay (NG Biotech)

5 **Author names and affiliations:**

6 Thomas Nicol^a, Caroline Lefeuvre^{ab}, Oriane Serri^a, Adeline Pivert^{ab}, Françoise Joubaud^c
7 Alexandra Ducancelle^{ab}, Françoise Lunel-Fabiani^{ab*}, and Hélène Le Guillou-Guillemette^{ab*}

8 ^a*Virology Department, Angers University Hospital, Angers, France*

9 ^b*HIFIH Laboratory EA 3859, LUNAM, Angers, France*

10 ^c*Department of Biochemistry and Genetics, Angers University Hospital, Angers, France*

11 **These two authors equally participated to the study*

12 **Corresponding author:** Hélène Le Guillou-Guillemette

13 PharmD, PhD

14 Postal address: Virology Department, Angers University Hospital, HIFIH Laboratory EA
15 3859, LUNAM, 4, rue Larrey 49000 Angers, France

16 Telephone number: (+33) 02 41 35 47 09

17 **Word count of the abstract: 249/250**

18 **Word count of the text: 2493/2500**

19 **Declaration of interest:**

20 The authors declare that they have no conflict of interest.

21 **Funding:**

22 This research did not receive any specific grant from funding agencies. NG-Test® IgG-IgM
23 COVID-19 rapid test cassettes (NG Biotech Laboratoires) were kindly provided by the
24 manufacturer.

1 **Abstract**

2 *Background:* The emergence of new SARS-CoV-2 has promoted the development of new
3 serological tests that could be complementary to RT-PCR. Nevertheless, the assessment of
4 clinical performances of available tests is urgently required as their use has just been initiated
5 for diagnose.

6 *Objectives:* The aim of this study was to assess the performance of three immunoassays for
7 the detection of SARS-CoV-2 antibodies.

8 *Methods:* Two automated immunoassays (Abbott SARS-CoV-2 CLIA IgG and Euroimmun
9 Anti-SARS-CoV-2 ELISA IgG/IgA assays) and one lateral flow immunoassay (LFIA NG-
10 Test® IgG-IgM COVID-19) were tested. 293 specimens were analyzed from patients with a
11 positive RT-PCR response, from patients with symptoms consistent with COVID-19 but
12 exhibiting a negative response to the RT-PCR detection test, and from control group
13 specimens. Days since symptoms onset were collected from clinical information sheet
14 associated with respiratory tract samples.

15 *Results:* Overall sensitivity for IgG was equivalent (around 80%) for CLIA, ELISA and LFIA.
16 Sensitivity for IgG detection, >14 days after onset of symptoms, was 100.0% for all assays.
17 Overall specificity for IgG was greater for CLIA and LFIA (more than 98%) compared to
18 ELISA (95.8%). Specificity was significantly different between IgA ELISA (78.9%) and IgM
19 LFIA (95.8%) ($p < 0.05$). The best agreement was observed between CLIA and LFIA assays
20 (97%; $k = 0.936$).

21 *Conclusion:* Excellent sensitivity for IgG detection was obtained >14 days after onset of
22 symptoms for all immunoassays. Specificity was also excellent for IgG CLIA and IgG LFIA.
23 Our study shows that NG-Test® is reliable and accurate for routine use in clinical
24 laboratories.

25 **Keywords:** SARS-CoV-2; COVID-19; performance; automated immunoassays; lateral flow

26 immunoassay

27

28

29 **Background**

30 A new acute respiratory syndrome named coronavirus disease 2019 (COVID-19) has emerged
31 from the region of Wuhan in China in December 2019. This infection, widespread all over the
32 world, is caused by a novel *Sarbecovirus* designated severe acute respiratory syndrome
33 coronavirus 2 (SARS-CoV-2), associated with severe morbidity and mortality [1–3]. The
34 detection of viral RNA by real time reverse transcriptase-Polymerase chain reaction (RT-
35 PCR) in respiratory tract samples is considered as the gold standard method for screening and
36 diagnosis in the early phase of infection. However, sensitivity is variable depending on
37 sample types, suitable sampling technique, the anatomic site, time of infection and viral load
38 [4–6]. Chest computed tomography (CT) may be helpful for the diagnosis, complementary to
39 RT-PCR, but it remains unspecific [7]. Development of new serological tests [8,9], readily
40 available and easier to perform compared to requirements of molecular assays in laboratories
41 [10], could be helpful as a complementary diagnostic tool and to increase the sensitivity of
42 tests especially in patients with late complications i.e. severe pneumonia. Different assays
43 have recently been commercialized: automated tests (enzyme-linked immunosorbent assays
44 [ELISA] or chemiluminescence enzyme immunoassays [CLIA]) or rapid detection test
45 (lateral flow immunoassays, LFIA). LFIA seems to be very attractive for large seroprevalence
46 studies because these tests can be used easily as point of care tests or in the laboratory, with a
47 result in less than 15 minutes. Serological tests can be used for symptomatic individuals for
48 which RT-PCR testing was either not performed at the time of acute illness or for which
49 nasopharyngeal swab result was found to be negative, and also for epidemiological studies
50 (close contacts screening, screening of health care workers ...) [11,12]. However, the
51 relevance of serological tests is highly related to their clinical performance, hence antibody
52 (Ab) assays with good sensitivity and specificity are needed. Despite a growing number of

53 available assays, related clinical performances are still scarce [13–17] or unknown and
54 individual studies are usually inconclusive. Moreover, the quality and diagnostic performance
55 of rapid tests have already been questioned in Spain and United Kingdom [18,19].

56 **Objectives**

57 The aim of the study was to assess the clinical performance of CE marked assays available in
58 Europe to detect SARS-CoV-2 antibodies: two automated immunoassays (Euroimmun and
59 Abbott assays) targeting two different proteins and also one lateral flow immunoassay (NG
60 Biotech).

61

62

63 **Methods**

64 **Specimens**

65 This retrospective study included 293 residual sera from patients with RT-PCR confirmed
66 SARS-CoV-2 infection, patients with symptoms consistent with COVID-19 but with a
67 negative RT-PCR result (clinical diagnosis of pneumonia of unknown etiology), and control
68 individuals (presumed negative). These samples were collected in the virology laboratory of
69 Angers University Hospital, France. Serum samples (n=141) obtained from 82 patients
70 (median age: 67 years) with confirmed COVID-19 by RT-PCR, performed in our laboratory
71 [20], were tested. 57 serum specimens obtained from 52 patients (median age: 64 years) with
72 symptoms consistent with COVID-19, but with negative RT-PCR results were analyzed.
73 Information about days since symptoms onset was determined by clinical information sheet
74 associated with respiratory tract samples. 50 residual serum samples presumed negative
75 collected before the emergence of SARS-CoV-2, in March 2019 and stored at -80°C were
76 used as control specimens.

77 Then, 25 serum samples with a potential cross-reaction to the SARS-CoV-2 immunoassays
78 were investigated (Table 1). Samples from 10 pregnant women and 10 sera from patients with
79 positive rheumatoid factor (RF) were also tested.

80 The study was approved by the Institutional Board of the Angers University Hospital.

81

82 **Table 1.** Selected specimens potentially containing cross-reacting antibodies with SARS-
 83 CoV-2.

Pathogen potentially cross-reactive with SARS-CoV-2	Number of specimens
Seasonal coronaviruses (HKU1, NL63, 229E, OC43)	2
Influenza A virus	3
Respiratory Syncytial Virus	3
Rhinovirus	3
Parainfluenzae virus	1
Acute EBV infection (positive for EBV VCA IgM and EBV VCA IgG)	7
Acute CMV infection (positive for CMV IgM)	1
<i>M. pneumonia</i> infection	2
Acute Hepatitis A infection	1
Acute hepatitis E infection	2

84

85 **Serological assays**

86 ***ELISA assay***

87 The Euroimmun Anti-SARS-CoV-2 ELISA IgG and IgA assays (Euroimmun, Luebeck,
 88 Germany) were performed according to the manufacturer's guidelines on the DS2® system,
 89 an automated microplate technology (Dynex Technologies GmbH, Denkendorf, Germany).
 90 The microplate wells are coated with recombinant S1 structural protein and the assay detects
 91 anti-SARS-CoV-2 IgG and IgA against the viral spike protein (Sp).

92

93 ***CLIA assay***

94 The Abbott SARS-CoV-2 IgG (Abbott, Ireland) was performed according to the
 95 manufacturer's instructions on the automated Abbott ARCHITECT i2000SR Instrument

96 The assay is a CLIA for qualitative detection of IgG antibodies against the SARS-CoV-2
97 nucleoprotein (Np) in serum or plasma.

98

99 ***Lateral flow test***

100 NG-Test® IgG-IgM COVID-19 (NG Biotech Laboratoires, Guipry-Messac, France) is an
101 immune colloidal technique intended for the qualitative detection of IgG and IgM antibodies
102 against the SARS-CoV-2 nucleoprotein in serum or plasma. 10 µL of specimen, were added
103 onto the sample loading area followed by 2 drops of sample dilution solution. The results
104 were read and interpreted 15 min after testing.

105

106 **Statistical analysis**

107 All statistical analyses were performed using IBM® SPSS® 15.0 Statistics software
108 (Statistical Package for Social Sciences, IBM Corp., Chicago, IL). To assess the sensitivity
109 and specificity, we choose the RT-PCR method as gold standard. Time from onset symptoms
110 was used to determine sensitivity and specificity. Grey zone was considered positive for the
111 statistical analyses. A *p value* <0.05 was considered statistically significant. The Cohen's
112 Kappa value was determined for agreement between assays.

113

114 **Results**

115 Sensitivities and specificities obtained with three immunoassays are summarized in Table 2.
116 The sensitivity of IgG ELISA at ≤ 7 days of symptoms was 28.1%, at 8-14 days 72.4%, and
117 >14 days was 100.0%. The sensitivity of IgG CLIA at ≤ 7 days of symptoms was 46.9%, at 8-
118 14 days 69%, and was 100.0% >14 days. Sensitivity of IgG LFIA at ≤ 7 days of symptoms
119 was 31.3%, at 8-14 days 69.0%, and was 100.0% >14 days. Overall sensitivity for IgG was
120 equivalent (around 80%) for CLIA, ELISA and LFIA. Overall specificity for IgG was greater
121 than 98% for CLIA and LFIA compared to ELISA (95.8%). Comparison of the sensitivity of
122 IgA ELISA (59.4%) and IgM LFIA (43.8%), during the first seven days after onset of
123 symptoms, was not significant ($p > 0.05$). By contrast, specificity was significantly different
124 between IgA ELISA and IgM LFIA ($p < 0.05$).

125 Among the control samples and the group of patients with negative RT-PCR, 26 false
126 positives were observed with IgA ELISA (17.3%): seven specimens from the cross-reactivity
127 study; seven from pre-epidemic specimens (March 2019); two from pregnant women; four
128 from patients with RF; six from patients with negative SARS-CoV-2 RT-PCR and symptoms
129 of pneumonia/dyspnea without a chest CT argument for COVID-19 or seroconversion
130 (median time between symptom onset and sera: 9.5 days). Fewer false positives were
131 observed with IgM LFIA: three specimens from the cross-reactivity study; one from pre-
132 epidemic sera; three from patients with negative RT-PCR result and symptoms of
133 pneumonia/dyspnea without a chest CT argument (including two specimens from the same
134 patient). Five false positives were observed with IgG ELISA (Figure 1): two pre-pandemic
135 specimens, one sample from pregnant woman, one sample from a patient with RF and one
136 with negative RT-PCR result (negative result with other assays). Only one false positive result
137 was observed with IgG CLIA and corresponded to a pre-pandemic specimen (Figure 1).

138 Using IgG LFIA, three false positives were observed; two were from a patient (negative RT-
139 PCR) for whom the etiology of pneumonia was undetermined.

140 All patients with positive RT-PCR were positive for serological assays. Two discordant
141 results between serologic assays and molecular method were reported: two patients clinically
142 adjudicated as COVID-19 but with negative RT-PCR. These two samples were tested positive
143 for all immunoassays (Figure 1). To determine the specificity for IgG of the three assays, we
144 excluded two specimens positive for serological assays but negative for RT-PCR because the
145 symptoms were strongly compatible with the COVID-19 and RT-PCR was performed 17-24
146 days after symptom onset.

147 Among patients with RT-PCR confirmed SARS-CoV-2 infection, there were only two
148 individuals without COVID-19 symptoms, but with a notion of contact with infected patients.
149 Both SARS-CoV-2 RT-PCR and serological assays were positives for these patients.

150

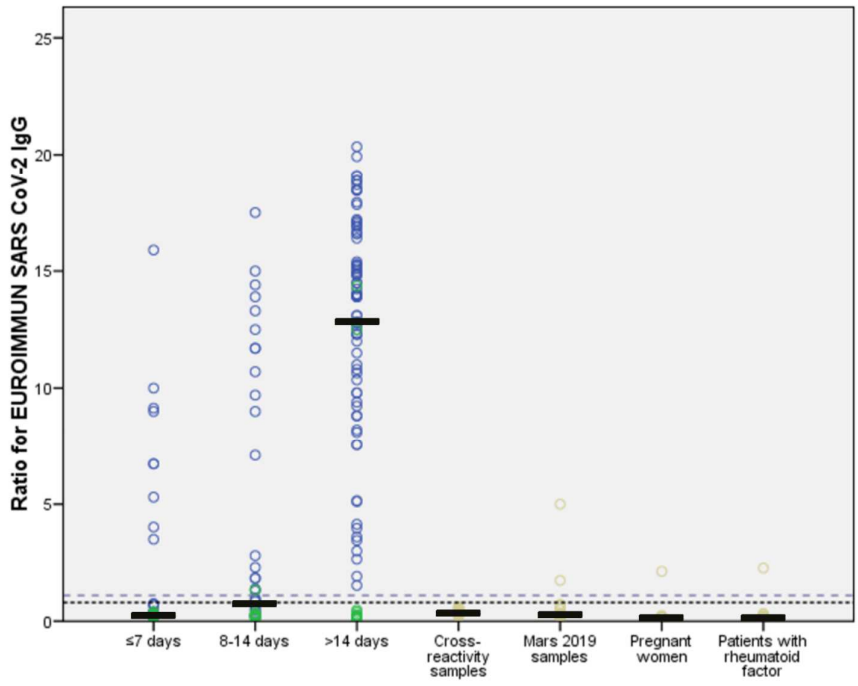
151 **Table 2.** Sensitivities of immunoassays for SARS-CoV-2 according to the onset of COVID-
 152 19 symptoms and specificities data. CI: confidence interval.

	Overall % (CI 95%)	Time from the symptom onset		
		0 to 7 days % (CI 95%)	8 to 14 days % (CI 95%)	15 or more days % (CI 95%)
ELISA assay				
IgG or IgA	Se: 87.4 (81.0-91.9%) Sp: 82.0 (75.1-87.3%)	Se: 59.4 (42.3-74.5%)	Se: 82.8 (65.5-92.4%)	Se: 100.0 (95.5-100.0%)
IgG	Se: 78.3 (70.9-84.3%) Sp: 96.7 (92.4-98.6%)	Se: 28.1 (42.3-74.5%)	Se: 72.4 (54.3-85.3%)	Se: 100.0 (95.5-100.0%)
IgA	Se: 86.7 (80.2-91.3%) Sp: 82.7 (75.8-87.9%)	Se: 59.4 (15.6-45.4%)	Se: 79.3 (61.6-90.2%)	Se: 100.0 (95.5-100.0%)
CLIA assay				
IgG	Se: 81.8 (74.7-87.3%) Sp: 99.3 (96.3-99.9%)	Se: 46.9 (30.9-63.6%)	Se: 69.0 (50.8-82.7%)	Se: 100.0 (95.5-100.0%)
Lateral flow immunoassay				
IgG or IgM	Se: 81.8 (74.7-87.3%) Sp: 95.3 (90.7-97.7%)	Se: 43.8 (28.2-60.7%)	Se: 72.4 (54.3-85.3%)	Se: 100.0 (95.5-100.0%)
IgG	Se: 78.3 (70.9-84.3%) Sp: 98.0 (94.3-99.3%)	Se: 31.3 (18.0-48.6%)	Se: 69.0 (50.8-82.7%)	Se: 100.0 (95.5-100.0%)
IgM	Se: 81.8 (74.7-87.3%) Sp: 95.3 (90.7-97.7%)	Se: 43.8 (28.2-60.7%)	Se: 72.4 (54.3-85.3%)	Se: 100.0 (95.5-100.0%)

153

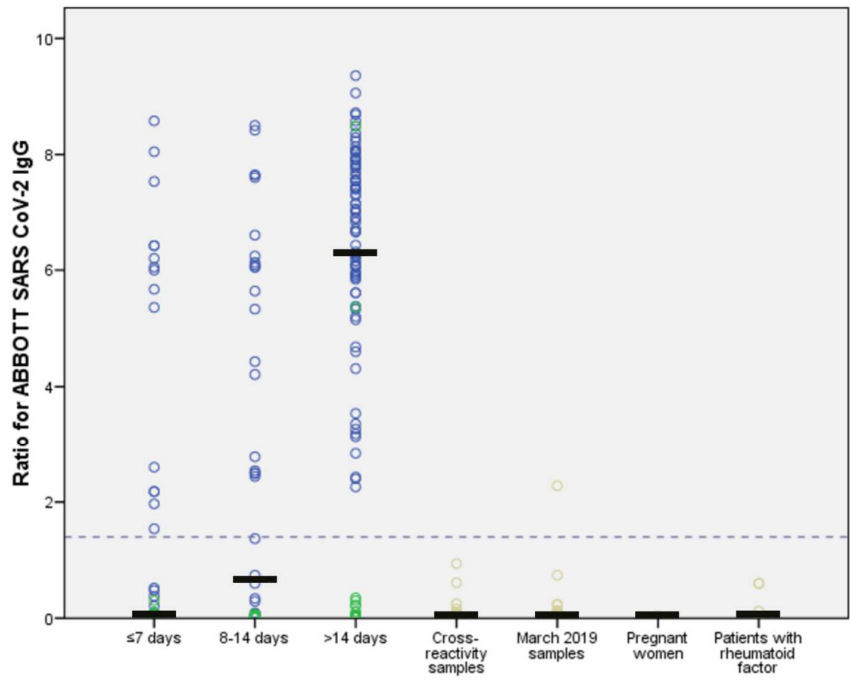
154

A $\frac{(+)}{(n)}$ $\frac{9}{56}$ $\frac{22}{44}$ $\frac{82}{98}$ $\frac{0}{25}$ $\frac{2}{50}$ $\frac{1}{10}$ $\frac{1}{10}$



155

B $\frac{(+)}{(n)}$ $\frac{15}{56}$ $\frac{20}{44}$ $\frac{82}{98}$ $\frac{0}{25}$ $\frac{1}{50}$ $\frac{0}{10}$ $\frac{0}{10}$



156

157 **Figure 1: Seropositivity of tested specimens with ELISA Euroimmun and CLIA Abbott**
158 **assays.** Seropositivity analysis in 95 presumed negatives control samples (cross-reactivity
159 samples, march 2019 samples, pregnant women samples, patients with RF samples), 57
160 samples from 52 patients with RT-PCR negative relative to days from symptom onset (≤ 7
161 days; 8-14 days; >14 days) and 141 samples from 82 patients with RT-PCR positive relative
162 to days from symptom onset. Blue circles correspond to sera from patients exhibiting a
163 positive RT-PCR result. Green circles correspond to sera from patients with negative RT-PCR
164 result. Ochre circles correspond to sera from individuals for whom RT-PCR detection has not
165 been performed. The black line represents the median of ratio. (+): number of seropositive
166 sera; (n): total number of specimens tested. **A)** Seropositivity with ELISA Euroimmun assay.
167 Dashed grey line represents cutoff for positivity (ratio ≥ 1.1). Dotted purple line corresponds
168 to cutoff for negativity (ratio < 0.8). **B)** Seropositivity with CLIA Abbott assay. Dashed grey
169 line represents cutoff for positivity (ratio ≥ 1.4).

170 Table 3 summarized overall agreement and agreement relative to the time of symptoms onset
171 between three immunoassays. Overall, excellent agreement was observed between the three
172 assays. The best agreement was observed between CLIA and LFIA (97%; Cohen kappa index
173 of 0.936). Even for the first week of symptoms onset, an excellent agreement was observed
174 between ELISA and LFIA assays (95%; $k=0.810$). However, poor agreement was observed
175 between ELISA and CLIA (89%; $k=0.687$). Overall agreement between IgG/IgA ELISA and
176 IgG/IgM LFIA was excellent (96%; $k=0.914$).

177

178 **Table 3.** Agreement between IgG serological assays.

		Euroimmun % (Kappa)	NG Biotech % (Kappa)
Overall	Abbott	96% (0.908)	97% (0.936)
	NG Biotech	96% (0.914)	
0 to 7 days	Abbott	89% (0.687)	91% (0.745)
	NG Biotech	95% (0.810)	
8 to 14 days	Abbott	95% (0.909)	98% (0.954)
	NG Biotech	93% (0.864)	
15 or more days	Abbott	100% (1.000)	99% (0.962)
	NG Biotech	99% (0.962)	

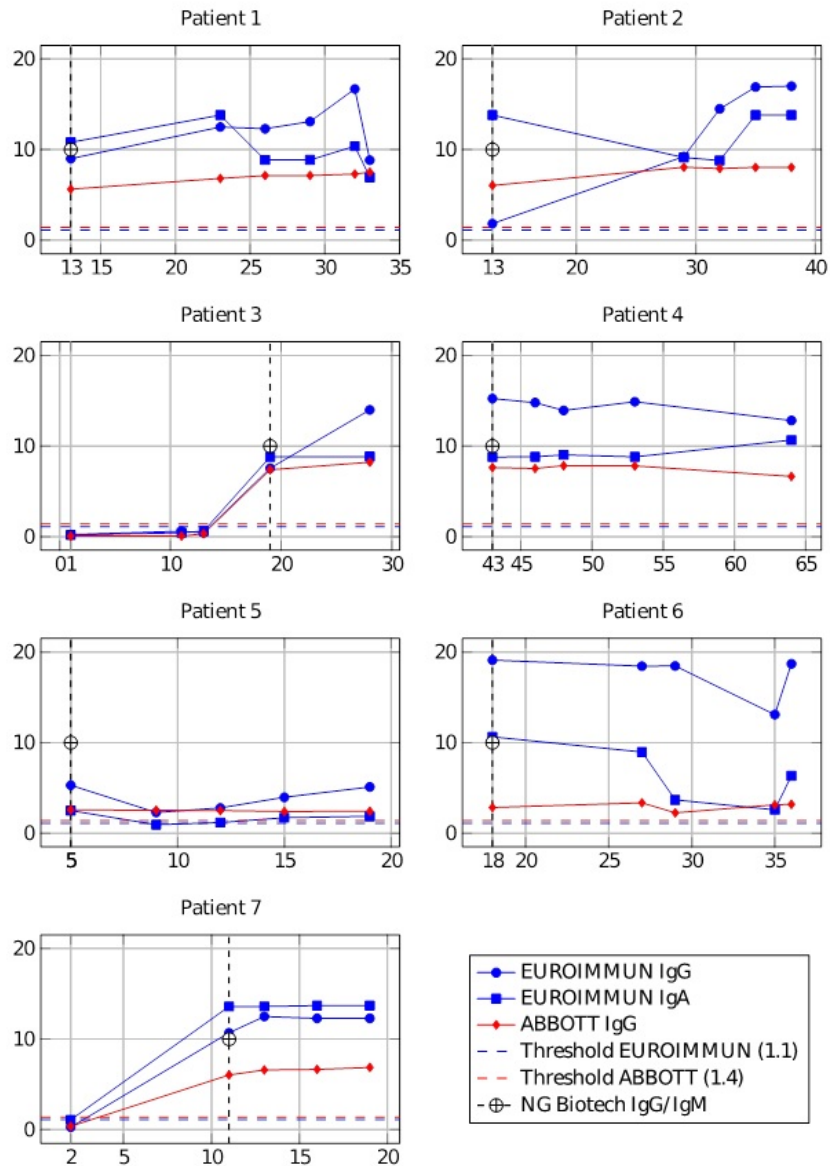
179

180 The IgA, IgM and IgG Ab kinetics were studied using specimens from seven patients
 181 (positive RT-PCR) with serial results and interesting kinetics (Figure 2). Then, five patients
 182 presented an earlier IgG seroconversion using CLIA compared to ELISA, the first week of
 183 symptom onset. Among these patients, we observed in three patients an IgM line with LFIA
 184 and IgA ELISA was positive for four patients.

185 Using LFIA, results were more easily interpretable for IgG line than for IgM line. IgM line
 186 was difficult for reading, notably for seven sera.

187

188



189

190 **Figure 2: Anti-SARS-CoV-2 antibodies seroconversion profiles for seven individuals.** X-
 191 axis: time from symptoms onset. Y-axis: interpretation ratio of two semi-quantitative
 192 immunoassays. Dotted black line represents the day of positivity of LFIA NG-Test® IgG-IgM
 193 COVID-19. The cutoff for positivity with ELISA Euroimmun assay is ≥ 1.1 (dotted blue line)
 194 and the cutoff for positivity with CLIA Abbott assay is ≥ 1.4 (dotted red line). Patients 1, 2, 4
 195 and 6 developed a prolonged immune response one month after symptoms onset and up 64
 196 days for patient 4. Patients 1, 2 and 7 had early seroconversion in the second week after
 197 symptoms onset and patient 5 had already seroconversion in the first week. Patient 3 had a

198 seroconversion in the third week after symptoms onset. Patient 5 produced fewer antibodies
199 compared to other patients and notably IgA production is close to the threshold of positivity.

200

201 **Discussion**

202 A strong clinical performance of assays in diagnosis and management of COVID-19 is
203 essential to quickly contain the COVID outbreak worldwide. Therefore, the development of
204 serological assays, routinely used in clinical laboratories to determine recent infection or
205 previous contact with viruses, is a good option complementary to RT-PCR method [21]. On
206 May, 2020, the French Health Authority (Haute Autorité de Santé) and Infectious Diseases
207 Society of America recommended that patients with symptoms consistent with COVID-19 but
208 having a positive result for SARS-CoV-2 by RT-PCR may be diagnosed by serological tests
209 [22,23]. Various immunoassays are available on the European market [24,25] and subjected to
210 European regulations with the mandatory CE marked for sales. Nevertheless, the European
211 Commission, in its April 2020 recommendations, allowed exceptionally the marketing of tests
212 that do not have the CE marked, in the interest of public health [22].

213 Here, we evaluated three different CE marked commercial immunoassays for detection of
214 SARS-CoV-2 antibodies in human serum and plasma. ELISA assay was performed on a semi-
215 automated microplate technology requiring high handling and with a limited capacity of tests
216 per day (90 tests per 4h). In contrast, CLIA assay is a fully automated random-access test and
217 that can perform over 4,000 tests per 24h. These two assays are used in clinical laboratories,
218 unlike LFIA, which can be used as a point of care test or in clinical laboratories and provides
219 a result within 15 min.

220 Performance of Euroimmun assay has been evaluated in some studies [13,14,26–29], showing
221 sensitivity for IgG between 85% and 95% >14 days after symptoms onset and specificity
222 between 95 and 100%. Few studies reported clinical performance of Abbott assay
223 [14,16,26,28]. Sensitivity for IgG was between 94% and 100% more than 14 days post
224 symptom onset and specificity between 99 and 100%. In our study, we showed a sensitivity

225 for IgG of 100% for CLIA Abbott and ELISA Euroimmun assays >14 days after symptoms
226 onset and an overall specificity for IgG of 78.3% and 81.8% with ELISA and CLIA
227 respectively. We carried out a large cross-reactivity study and more false positives results
228 were observed using ELISA than CLIA as previously described [14].

229 Recently, many CE marked LFIA became available. Two studies showed that sensitivity and
230 specificity were similar to those of Euroimmun assay [13,29]. However, to our knowledge,
231 only one study compared clinical performance between CLIA Abbott and LFIA [30] and no
232 study described diagnostic performance of NG-Test®. Here, we observed an excellent
233 agreement for IgG between CLIA and LFIA 15 days after onset symptoms ($k=0.810$), and an
234 excellent sensitivity and specificity for both assays. LFIA advantages are the ability to reach
235 larger population groups, when used in point-of-care, and to evaluate the herd immunity
236 without saturating the capacity of laboratories. However, these devices must be used with
237 caution. Trained staff or automated reader devices are needed for good interpretation of result.
238 Traceability of results may be at fault in case of use at the point-of-care and results may not
239 be reported to the health authorities for seroprevalence studies.

240 To evaluate sensitivity, some manufacturers or authors used the time from positive RT-PCR
241 rather than the time from symptom onset. However, there is a risk of misestimating sensitivity
242 as some patients presented late after the onset of symptoms with disease progression at time
243 of the first PCR testing. Then, sensitivity and specificity must be interpreted with caution. The
244 use of RT-PCR as gold standard may decrease the real number of patients infected by SARS-
245 CoV-2 due to false negative results.

246 In our study, we observed false positive results with IgA ELISA and few with IgM LFIA. No
247 false positive with IgM LFIA were observed with for RF specimens whereas interferences
248 were described with some other immunoassays [31]. Elslande et al pointed out that the ELISA
249 IgA should not be used for the screening of asymptomatic persons. It might be better not to

250 measure IgM or IgA since it may result in a significant number of false-positive results
251 without improving diagnostic performance. [29]. It would appear here that IgM detection with
252 the LFIA provides a gain in diagnostic performance.

253 Developed immunoassays target either the Sp or the Np of SARS-CoV-2 [32], involving
254 different immune Ab responses. However, related studies are controversial. Some studies
255 described that early antibody response was targeted against Np and then Sp inducing an
256 earlier positivity of the tests targeting Np [14,33]. By contrast, another study revealed that the
257 Sp-based ELISA was more sensitive than the Np-based one in the detection of IgM [34].
258 Here, we did not observe any significant difference between sensitivity of IgA ELISA and
259 IgM LFIA which detect two different targets. Positive results with serological tests do not
260 indicate the presence of neutralizing antibodies, i.e. possible protective immunity to SARS-
261 CoV-2 [35], but are only indicative of a contact with SARS-CoV-2

262 One of the strong points of our study is the use of large number of samples for cross
263 reactivities study with other pathogens and other causes of false positive results ie. pregnancy
264 and individuals with RF. Among symptomatic patients with positive RT-PCR, most of
265 patients were elderly with a potential risk of low immune response.

266 Some limitations of the study are the subjectivity in the perception of symptoms by patients,
267 in particular for elderly patient. Our study included few patients with asymptomatic infections
268 and positive RT-PCR because specific indications of tests in France were, at the time of the
269 study, mainly limited to patients who were symptomatic, due in particular to the shortage of
270 tests.

271 In conclusion, our study showed equivalent clinical performance for IgG of three
272 immunoassays (ELISA, CLIA and LFIA) >14 days after symptoms onset. The three assays
273 had, as expected, a poor sensitivity during first days of symptom onset. Therefore, serological

274 tests can be useful to confirm past COVID-19, to do epidemiologic studies 15 days after
275 symptoms onset [36] or to identify people who could return to the workplace, even if its use is
276 still widely discussed [37] . For asymptomatic patients with RT-PCR negative, a higher
277 threshold must be used [16]. A lower threshold (8-14 days) should be used for symptomatic
278 patients >7 days with negative RT-PCR and clinical presentation consistent with COVID-19.
279 Currently, it is not clear whether IgG antibodies are protective against reinfection [38].
280 Finally, even if the LFIA is reliable on serum or plasma, studies should be conducted to
281 evaluate the performance on fingerstick; a process commonly used for seroprevalence studies.

282 **Acknowledgments**

283 The authors thank the laboratory technicians who helped us.

284 The authors thank Thomas Le Guillou for proofreading the English manuscript.

285

286

287

288

289 **References**

- 290 [1] World Health Organization, Coronavirus disease 2019, (2020).
291 <https://www.who.int/emergencies/diseases/novel-coronavirus-2019> (accessed May 23,
292 2020).
- 293 [2] N. Zhu, D. Zhang, W. Wang, X. Li, B. Yang, J. Song, X. Zhao, B. Huang, W. Shi, R. Lu,
294 P. Niu, F. Zhan, X. Ma, D. Wang, W. Xu, G. Wu, G.F. Gao, W. Tan, China Novel
295 Coronavirus Investigating and Research Team, A Novel Coronavirus from Patients with
296 Pneumonia in China, 2019, *N Engl J Med.* 382 (2020) 727–733.
297 <https://doi.org/10.1056/NEJMoa2001017>.
- 298 [3] X. Yang, Y. Yu, J. Xu, H. Shu, J. Xia, H. Liu, Y. Wu, L. Zhang, Z. Yu, M. Fang, T. Yu,
299 Y. Wang, S. Pan, X. Zou, S. Yuan, Y. Shang, Clinical course and outcomes of critically
300 ill patients with SARS-CoV-2 pneumonia in Wuhan, China: a single-centered,
301 retrospective, observational study, *Lancet Respir Med.* 8 (2020) 475–481.
302 [https://doi.org/10.1016/S2213-2600\(20\)30079-5](https://doi.org/10.1016/S2213-2600(20)30079-5).
- 303 [4] S.K. Vashist, In Vitro Diagnostic Assays for COVID-19: Recent Advances and Emerging
304 Trends, *Diagnostics.* 10 (2020) 202. <https://doi.org/10.3390/diagnostics10040202>.
- 305 [5] M.J. Loeffelholz, Y.-W. Tang, Laboratory diagnosis of emerging human coronavirus
306 infections - the state of the art, *Emerg Microbes Infect.* 9 (2020) 747–756.
307 <https://doi.org/10.1080/22221751.2020.1745095>.
- 308 [6] Y. Yang, M. Yang, C. Shen, F. Wang, J. Yuan, J. Li, M. Zhang, Z. Wang, L. Xing, J.
309 Wei, L. Peng, G. Wong, H. Zheng, M. Liao, K. Feng, J. Li, Q. Yang, J. Zhao, Z. Zhang,
310 L. Liu, Y. Liu, Evaluating the accuracy of different respiratory specimens in the
311 laboratory diagnosis and monitoring the viral shedding of 2019-nCoV infections,
312 medRxiv, 2020. <https://doi.org/10.1101/2020.02.11.20021493>.

- 313 [7] X. Xie, Z. Zhong, W. Zhao, C. Zheng, F. Wang, J. Liu, Chest CT for Typical 2019-nCoV
314 Pneumonia: Relationship to Negative RT-PCR Testing, *Radiology*. (2020) 200343.
315 <https://doi.org/10.1148/radiol.2020200343>.
- 316 [8] A. Petherick, Developing antibody tests for SARS-CoV-2, *Lancet*. 395 (2020) 1101–
317 1102. [https://doi.org/10.1016/S0140-6736\(20\)30788-1](https://doi.org/10.1016/S0140-6736(20)30788-1).
- 318 [9] N.M.A. Okba, M.A. Müller, W. Li, C. Wang, C.H. GeurtsvanKessel, V.M. Corman,
319 M.M. Lamers, R.S. Sikkema, E. de Bruin, F.D. Chandler, Y. Yazdanpanah, Q. Le
320 Hingrat, D. Descamps, N. Houhou-Fidouh, C.B.E.M. Reusken, B.-J. Bosch, C. Drosten,
321 M.P.G. Koopmans, B.L. Haagmans, Severe Acute Respiratory Syndrome Coronavirus 2-
322 Specific Antibody Responses in Coronavirus Disease 2019 Patients, *Emerging Infect Dis*.
323 26 (2020). <https://doi.org/10.3201/eid2607.200841>.
- 324 [10] World Health Organization, Laboratory biosafety guidance related to the novel
325 coronavirus (2019-nCoV), (2020). [https://www.who.int/docs/default-](https://www.who.int/docs/default-source/coronaviruse/laboratory-biosafey-novel-coronavirus-version-1-1.pdf?sfvrsn=912a9847_2)
326 [source/coronaviruse/laboratory-biosafey-novel-coronavirus-version-1-](https://www.who.int/docs/default-source/coronaviruse/laboratory-biosafey-novel-coronavirus-version-1-1.pdf?sfvrsn=912a9847_2)
327 [1.pdf?sfvrsn=912a9847_2](https://www.who.int/docs/default-source/coronaviruse/laboratory-biosafey-novel-coronavirus-version-1-1.pdf?sfvrsn=912a9847_2) (accessed May 27, 2020).
- 328 [11] Y. Xu, M. Xiao, X. Liu, S. Xu, T. Du, J. Xu, Q. Yang, Y. Xu, Y. Han, T. Li, H. Zhu,
329 M. Wang, Significance of serology testing to assist timely diagnosis of SARS-CoV-2
330 infections: implication from a family cluster, *Emerg Microbes Infect*. 9 (2020) 924–927.
331 <https://doi.org/10.1080/22221751.2020.1752610>.
- 332 [12] Y.-W. Tang, J.E. Schmitz, D.H. Persing, C.W. Stratton, The Laboratory Diagnosis of
333 COVID-19 Infection: Current Issues and Challenges, *J Clin Microbiol*. (2020).
334 <https://doi.org/10.1128/JCM.00512-20>.
- 335 [13] I. Montesinos, D. Gruson, B. Kabamba, H. Dahma, S. Van den Wijngaert, S. Reza, V.
336 Carbone, O. Vandenberg, B. Gulbis, F. Wolff, H. Rodriguez-Villalobos, Evaluation of
337 two automated and three rapid lateral flow immunoassays for the detection of anti-SARS-

338 CoV-2 antibodies, *J Clin Virol.* 128 (2020) 104413.
339 <https://doi.org/10.1016/j.jcv.2020.104413>.

340 [14] M.S. Tang, K.G. Hock, N.M. Logsdon, J.E. Hayes, A.M. Gronowski, N.W. Anderson,
341 C.W. Farnsworth, Clinical Performance of Two SARS-CoV-2 Serologic Assays, *Clin*
342 *Chem.* (2020). <https://doi.org/10.1093/clinchem/hvaa120>.

343 [15] Y. Pan, X. Li, G. Yang, J. Fan, Y. Tang, J. Zhao, X. Long, S. Guo, Z. Zhao, Y. Liu, H.
344 Hu, H. Xue, Y. Li, Serological immunochromatographic approach in diagnosis with
345 SARS-CoV-2 infected COVID-19 patients, *J Infect.* (2020).
346 <https://doi.org/10.1016/j.jinf.2020.03.051>.

347 [16] A. Bryan, G. Pepper, M.H. Wener, S.L. Fink, C. Morishima, A. Chaudhary, K.R.
348 Jerome, P.C. Mathias, A.L. Greninger, Performance Characteristics of the Abbott
349 Architect SARS-CoV-2 IgG Assay and Seroprevalence in Boise, Idaho, *J Clin Microbiol.*
350 (2020). <https://doi.org/10.1128/JCM.00941-20>.

351 [17] Z. Zainol Rashid, S.N. Othman, M.N. Abdul Samat, U.K. Ali, K.K. Wong, Diagnostic
352 performance of COVID-19 serology assays, *Malays J Pathol.* 42 (2020) 13–21.

353 [18] E. Sevillano, Covid-19 in Spain: Unreliability of new tests delays effort to slow
354 coronavirus spread in Spain | Society | EL PAÍS in English, (2020).
355 [https://english.elpais.com/society/2020-03-27/unreliability-of-new-tests-delays-effort-to-](https://english.elpais.com/society/2020-03-27/unreliability-of-new-tests-delays-effort-to-slow-coronavirus-spread-in-spain.html)
356 [slow-coronavirus-spread-in-spain.html](https://english.elpais.com/society/2020-03-27/unreliability-of-new-tests-delays-effort-to-slow-coronavirus-spread-in-spain.html) (accessed June 9, 2020).

357 [19] D. Brennan, U.K. says millions of coronavirus test kits bought from China are
358 unreliable for most patients, *Newsweek.* (2020). [https://www.newsweek.com/uk-says-](https://www.newsweek.com/uk-says-millions-coronavirus-test-kits-bought-china-unreliable-most-patients-1496506)
359 [millions-coronavirus-test-kits-bought-china-unreliable-most-patients-1496506](https://www.newsweek.com/uk-says-millions-coronavirus-test-kits-bought-china-unreliable-most-patients-1496506) (accessed
360 June 9, 2020).

361 [20] F.-X. Lescure, L. Bouadma, D. Nguyen, M. Parisey, P.-H. Wicky, S. Behillil, A.
362 Gaymard, M. Bouscambert-Duchamp, F. Donati, Q. Le Hingrat, V. Enouf, N. Houhou-

363 Fidouh, M. Valette, A. Mailles, J.-C. Lucet, F. Mentre, X. Duval, D. Descamps, D.
364 Malvy, J.-F. Timsit, B. Lina, S. van-der-Werf, Y. Yazdanpanah, Clinical and virological
365 data of the first cases of COVID-19 in Europe: a case series, *The Lancet Infectious*
366 *Diseases*. 20 (2020) 697–706. [https://doi.org/10.1016/S1473-3099\(20\)30200-0](https://doi.org/10.1016/S1473-3099(20)30200-0).

367 [21] Z. Yongchen, H. Shen, X. Wang, X. Shi, Y. Li, J. Yan, Y. Chen, B. Gu, Different
368 longitudinal patterns of nucleic acid and serology testing results based on disease severity
369 of COVID-19 patients, *Emerg Microbes Infect.* 9 (2020) 833–836.
370 <https://doi.org/10.1080/22221751.2020.1756699>.

371 [22] Haute Autorité de Santé, Place des tests sérologiques dans la stratégie de prise en
372 charge de la maladie COVID-19, (2020) 37.

373 [23] Infectious Diseases Society of America, IDSA COVID-19 antibody primer. May 4,
374 2020. [https://www.idsociety.org/globalassets/idsa/public-health/covid-19/idsa-covid-19-](https://www.idsociety.org/globalassets/idsa/public-health/covid-19/idsa-covid-19-antibody-testing-primer.pdf)
375 [antibody-testing-primer.pdf](https://www.idsociety.org/globalassets/idsa/public-health/covid-19/idsa-covid-19-antibody-testing-primer.pdf) (accessed June 10, 2020).

376 [24] Ministère des Solidarités et de la Santé, Plateforme COVID-19, (2020). [https://covid-](https://covid-19.sante.gouv.fr/tests)
377 [19.sante.gouv.fr/tests](https://covid-19.sante.gouv.fr/tests) (accessed May 25, 2020).

378 [25] SARS-CoV-2 diagnostic pipeline. (2020). <https://www.finddx.org/testing-matters/>
379 (accessed June 9, 2020).

380 [26] N. Kohmer, S. Westhaus, C. Rühl, S. Ciesek, H.F. Rabenau, Brief clinical evaluation
381 of six high-throughput SARS-CoV-2 IgG antibody assays, *J Clin Virol.* 129 (2020)
382 104480. <https://doi.org/10.1016/j.jcv.2020.104480>.

383 [27] N. Kohmer, S. Westhaus, C. Rühl, S. Ciesek, H.F. Rabenau, Clinical performance of
384 different SARS-CoV-2 IgG antibody tests, *J Med Virol.* (2020).
385 <https://doi.org/10.1002/jmv.26145>.

- 386 [28] E.S. Theel, J. Harring, H. Hilgart, D. Granger, Performance Characteristics of Four
387 High-Throughput Immunoassays for Detection of IgG Antibodies against SARS-CoV-2, J
388 Clin Microbiol. (2020). <https://doi.org/10.1128/JCM.01243-20>.
- 389 [29] J. Van Elslande, E. Houben, M. Depypere, A. Brackenier, S. Desmet, E. André, M.
390 Van Ranst, K. Lagrou, P. Vermeersch, Diagnostic performance of 7 rapid IgG/IgM
391 antibody tests and the Euroimmun IgA/IgG ELISA in COVID-19 patients, Clin Microbiol
392 Infect. (2020). <https://doi.org/10.1016/j.cmi.2020.05.023>.
- 393 [30] S. Dellière, M. Salmona, M. Minier, A. Gabassi, A. Alanio, J. Le Goff, C. Delaugerre,
394 M.-L. Chaix, Evaluation of COVID-19 IgG/IgM Rapid Test from Orient Gene Biotech, J
395 Clin Microbiol. (2020) JCM.01233-20. <https://doi.org/10.1128/JCM.01233-20>.
- 396 [31] Q. Wang, Q. Du, B. Guo, D. Mu, X. Lu, Q. Ma, Y. Guo, L. Fang, B. Zhang, G. Zhang,
397 X. Guo, A Method To Prevent SARS-CoV-2 IgM False Positives in Gold
398 Immunochromatography and Enzyme-Linked Immunosorbent Assays, J Clin Microbiol.
399 58 (2020). <https://doi.org/10.1128/JCM.00375-20>.
- 400 [32] N. Sethuraman, S.S. Jeremiah, A. Ryo, Interpreting Diagnostic Tests for SARS-CoV-
401 2, JAMA. (2020). <https://doi.org/10.1001/jama.2020.8259>.
- 402 [33] M.Z. Tay, C.M. Poh, L. Rénia, P.A. MacAry, L.F.P. Ng, The trinity of COVID-19:
403 immunity, inflammation and intervention, Nat Rev Immunol. 20 (2020) 363–374.
404 <https://doi.org/10.1038/s41577-020-0311-8>.
- 405 [34] W. Liu, L. Liu, G. Kou, Y. Zheng, Y. Ding, W. Ni, Q. Wang, L. Tan, W. Wu, S. Tang,
406 Z. Xiong, S. Zheng, Evaluation of Nucleocapsid and Spike Protein-Based Enzyme-Linked
407 Immunosorbent Assays for Detecting Antibodies against SARS-CoV-2, J Clin Microbiol.
408 58 (2020). <https://doi.org/10.1128/JCM.00461-20>.

- 409 [35] N. Kohmer, S. Westhaus, C. Rühl, S. Ciesek, H.F. Rabenau, Clinical performance of
410 SARS-CoV-2 IgG antibody tests and potential protective immunity, *Microbiology*, 2020.
411 <https://doi.org/10.1101/2020.05.08.085506>.
- 412 [36] A.K. Winter, S.T. Hegde, The important role of serology for COVID-19 control,
413 *Lancet Infect Dis.* (2020). [https://doi.org/10.1016/S1473-3099\(20\)30322-4](https://doi.org/10.1016/S1473-3099(20)30322-4).
- 414 [37] M.C. Weinstein, K.A. Freedberg, E.P. Hyle, A.D. Paltiel, Waiting for Certainty on
415 Covid-19 Antibody Tests — At What Cost?, *N Engl J Med.* (2020)p2017739.
416 <https://doi.org/10.1056/NEJMp2017739>.
- 417 [38] J.G. Melgaço, T. Azamor, A.P.D. Ano Bom, Protective immunity after COVID-19 has
418 been questioned: What can we do without SARS-CoV-2-IgG detection?, *Cell. Immunol.*
419 353 (2020) 104114. <https://doi.org/10.1016/j.cellimm.2020.104114>.
- 420