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Original article

Live benthic foraminiferal faunas along a bathymetrical transect (140–4800 m) in the Bay of Biscay (NE Atlantic)

Les foraminifères benthiques vivants le long d'un transect bathymétrique (140–4800 m) dans le Golfe de Gascogne (Atlantique NE)

Meryem Mojtahid^{a,b,c,*}, Clémentine Griveaud^{b,c}, Christophe Fontanier^{b,c}, Pierre Anschutz^d, Fransiscus Josephus Jorissen^{b,c}

^a National Oceanography Centre, Southampton, Waterfront campus, European way, Southampton SO14 3ZH, United Kingdom

^b Laboratoire d'étude des bio-indicateurs actuels et fossiles (BIAF), UFR Sciences, université d'Angers, 2, boulevard Lavoisier, 49045 Angers cedex, France

^c Laboratoire d'étude des bio-indicateurs Marins (LEBIM), Ker Chalon, Port Joinville, 85350 Ile d'Yeu, France

^d UMR-CNRS 5805 EPOC (Environnements et paléo-environnements océaniques), site de Talence, université de Bordeaux 1, avenue des Facultés, 33405 Talence cedex, France

Abstract

In a 10-stations bathymetrical transect in the Bay of Biscay, we observed important changes in the density, composition and microhabitats of live foraminiferal faunas from the outer continental shelf to the abyssal plain. Four zones are recognised: (1) at the upper continental shelf (140 m water depth), foraminiferal densities are very high and the superficial sediment is occupied by *Bolivina subaenariensis* and *Valvulineria bradyana*. *Globobulimina* spp., *Chilostomella oolina* and *Nonion fabum* dominate the infaunal niches, which are positioned close to the sediment–water interface due to a strong compaction of the vertical succession of redox zones. (2) At the upper continental slope stations (300–1000 m), foraminiferal densities are high and the superficial sediments are dominated by *Uvigerina mediterranea/peregrina*. Deeper in the sediment, intermediate infaunal niches are occupied by *Melonis barleeanus*. Due to a deeper oxygen penetration, the deep infaunal taxa *Globobulimina* spp. and *C. oolina* live at a considerable depth in the sediment. (3) At the mid and lower slope stations (1000–2000 m) in the superficial sediment *Cibicides kullenbergi* and *Hoeglundina elegans* progressively replace *U. mediterranea*. *U. peregrina* is still a dominant taxon, reflecting its preference for a somewhat intermediate organic flux level. Deep infaunal taxa become increasingly rare. (4) At the lower slope and abyssal plane stations (deeper than 2000 m), faunal densities are very low and the fauna is composed exclusively by shallow infaunal species, such as *Nuttallides umboniferus* and *Melonis pompilioides*. The foraminiferal data together with the pore water data in the sediment give evidence of the presence of a trophic gradient from very eutrophic settings at the upper continental shelf towards oligotrophic settings at the abyssal area.

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Keywords: Benthic foraminifera; Bathymetric transect; Trophic levels; Redox conditions; Microhabitat; Bay of Biscay

Résumé

Le long d'un transect bathymétrique de 10 stations dans le Golfe de Gascogne, nous observons des changements importants de la densité, la composition et des microhabitats des faunes de foraminifères benthiques, du plateau continental vers la plaine abyssale. Nous avons pu distinguer quatre zones successives : (1) la partie externe du plateau continental (140 m de profondeur) est caractérisée par des densités très élevées de foraminifères et le sédiment de surface est habité par *Bolivina subaenariensis* et *Valvulineria bradyana*. Les niches plus profondes, se trouvant près de l'interface eau–sédiment en raison de la compaction de la succession verticale des zones redox, sont dominées par *Globobulimina* spp., *Chilostomella oolina* et *Nonion fabum*. (2) Dans les environnements du haut de la pente continentale (300–1000 m), les densités sont élevées et le sédiment superficiel est occupé par *Uvigerina mediterranea/peregrina*. Dans le sédiment, *Melonis barleeanus* occupe les niches intermédiaires. En raison d'une pénétration d'oxygène maximale, les taxons endopéliques *Globobulimina* spp. et *C. oolina* vivent profondément dans le sédiment.

* Corresponding author.

E-mail address: Meryem.Mojtahid@noc.soton.ac.uk (M. Mojtahid).

(3) Dans les environnements du bas de pente (1000–2000 m), les espèces superficielles *Cibicoides kullenbergi* et *Hoeglundina elegans* remplacent progressivement *U. mediterranea*. *U. peregrina* reste dominante, ce qui reflète sa préférence pour des flux organiques intermédiaires. Les espèces endobiontes profondes se raréfient. (4) Dans les environnements du bas de pente et de la plaine abyssale (plus de 2000 m), les densités sont faibles et la faune est composée exclusivement d'espèces superficielles adaptées aux faibles apports organiques, comme *Nuttallides umboniferus* et *Melonis pompilioides*. Les faunes de foraminifères, ainsi que la pénétration d'oxygène dans le sédiment témoignent d'un gradient trophique allant de conditions très eutrophiques dans la partie externe du plateau continental à des conditions oligotrophiques dans les environnements de bas de pente et de la plaine abyssale.

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Mots clés : Foraminifères benthiques ; Transect bathymétrique ; Conditions redox ; Niveaux trophiques ; Microhabitat ; Golfe de Gascogne

1. Introduction

Benthic foraminifera are among the most abundant protists in marine ecosystems. In deep-sea environments, they may represent more than 50% of the total biomass (Gooday et al., 1992). Due to their short life cycle, high diversity and specific ecological requirements, foraminifera adapt rapidly to the physicochemical conditions of their environment (Murray, 2006). They can respond quickly to the input of organic material (e.g., Gooday, 1988; Silva et al., 1996; Kitazato et al., 2000) by increasing their density (e.g., Kitazato et al., 2000; Fontanier et al., 2003), changing the faunal composition or modifying their microhabitat (Linke and Lutze, 1993; Jorissen et al., 1995; Ohga and Kitazato, 1997). For a long time, certain species have been associated to specific bathymetric ranges (e.g., Phleger, 1951; Parker, 1958; Pujos-Lamy, 1973; Pflum and Frerichs, 1976; Lutze, 1980). Nowadays, it seems accepted that the correlation of these associations with water depth is not very robust (e.g., see review in Jorissen et al., 2007) and that the distributional patterns of foraminiferal species are more related to the organic matter flux to the sea floor and the oxygen content of bottom and pore waters (e.g., Lutze, 1980; Lutze and Coulbourn, 1984; Altenbach and Sarthein, 1989; Loubere, 1991, 1994; Jorissen et al., 1995; Fariduddin and Loubere, 1997; Gooday, 2003; Jorissen et al., 2007). However, other factors (e.g., sedimentary processes, the nature of the substrate, or biological processes, such as competition and predation) also influence the spatial distributional patterns of benthic foraminifera, and may become predominant in specific settings. In open ocean deep-sea environments, the organic matter flux that arrives at the sea floor mostly originates from surface primary production. Because of partial remineralisation in the water column, the amount of organic matter that reaches the sediment and the proportion of labile organic matter decrease with increasing water depth (e.g., Suess, 1980; Berger and Wefer, 1990; Herguera, 1992). In general, the flux of metabolisable organic matter is therefore inversely proportional to water depth.

In a number of previous studies, Fontanier et al. (2002, 2003, 2006) and Langezaal et al. (2006) described the composition, densities and microhabitats of live benthic foraminiferal faunas in the Bay of Biscay along a 5-station bathymetric transect from 140 to 2000 m depth. Fontanier et al. (2002) showed decreasing faunal densities with increasing water depth and decreasing estimated organic matter flux, and a bathymetrical succession of species, some having a restricted bathymetric range. Certain species adapt to dysoxic/anoxic conditions within the sediments,

and show varying microhabitats from one station to another, in function of the precise depth of successive redox boundaries. Fontanier et al. (2003, 2006) described the seasonal and interannual variability of benthic foraminiferal faunas at two stations situated at 550 and 1000 m water depths. The seasonal variability of the foraminiferal faunas at a 140 m deep outer shelf station has been described by Langezaal et al. (2006). These authors tried to relate temporal faunal changes to estimations of surface water primary production, which were based on satellite images. This method can perhaps yield acceptable estimates in open ocean settings, where the organic matter flux has an important vertical component. On the contrary, in more marginal environments, lateral sediment transport mechanisms (e.g., by nepheloid layers, or by gravitational processes) tend to be important, and a large part of the organic matter arriving at the sea floor may result from lateral supplies. Schmidt et al. (2009) argue that this is also the case for the Bay of Biscay, especially for sites that are situated in the vicinity of submarine canyons. In such cases, flux equations do no longer give realistic results. Therefore, in this study we use the redox conditions, not only to assess the remineralisation pathways in the sediment, but also as a proxy of the trophic conditions of the benthic ecosystem. At the sediment–water interface, oxygen migrates from the bottom waters into the sediment by molecular diffusion and macrofaunal bio-irrigation. In the superficial sediment, (aerobic) microorganisms preferentially use oxygen for the degradation of organic matter, thereby obtaining the energy necessary for their life functions (growth, reproduction, etc.). Oxygen becomes therefore depleted at a depth of a few millimetres to tens of centimetres below the water/sediment interface, depending on the intensity of organic matter degradation. In eutrophic ecosystems, all oxygen is consumed before all labile organic carbon has been remineralised. In the absence of external disturbances (e.g., sediment burrowing), which can transport oxygen to deeper sediment layers, organic matter degradation then continues by anaerobic pathways, using other electron acceptors, such as nitrates, metal oxides (iron and manganese) and sulfates. In general, the oxygen penetration depth in the sediment is therefore inversely proportional to the amount of organic carbon arriving at the sea floor.

In this paper, we will reconsider the live benthic foraminiferal faunas along the sample transect initially described by Fontanier et al. (2002). We will present new data for five additional sampling stations. These new data will allow us to describe in more detail the influence of the environmental parameters on the density, specific composition and vertical distribution of benthic foraminiferal faunas along the transect. Two of the five new

stations are positioned at the upper slope (at 320 m and 800 m water depth), one at mid-slope (1600 m), whereas two stations are located at the lower slope (2400 m) and on the abyssal plain (4800 m). The main objectives of this paper are:

- to present and discuss variation of the foraminiferal density with increasing water depth and thus, probably with a diminishing organic flux, using a combination of existing data for a five station transect in the Bay of Biscay and new data from five complementary stations (320 m, 800 m, 1600 m, 2400 m and 4800 m water depth);
- to better define the ecological characteristics of the main foraminiferal species, and to explain the microhabitat changes

in response to the trophic conditions at the sediment–water interface.

2. Sampling area, material and methods

2.1. Regional setting

The Bay of Biscay (Fig. 1) is an oceanic embayment, partially closed to the south by the Spanish coast and to the east by the French coast. Water masses in the Bay of Biscay originate from the North Atlantic drift. Surface waters arrive from the north, along the Irish continental shelf, and leave at Cape Finisterre. The surface water patterns are strongly constrained

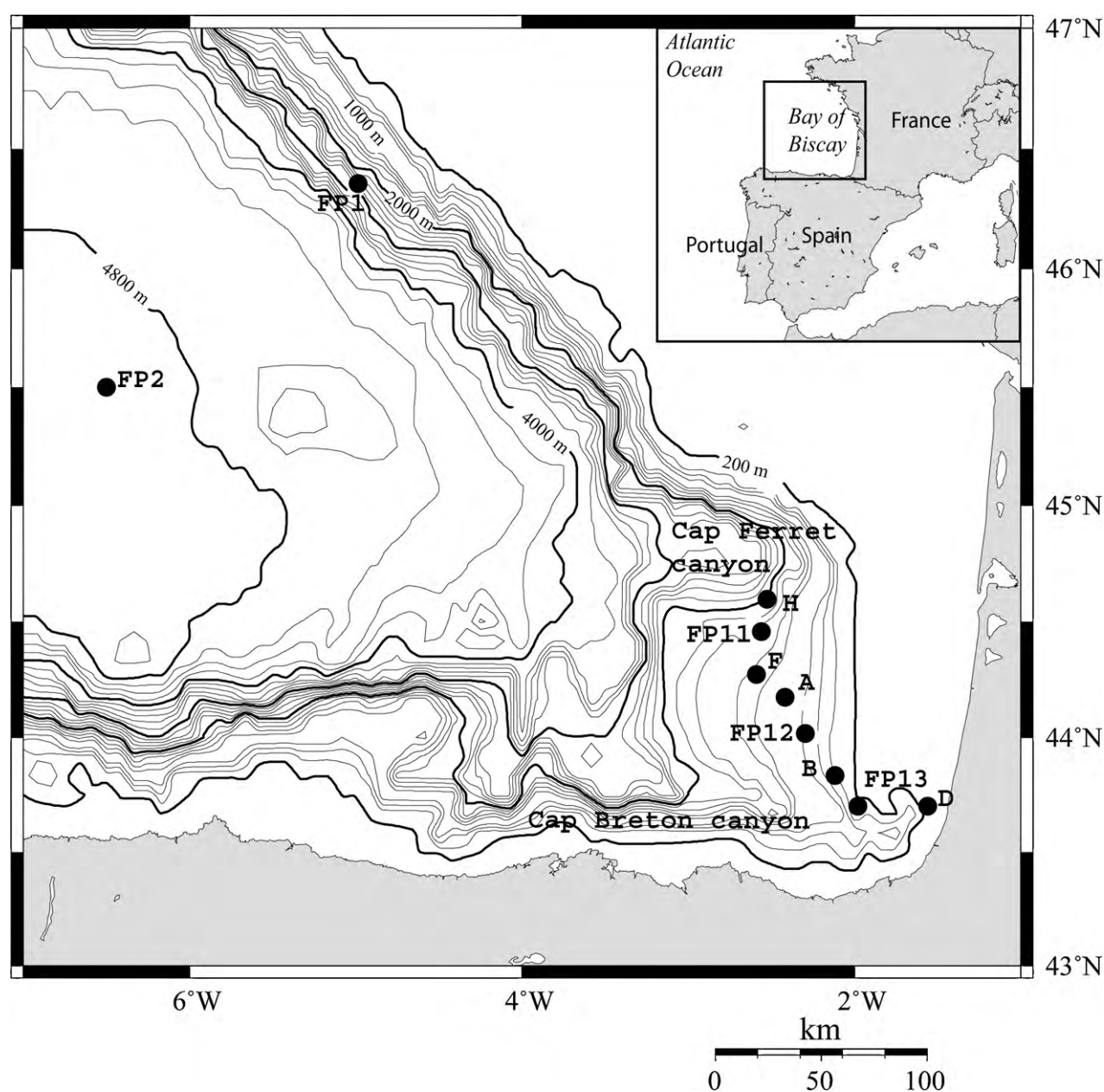


Fig. 1. Study area, bathymetry and geographical position of the 10 stations.
Site d'étude, bathymétrie et localisation des 10 stations.

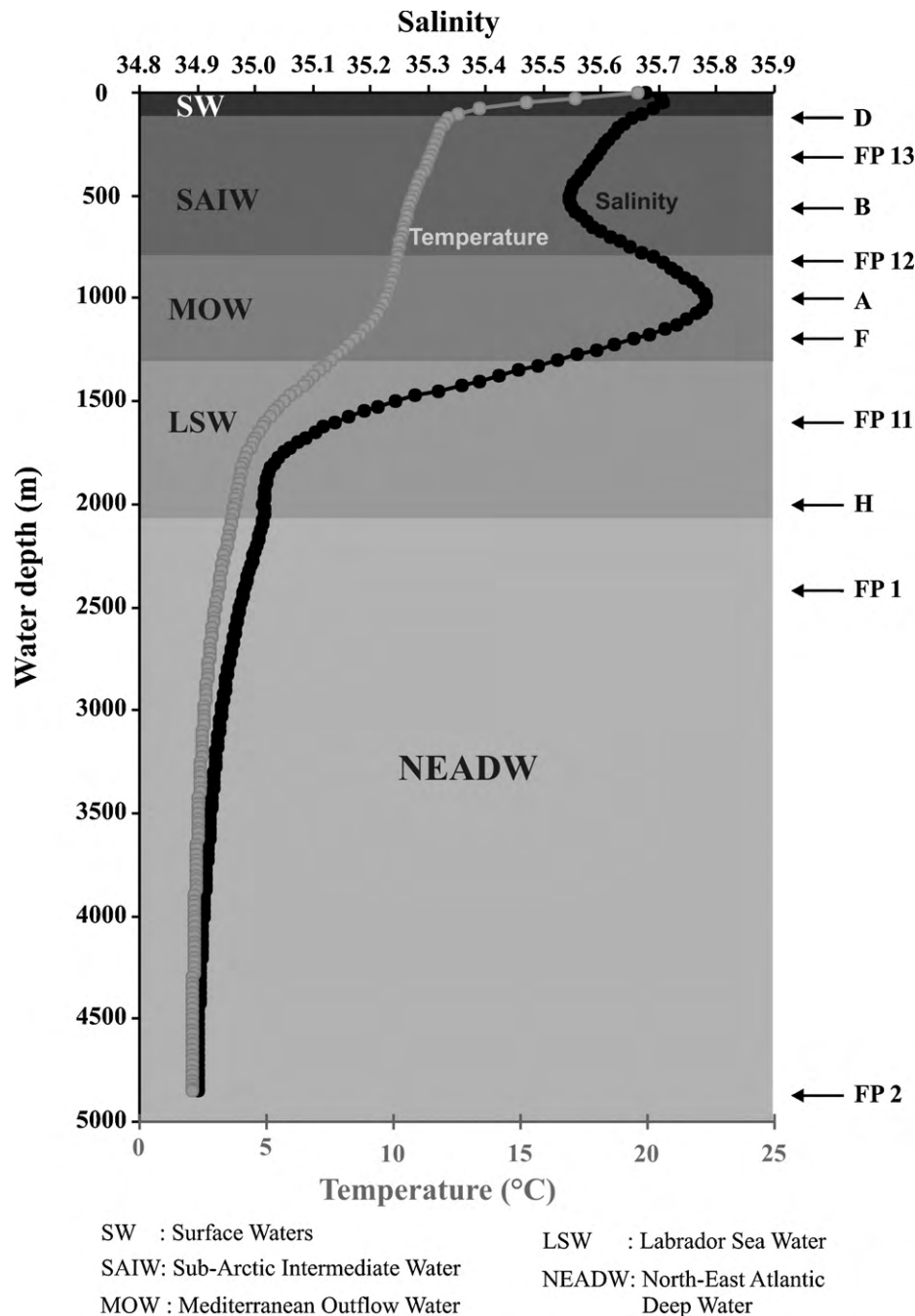


Fig. 2. Salinity and temperature CTD profiles at station FP2 (Van Aken, unpublished data) and the localisation of water masses (After Van Aken, 2000a, 2000b, 2001).

Profils CTD de salinité et température à la station FP2 (Van Aken, données non-publiées) et localisation des masses d'eau (d'après Van Aken, 2000a, 2000b, 2001).

by seasonal variations of the thermocline and the mixed layer (Tréguer et al., 1979). Below the surface waters (~150 m), the Sub-Arctic Intermediate Waters (SAIW) are present down to approximately 800 m depth (Fig. 2) presenting a sub-surface salinity minimum. Between 800 m and 1300 m, a branch of the Mediterranean Outflow Water (MOW) is present. This branch is slightly less oxygenated (3.8 mL.L^{-1} , Le Floch, 1968) and presents a higher salinity (35.8) in comparison to the surrounding waters (Fig. 2). Below the Mediterranean waters and until about 2000 m depth, the Labrador Sea Waters (LSW) are found

(Fig. 2). Deeper, the cold and low salinity North-East Atlantic Deep Waters (NEADW) are present (Fig. 2).

The continental slope bordering the French shelf deepens gradually, and is interrupted by two large canyons (Cap Ferret and Cap Breton Canyons) (Fig. 1). Vertical fluxes represent generally the main sedimentary component in open slope environments (Heussner et al., 1999), but lateral advection may dominate sedimentary processes in and close to the canyons (Heussner et al., 1999; Schmidt et al., 2009). Surface primary production values vary from 145 to 170 $\text{gC/m}^2/\text{yr}$ in the Bay

of Biscay (Laborde et al., 1999). A clear primary production maximum is observed in spring; primary production values are minimal in summer and winter (Tréguer et al., 1979; Laborde et al., 1999).

2.2. Sediment sampling

Stations FP1 and FP2 were sampled in August 2003, station FP11 in May 2004, and stations FP12 and FP13 in August 2005 (Table 1). Stations D, B and A were sampled several times between 1997 and 2001, station F was sampled in January 1998 and station H in October 1998. In the study of Fontanier et al. (2002), only one core per station was investigated. In the present paper, we use also the additional cores analysed in later studies (Fontanier et al., 2003, 2006; Langezaal et al., 2006) at stations D, B and A (Table 1). For these three stations, averaged values are given to describe the faunas of each station. Only cores where the entire ten first centimetres of the sediment were analysed have been considered. Sediments were collected between 140 and 4800 m water depth using classical Barnett multicorers (Barnett et al., 1984). The corer used during the FORAMPROX1 cruise (stations FP1 and FP2) contained 8 Plexiglas tubes of 6.3 cm internal diameter (core surface: $\sim 31 \text{ cm}^2$) whereas the corer used during all other cruises took 8 cores with an internal diameter of 9.5 cm (core surface: $\sim 71 \text{ cm}^2$). This sampling method is the most efficient to obtain an undisturbed sediment and water–sediment interface. The ten stations have been selected to compose a SE–NW bathymetrical/trophic transect, ranging from the outer shelf to bathyal open slope environments in the Bay of Biscay.

2.3. Geochemistry

At stations FP11, FP12 and FP13, overlying bottom waters were collected immediately after core recovery in order to

measure the dissolved oxygen concentrations using the Winkler titration method (Strickland and Parsons, 1972). Profiles of pore water oxygen were obtained onboard with a cathode-type mini-electrode (Revsbeck, 1983; Helder and Bakker, 1985; Revsbeck and Jørgensen, 1986). At stations FP1 and FP2, no measurements were performed of bottom water oxygen concentrations; they were estimated using World Ocean Atlas 2005 data (<http://www.nodc.noaa.gov/>). For these cores, the depth of the zero oxygen level was estimated on the basis of the manganese and nitrate profiles. At station FP2, we tentatively placed the zero oxygen level at the intersection of the nitrate and Mn^{2+} profiles. At station FP1, Mn^{2+} profiles were not determined below 9 cm, and our estimated depth of the zero oxygen level is uniquely based on the nitrate profile. See Fontanier et al. (2002, 2003, 2006) and Langezaal et al. (2006) for a description of the other geochemical methods used. Temperature and salinity data listed in Table 1 are the results of a compilation of Conductivity–Temperature–Depth (CTD) measurements performed during three years in the Bay of Biscay (Van Aken, 2000a, 2000b, 2001).

2.4. Foraminifera

For faunal analysis, entire cores were sliced horizontally for each station; every half cm until 4 cm depth, and every cm between 4 and 10 cm (see Fontanier et al., 2002, 2003, 2006; Langezaal et al., 2006 for the sampling strategy used at stations D, B, A, H and F). Sediments were stored in 500 cm^3 bottles, which were filled with 95% ethanol containing 1.0 to 1.2 g/L Rose Bengal stain. Rose Bengal staining is commonly used to identify live foraminifera (Walton, 1952). All samples were gently shaken for several minutes in order to obtain a homogeneous mixture. In the laboratory, they were sieved through 63 and 150 μm mesh screens, and the sieve residues were stored in 95% ethanol. Stained foraminifera belonging to the greater than

Table 1

Water depth, number of studied cores, core surface, sampling period, geographical position, bottom water oxygen (BWO), oxygen penetration depth (OPD) in the sediment (see text for full explanation), bottom water salinity and temperature. Stations in bold (FP13, FP12, FP11, FP1 and FP2) are the five newly studied stations. The other stations (D, B, A, F and H) have been studied by Fontanier et al. (2002, 2003, 2006) and Langezaal et al. (2006). At stations FP1 and FP2, BWO values are estimated using World Ocean Atlas 2005 data and OPD values are estimated on the basis of nitrate and Mn^{2+} profiles (see methods section Geochemistry).

Profondeur d'eau, nombre des carottes étudiées, surface des carottes, période d'échantillonnage, position géographique, oxygène des eaux de fond (BWO), pénétration d'oxygène dans le sédiment (OPD), salinité et température des eaux de fond. Les stations en gras (FP13, FP12, FP11, FP1 et FP2) sont les cinq nouvelles stations. Les autres stations (D, B, A, F et H) ont été étudiées par Fontanier et al. (2002, 2003, 2006) et Langezaal et al. (2006). Aux stations FP1 et FP2, l'oxygénation des eaux de fond a été estimée à partir des données du « World Ocean Atlas 2005 » et les valeurs de pénétration d'oxygène dans le sédiment ont été estimées sur la base des profils de nitrates et de manganèse (voir méthodes section Géochimie).

Station	Number of studied cores	Core surface (cm^2)	Sampling period	Depth (m)	Lat.	Long.	BWO $\mu\text{mol/L}$	OPD (cm)	Bottom Sal. (‰)	Bottom Temp. ($^{\circ}\text{C}$)
D	13	70.85	Oct-97 to Apr-01	140	43°42'N	1°34'W	223	0.54	35.6	12.1
FP13	1	70.85	Aug-05	320	43°42'N	1°59'W	218	2	35.6	11.4
B	11	70.85	Oct-97 to Apr-01	550	43°50'N	2°03'W	210	1.9	35.6	10.8
FP12	1	70.85	Aug-05	800	44°00'N	2°15'W	175	2.5	35.7	10.2
A	12	70.85	Oct-97 to Apr-01	1000	44°10'N	2°20'W	195	2.8	35.8	9.7
F	1	70.85	Jan-98	1200	44°17'N	2°45'W	210	6.3	35.6	8.4
FP11	2	70.85	May-04	1600	44°27'N	2°39'W	250	4.5	35.3	5.8
H	1	70.85	Oct-98	2000	44°32'N	2°37'W	261	6.23	35.1	4.1
FP1	6	31.16	Aug-03	2400	46°20'N	5°00'W	268	9	35	3.3
FP2	6	31.16	Aug-03	4800	45°30'N	6°30'W	268	8	34.9	2.5

150 μm fraction were sorted from wet samples, and stored in Chapman slides. For some of the stations, the 63–150 μm fraction was studied for the uppermost levels. The data for this small size fraction are not considered in this paper. A potential problem of the Rose Bengal staining technique is the fact that Rose Bengal may stain the protoplasm of dead foraminifera, which may be relatively well preserved for a considerable period of time under the anoxic conditions that generally prevail deeper in the sediment (Bernhard, 1988; Corliss and Emerson, 1990; Bernhard et al., 2006). Consequently, a strict application of the staining criteria is most times easy in superficial samples, but may become more critical in the deeper levels. In all cases, we applied our staining criteria (all chambers except the last one stained brightly pink) very strictly, and compared doubtful individuals with perfectly stained individuals of the same species found in superficial sediment layers. Non-transparent agglutinated and miliolid taxa were broken on many occasions for inspection of the interior of the test. We tried to identify most of live foraminifera at species level.

All data presented by Fontanier et al. (2002, 2003, 2006) and Langezaal et al. (2006) are based on the study of single cores with a surface area of $\sim 71 \text{ cm}^2$. When data on more than one core are available (either replicate cores or cores at different sampling periods), average values (with standard errors) are used. New data are based on a single $\sim 71 \text{ cm}^2$ core for stations FP12 and FP13 and on two $\sim 71 \text{ cm}^2$ cores for station FP11. Finally, for stations FP1 and FP2, 6 cores with a surface area of $\sim 31 \text{ cm}^2$ were studied. In order to be able to compare the cores, all densities have been standardized for a sediment surface of 50 cm^2 . For all vertical profiles, densities are standardized for 50 cm^3 sediment volume (see appendices for raw data).

In order to describe the vertical distribution of the total faunas or individual taxa, we use the Average Living Depth (ALD_x , Jorissen et al., 1995), which allows a rapid description of the microhabitat patterns. The ALD_x is calculated with the following formula:

$$\text{ALD}_x = \sum_{i=0,x} (n_i \times D_i) / N$$

- x = lower boundary of deepest sample
- n_i = number of individuals in interval i
- D_i = midpoint of sample interval i
- N = total number of individuals for all levels.

For all stations, ALD_{10} was calculated for the whole fauna as well as for individual taxa, on the basis of the numbers of stained individuals found in the successive sediment slices. Isolated individuals separated from the main population by more than 1 cm of “sterile” sediment (without live individuals of the studied taxa) were not considered in the calculations of the ALD_{10} . We suppose that such isolated individuals have been transported downward (outside their normal microhabitat) by bioturbation, or correspond to dead organisms that have been counted erroneously. In the data sheets, the latter individuals are indicated in italics. After the first classification with four main microhabitats proposed by Corliss and Chen (1988), it was argued that only species living on elevated substrates can be considered as

“epifaunal” (Buzas et al., 1993). Therefore, in the soft bottom communities described in this study, we recognize only three different groups: shallow infaunal, intermediate infaunal and deep infaunal species.

3. Results

Fontanier et al. (2002, 2003, 2006) and Langezaal et al. (2006) have presented results of foraminiferal densities, species composition and microhabitats for 5 stations (D, B, A, F and H) located on the continental slope of the Bay of Biscay, at water depths ranging from 140 to 2000 m. In this paper, we investigated live faunas from five additional stations: FP13 (320 m), FP12 (800 m), FP11 (1600 m), FP1 (2400 m) and FP2 (4800 m).

3.1. Benthic geochemical processes

For the 10 stations of the bathymetric transect, data for bottom water oxygen concentration, inferred oxygen penetration depth in the sediment, bottom water salinity and temperature are summarized in Table 1. The pore water profiles of the 5 new stations are presented in Fig. 3.

Bottom water oxygen concentration varies ranging from 175 $\mu\text{mol/L}$ at station FP12 (bathed by Mediterranean Outflow Waters; Fig. 2) to 268 $\mu\text{mol/L}$ at stations FP1 and FP2 (bathed by North-East Atlantic Deep Waters; Fig. 2). The depth of oxygen penetration into the sediment is minimal ($\sim 0.5 \text{ cm}$) at the outer shelf station D (140 m), and progressively increases with water depth, until maximum values of about 9 cm are reached at 2400 and 4800 m depth, respectively (Fig. 4[A]).

3.2. Live foraminiferal densities and composition

Fig. 4(B, C) shows densities of live benthic foraminifera plotted against the water depth and the oxygen penetration depth in the sediment. For stations that were sampled several times (D, B and A), during different sampling campaigns, the arithmetic average of foraminiferal densities, and standard deviations are represented on the graph. Decreasing densities of live faunas are observed with increasing water depth and increasing oxygen penetration depth. Densities range from 1430 ind./ 50 cm^2 at the shallowest station D (140 m) to 28 ind./ 50 cm^2 at the deepest station FP2 (Table 2).

Live faunas from the previously studied stations are described in detail in Fontanier et al. (2002, 2003, 2006) and Langezaal et al. (2006). Here, we will only describe the five newly added stations (Fig. 5):

- at station **FP13 (320 m)**, a total of 485 live individuals are found in the studied core ($\sim 71 \text{ cm}^2$), corresponding to a density of 342 ind./ 50 cm^2 (Table 2; Appendix A). The maximal density, of 141 ind./ 50 cm^3 , is found in the second half centimetre of the sediment (Fig. 5). Faunal density decreases progressively until 10 cm depth where only $\sim 1 \text{ ind./}50 \text{ cm}^3$ is found. Major taxa ($\geq 5.0\%$ of the total fauna) are *Bigenenerina nodosaria* (19.8%), *Globobulimina* spp. (17.5%), *Melonis barleeanus* (13.6%) and *Chilostomella oolina* (7.6%);

Table 2

Absolute (N) and relative densities (%) of the main live species (present with $\geq 5\%$ in at least one station) at the ten stations of the bathymetric transect in the first 10 cm of the sediment. An averaged value is given for stations D, B and A that were sampled at several periods. The data for station FP11 are totals, based on the study of 2 cores with a $\sim 71 \text{ cm}^2$ surface. Both for stations FP1 and FP2, 6 cores with a $\sim 31 \text{ cm}^2$ were studied. The table presents the total number of live foraminifera found in these 6 cores.

Densités absolues (N) et relatives (%) des espèces vivantes majoritaires (présentes à $\geq 5\%$ dans au moins une station) dans les 10 premiers cm du sédiment au niveau des 10 stations du transect bathymétrique. Une valeur moyenne est donnée pour les stations D, B et A ayant été échantillonnées à plusieurs périodes. Les données pour la station FP11 sont des totaux basés sur l'étude de deux carottes de $\sim 71 \text{ cm}^2$ de surface. Pour chacune des stations FP1 et FP2, 6 carottes de $\sim 31 \text{ cm}^2$ de surface ont été étudiées. Le nombre total des foraminifères vivants trouvés dans ces 6 carottes est donné dans le tableau.

Stations	D	FP13	B	FP12	A	F	FP11	H	FP1	FP2
Depth (m)	140	320	550	800	20550	1200	1600	4800	2400	4800
Sediment surface (cm ²)	71	71	71	71	71	71	142	71	186	186
N	%	N	%	N	%	N	%	N	%	N
<i>Perforates</i>	651	32.1	1	0.2	7	1.6	1	3.1	19	11.3
<i>Bolivina subaenariensis</i>									1.7	6.0
<i>Bulimina alazanensis</i>								1	0.6	
<i>Bulimina inflata</i>	223	11.0	37	7.6	7		81	25.4	3	1.8
<i>Chilostomella oolina</i>				1.2			11	3.4		
<i>Cibicides kullenbergi</i>						8	6.6			
<i>Cibicides robertsonianus</i>						3	2.5	5.0	2.2	4.8
<i>Gavelinopsis translucens</i>	6	0.3	85	17.5	37	6.4	2	0.8	3	
<i>Globobulimina</i> spp.						5.0	1	0.9	5	
<i>Gyroldina orbicularis</i>	6	0.3	1	0.2	2	0.3	2	1.4	18	1.0
<i>Hoeglundina elegans</i>						1.8	12	5.3	10.1	5.4
<i>Melonis berleanus</i>						9.7	7	21.9	32.4	
<i>Melonis pompilioides</i>						3.0	6	1.9		7
<i>Nonion fabum</i>	366	18.0								
<i>Nuttallides umboniferus</i>			2	0.4	1	0.7	8	1.9	3	1.8
<i>Uvigerina elongatastrata</i>			14	2.9	31	5.3				17.3
<i>Uvigerina mediterranea</i>			18	3.7	180	31.0	67	47.9		
<i>Uvigerina peregrina</i>	43	2.1	2	0.4	54	9.3	2	1.4	25.1	9
<i>Valvulineria bradyana</i>	210	10.4					24	7.5	5.4	
<i>Porcellaneous</i>										
<i>Pyrgo elongata</i>	15	0.7	9	1.9			16	5.0		8.7
<i>Quinqueloculina</i> sp. 1			0	0.1	1	0.1	7	0.6	1	
<i>Fossilizing Agglutinates</i>										
<i>Bigenerina nodosaria</i>			96	19.8	7	1.2	7	2.2	6	3.6
<i>Eggerella bradyi</i>					3	0.6				29
<i>Non-fossilizing Agglutinates</i>										
<i>Ammobaculites agglutinans</i>					16	3.5			11	6.5
<i>Clavulina cylindrica</i>	105	5.2	18	3.7	18	3.1				3
<i>Cribrostomoides</i> spp.	2	0.1	28	4.8	10	4.8	13	4.1	2.2	3
<i>Lagenammina pseudodifflugiformis</i>					9	2.0	4	3.3	9	5.4
<i>Psammosphaera</i> spp.			4	0.8	1	0.1	9	5.7		3
<i>Reophax dentaliniformis</i>			2	0.4	1	0.2	3	0.9	9	5.4
<i>Total densities</i>	2031	485	581	140	440	122	319	179	168	104
<i>Total standardized density/50 cm²</i>	1430	342	409	99	310	86	112	126	45	28

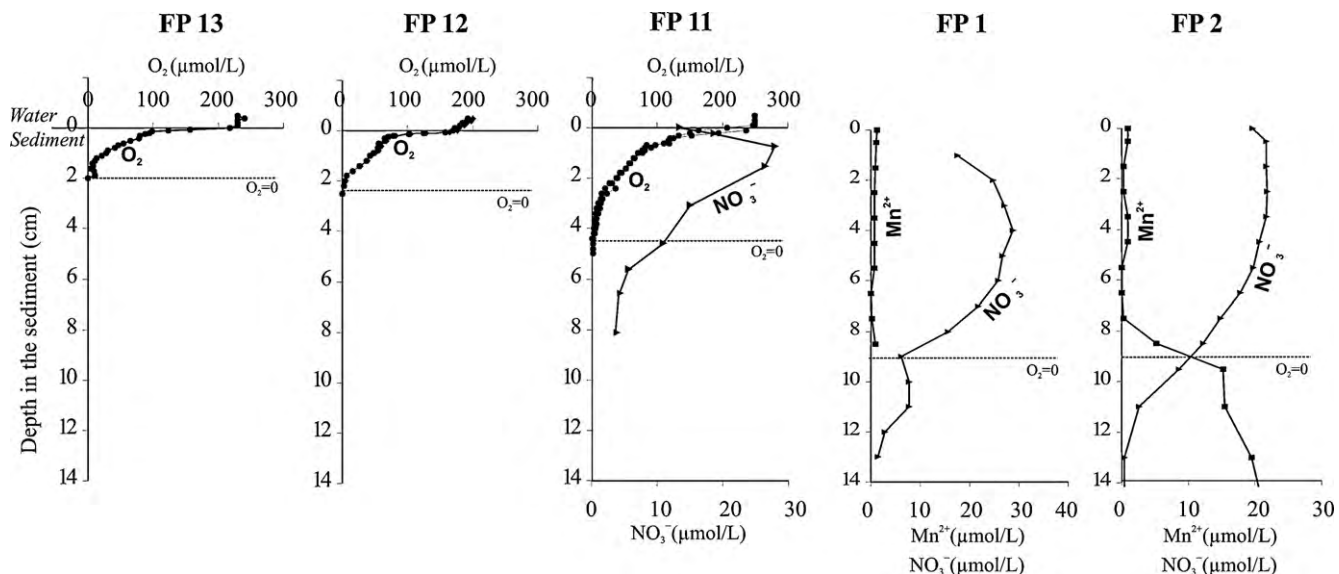


Fig. 3. Pore water data at the five new stations FP13 (320 m), FP12 (800 m), FP11 (1600 m), FP1 (2400 m) and FP2 (4800 m). At stations FP1 and FP2, the oxygen penetration limit is estimated on the basis of nitrate and Mn^{2+} profiles (see methods section Geochemistry).

Données géochimiques du sédiment au niveau des cinq nouvelles stations FP13 (320 m), FP12 (800 m), FP11 (1600 m), FP1 (2400 m) et FP2 (4800 m). Aux stations FP1 et FP2, l'estimation de la limite de pénétration d'oxygène dans le sédiment est basée sur les profils de nitrates et de manganèse (voir méthodes, section Géochimie).

- at station **FP12 (800 m)**, 140 live benthic foraminifera were found in the studied core ($\sim 71 \text{ cm}^2$), corresponding to 99 ind./50 cm^2 (Table 2; Appendix B). A maximum density is found in the first half centimetre of the sediment ($\sim 112 \text{ ind./50 cm}^3$) (Fig. 5). The density decreases rapidly towards deeper sediment levels. Below 3 cm, live fauna is nearly absent. Major taxa ($\geq 5.0\%$ of the total fauna) are *Uvigerina mediterranea* (47.9%), *Cribr stomoides* spp. (7.1%), *Psammosphaera* spp. (6.4%) and *Melonis barleeanus* (5.0%);
- at station **FP11 (1600 m)**, 2 cores (surface area $\sim 71 \text{ cm}^2$) were studied, which contained a total of 319 live foraminifera, corresponding to 112 ind./50 cm^2 (Table 2; Appendix C). The maximal density (99 ind./50 cm^3) is found in the first half centimetre (Fig. 5). The density decreases very rapidly down core, attaining less than 10 ind./50 cm^3 in the 1–1.5 cm level. Faunal density remains below this value until the bottom of the core. The live foraminiferal assemblage is dominated by *Cibicides kullenbergi* (25.4%), *Hoeglundina elegans* (21.9%), *Uvigerina peregrina* (7.5%), *Gyroidina orbicularis* (5.3%), *Gavelinopsis translucens* (5.0%) and *Pyrgo elongata* (5.0%);
- at station **FP1 (2400 m)**, a total density of 168 individuals was obtained for the 6 studied cores (each with a $\sim 31 \text{ cm}^2$ surface area), corresponding to 45 ind./50 cm^2 (Table 2; Appendix D). Foraminifera are strongly concentrated in the first half cm of the sediment (65 ind./50 cm^3) (Fig. 5). About 10 ind./50 cm^3 are found in the 0.5–1 cm and 1–1.5 cm intervals, whereas only some specimens are found between 1.5 and 2.5 cm depth. No live specimens have been found deeper in the core. This station is characterized by the presence of 45 different taxa, no species being strongly predominant. On Fig. 5, only species that occur with more than 5.0% are represented. Major species

are *Bulimina alazanensis* (11.3%), *Bulimina inflata* (6.0%), *Uvigerina peregrina* (5.4%), *Hoeglundina elegans* (5.4%), and three agglutinated species: *Ammobaculites agglutinans* (6.5%), *Cribr stomoides* spp. (5.4%) and *Reophax dentaliniformis* (5.4%);

- station **FP2 (4800 m)** is characterized by the lowest foraminiferal densities in the sampled transect: a total of 104 individuals was found in the 6 studied cores (each with a $\sim 31 \text{ cm}^2$ surface area), corresponding to 28 ind./50 cm^2 (Table 2; Appendix E). A maximum of 32 ind./50 cm^3 is present in the first half centimetre of the sediment (Fig. 5). Faunas are dominated by *Eggerella bradyi* (27.9%), *Nuttallides umboniferus* (17.3%), *Quinqueloculina* sp. 1 (8.7%) and *Melonis pompilioides* (6.7%).

3.3. Microhabitats of live foraminifera

For the newly analysed stations, the vertical distribution of the total assemblage is shown in Fig. 5. ALD_{10} values for total faunas and for the dominant species are presented in Fig. 6 and Table 3:

- at station FP13 (320 m), the ALD_{10} of the entire fauna is 2.3 cm. *Bigenerina nodosaria* is a shallow infaunal taxon ($ALD_{10} = 0.7 \text{ cm}$), *Melonis barleeanus* occupies intermediate infaunal niches ($ALD_{10} = 2.0 \text{ cm}$) and *Chilostomella oolina* and *Globobulimina* spp. are deep infaunal species ($ALD_{10} = 4.8$ and 4.6 cm , respectively);
- at station FP12 (800 m), the ALD_{10} of the entire fauna is 0.8 cm. *Uvigerina mediterranea*, *Cribr stomoides* spp. and *Psammosphaera* spp. are shallow infau-

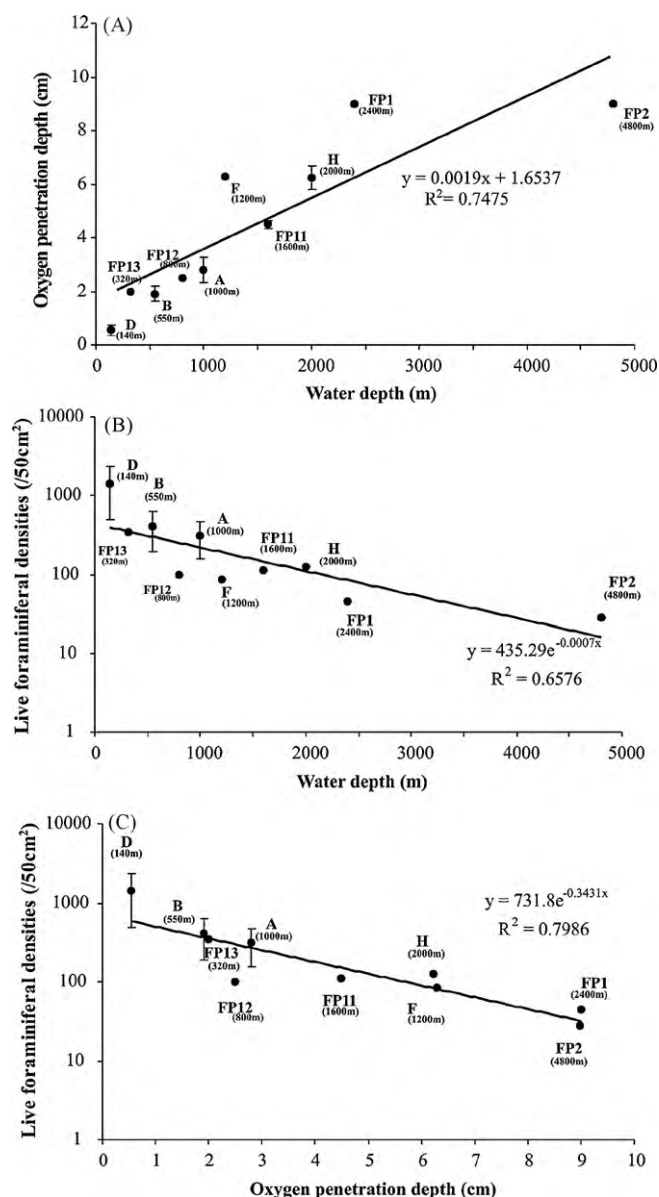


Fig. 4. **A.** Relationship between the oxygen penetration depth and the water depth along the 10-stations bathymetric transect. At stations FP1 and FP2, oxygen penetration depth into sediment was estimated following nitrate and Mn^{2+} sediment concentrations. See methods, section Geochemistry for full explanation. The error bars represent the variability in oxygen profiles at the stations sampled and/or measured several times. **B.** Live foraminiferal densities (standardized for a 50 cm² sediment surface, for a 10-cm-long core) as a function of water depth; **C.** Live foraminiferal densities (standardized for a 50 cm² sediment surface, for a 10-cm-long core) as a function of oxygen penetration depth into the sediment. The error bars in Fig. 4 (B, C) represent the variability in foraminiferal densities at the stations sampled and/or measured several times. **A.** Profondeur de pénétration d'oxygène dans le sédiment en fonction de la profondeur d'eau dans les 10 stations du transect bathymétrique. Aux stations FP1 et FP2, la pénétration d'oxygène dans le sédiment est estimée à partir des mesures de nitrates et de manganèse (voir méthodes, section Géochimie). Les bars d'erreurs représentent la variabilité des mesures d'oxygène aux stations échantillonnées et/ou mesurées plusieurs fois; **B.** Densités des foraminifères vivants (standardisées pour une surface de 50 cm², pour une carotte de 10 cm de long) en fonction de la profondeur d'eau; **C.** Densités des foraminifères vivants (standardisées pour une surface de 50 cm², pour une carotte de 10 cm de long) en fonction de la pénétration d'oxygène dans le sédiment. Les bars d'erreur sur la Fig. 4 (B, C) représentent la variabilité des densités des foraminifères au niveau des stations échantillonnées et/ou mesurées plusieurs fois.

nal species ($ALD_{10} = 0.7$, 0.7 , and 0.3 cm, respectively) while *M. barleeanus* inhabits intermediate microhabitats ($ALD_{10} = 1.1$ cm);

- at station FP11 (1600 m), the ALD_{10} of the entire fauna is 1.3 cm. *Hoeglundina elegans* and *Gavelinopsis translucens* have the most superficial microhabitat ($ALD_{10} = 0.3$ cm), followed by *Cibicidoides kullenbergi* ($ALD_{10} = 0.5$ cm), *Pyrgo elongata* and *Uvigerina peregrina* ($ALD_{10} = 0.6$ cm). *Gyroidina orbicularis* ($ALD_{10} = 1.5$ cm) occupies a somewhat deeper microhabitat;
- at station FP1 (2400 m), the ALD_{10} of the entire fauna is only 0.5 cm. *H. elegans* and *Bulimina inflata* are only present in the first half centimetre ($ALD_{10} = 0.3$ cm). *Ammobaculites agglutinans*, *U. peregrina*, *Bulimina alazanensis* and *Cribrostomoides* spp. are present at the surface and slightly deeper in the sediment ($ALD_{10} = 0.3$, 0.4 , 0.5 and 0.7 cm, respectively), whereas *Reophax dentaliniformis* reaches a maximal density at 1–1.5 cm sediment level ($ALD_{10} = 0.9$ cm);
- at station FP2 (4800 m), all dominant species are restricted to the superficial sediment layer. *Melonis pompilioides*, *Nuttallides umboniferus*, *Eggerella bradyi* and *Quinqueloculina seminula* have an ALD_{10} between 0.3 and 0.6 cm. The ALD_{10} of the total fauna is 0.5 cm.

Fig. 6 (upper panel) shows the spatial evolution of the ALD_{10} of the total faunas along the bathymetrical transect. The ALD_{10} is maximum (> 1.5 cm) at the outer shelf and upper slope stations. It varies between 0.8 and 1.3 cm for the stations between 550 and 2000 m depth, and becomes minimal (about 0.5 cm) at the two deepest stations (2400 and 4800 depth). This general decrease in ALD_{10} values with increasing water depths seems to be largely caused by a strong decrease in the density of intermediate and deep infaunal taxa (*C. oolina*, *Globobulimina* spp., *G. orbicularis*, *M. barleeanus*, *N. fabum* and *U. elongatastriata*) as shown in the upper panel of Fig. 6.

The lower panel of Fig. 6 shows the ALD_{10} of three species that are representative for the three main microhabitat patterns that we find in the Bay of Biscay: *Uvigerina peregrina* occupies a superficial infaunal microhabitat, *Melonis barleeanus* is always found in an intermediate infaunal niche and *Globobulimina* spp. systematically lives in a deep infaunal microhabitat. In Fig. 5, we can notice that the superficial taxon *U. peregrina* has a tendency towards a shallower microhabitat at deeper sites, whereas the intermediate and deep infaunal species *M. barleeanus* and *Globobulimina* spp. deepen their microhabitat towards deeper sites. The trend of the deeper infaunal species corresponds to a similar tendency observed for oxygen penetration into the sediment, which also increases towards deeper sites.

4. Discussion

4.1. Pore water oxygenation

Along the bathymetric/trophic transect, the oxygen penetration depth into the sediment shows a gradual increase from shallow to deeper sites. Although the oxygen penetration depth is controlled by several parameters (supply of labile

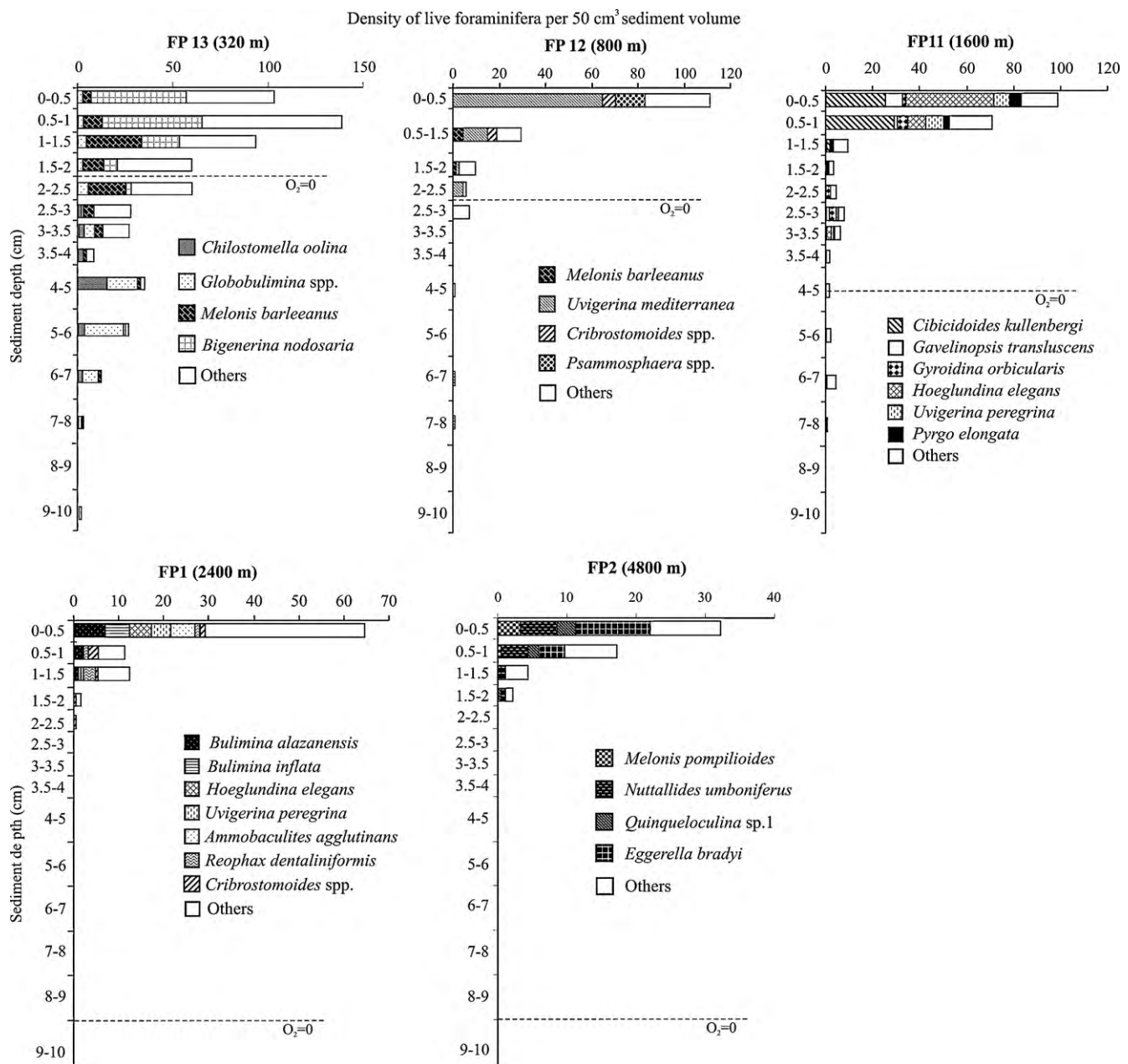


Fig. 5. Foraminiferal distribution (number of individuals found in each level, standardized for a 50 cm³ sediment volume) of the main species (present with a percentage higher than 5%) at stations FP13 (320 m), FP12 (800 m), FP11 (1600 m), FP1 (2400 m) and FP2 (4800 m). The dashed lines represent the maximum oxygen penetration depths.

Distribution (nombre d'individus trouvés à chaque niveau de la carotte, standardisé pour 50 cm³ de sédiment) des espèces majoritaires (présentes à $\geq 5\%$ dans au moins une station) au niveau des stations FP13 (320 m), FP12 (800 m), FP11 (1600 m), FP1 (2400 m) et FP2 (4800 m). Les lignes en pointillés représentent les limites maximum de pénétration d'oxygène dans le sédiment.

organic carbon, sediment porosity and bioturbation), it is mainly determined by the balance between downward diffusion and oxygen consumption in the sediment, due to aerobic organic carbon remineralisation and the oxidation of reduced products of anaerobic mineralization (e.g., Hyacinthe et al., 2001). It appears therefore that the very clear tendency of deeper oxygen penetration with increasing water depth reflects a strong decrease of metabolisable organic particles in function of water depth.

4.2. Densities of live foraminifera

Although we have no direct way to quantify the trophic level at any of our sampling stations, foraminiferal assemblages and pore water characteristics should vary in function of the amount of metabolisable organic carbon. As we saw in the previous paragraph, the oxygen penetration in the sediment should increase with a decreasing labile organic carbon flux to the sea floor. Simultaneously, the density of the foraminiferal

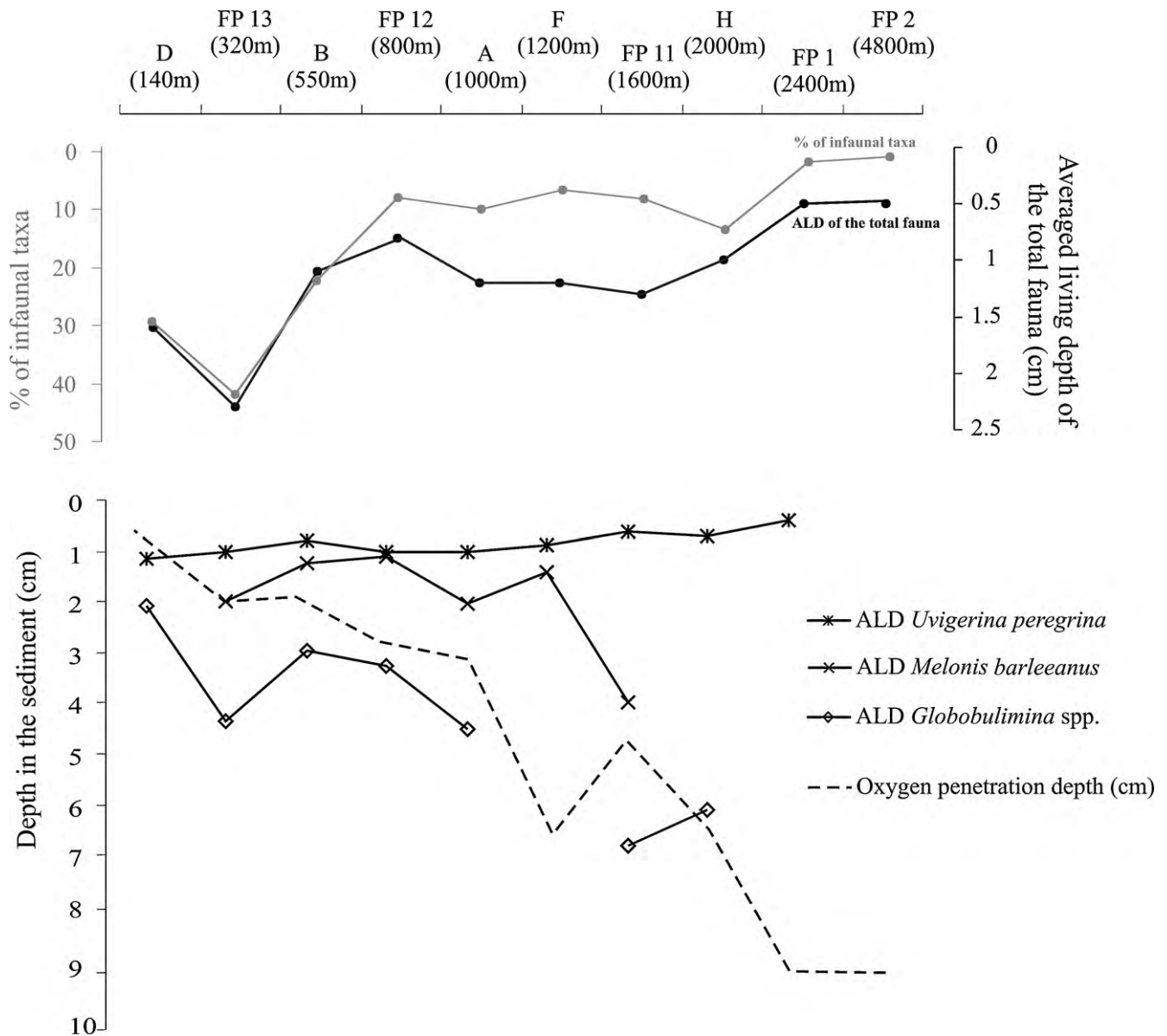


Fig. 6. Upper panel: ALD₁₀ of the total live fauna along the 10-stations bathymetric transect and cumulative percentages of infaunal taxa occurring with more than 5 % in at least one station (*C. oolina*, *Globobulimina* spp., *G. orbicularis*, *M. barleeanus*, *N. fabum* and *U. elongastriata*). Lower panel: oxygen penetration depth into the sediment and ALD₁₀ of three important species presenting the three main microhabitats found in the Bay of Biscay.

Partie supérieure : ALD₁₀ de la faune vivante totale au niveau des 10 stations du transect bathymétrique, ainsi que les pourcentages cumulatifs des taxons endopéiques présents à ≥ 5 % dans au moins une station (*C. oolina*, *Globobulimina* spp., *G. orbicularis*, *M. barleeanus*, *N. fabum* et *U. elongastriata*). Partie inférieure : pénétration d'oxygène dans le sédiment et ALD₁₀ de trois espèces importantes représentant les trois principaux microhabitats dans les sédiments du Golfe de Gascogne.

faunas decreases in function of water depth and oxygen penetration depth, logically revealing a strong correlation between the two parameters as well. It appears that both the density of the foraminiferal faunas and oxygen penetration depth into the sediment respond clearly to the diminishing flux of metabolisable organic carbon towards deeper waters. In the following paragraphs, we will therefore use both parameters to characterise the trophic level of our stations in a qualitative way.

On the basis of the total densities of the live foraminiferal faunas, and oxygen penetration depth into the sediment, we can differentiate 4 trophic zones along our sampling transect:

- at the shallowest station D (140 m), located at the outer continental shelf of the Bay of Biscay, live benthic foraminifera are present with very high densities (1430 ± 939 ind./50 cm²) and the oxygen penetration depth in the sediment is minimal (~0.5 cm). At this station, important differences between densities recorded during different sampling periods reflect the seasonal variation of organic matter deposition (Langezaal et al., 2006). This zone seems to present the most eutrophic conditions along the bathymetric transect;
- at the upper slope stations between 320 and 1000 m depth (FP13, B, FP12, A), live benthic foraminifera are present with

Table 3

Average Living Depth (ALD₁₀) of the main foraminiferal species (present with $\geq 5\%$ in at least one station) at the ten stations of the bathymetric transect. Between parentheses is indicated the number of individuals on which the calculation is based. An averaged value is given for stations D, B and A that were sampled at several periods.

ALD₁₀ des espèces vivantes majoritaires (présentes à $\geq 5\%$ dans au moins une station) au niveau des 10 stations. Le nombre d'individus utilisé pour les calculs est indiqué entre parenthèses. Une valeur moyenne est donnée pour les stations D, B et A ayant été échantillonnées à plusieurs périodes.

Stations	D	FP13	B	FP12	A	F	FP11	H	FP1	FP2
Depth (m)	140	320	550	800	20550	320550	1600	4800	2400	4800
<i>Perforates</i>										
<i>Bolivina subaenariensis</i>	0.8(651)	2.8(1)	4.5(1)							
<i>Bulimina alazanensis</i>									0.5(19)	
<i>Bulimina inflata</i>					1.1(7)	0.1(1)	0.5 (7)	0.5(3)	0.3(10)	
<i>Chilostomella oolina</i>	1.3 (223)	4.8 (37)	3.0(7)					6.5(1)		
<i>Cibicidoides kullenbergi</i>							0.5 (80)		0.8(3)	
<i>Cibicidoides robertsonianus</i>					0.4(1)	1.4(8)	0.8(11)			
<i>Gavelinopsis translucens</i>			0.1(1)			2(0.1)	0.3(15)	0.9 (4)	0.3(8)	
<i>Globobulimina</i> spp.	2.1 (6)	4.6 (84)	2.9 (37)	3.1(2)	3.1 (2)	7.5 (1)	2.9 (37)	6.1(5)		
<i>Gyroidina orbicularis</i>		0.3(1)	0.9 (2)	1.0 (2)	2.2 (9)		1.5(15)	1.3(1)	0.6(3)	1.3(1)
<i>Hoeglundina elegans</i>	1.7 (6)			1.0(1)	0.8 (42)	0.5 (12)	0.3 (68)	0.6(58)	0.3 (9)	
<i>Melonis berleeanus</i>		2.0 (66)	1.2(53)	1.1(7)	1.9(13)	1.4 (7)	4.0 (6)			
<i>Melonis pompilioides</i>										0.3 (7)
<i>Nonion fabum</i>	2.4 (366)		1.3(1)							
<i>Nuttallides umboniferus</i>			0.9(3)		0.6 (9)				0.3 (3)	0.5(18)
<i>Uvigerina elongatastrata</i>		2.3(14)	1.5(31)		1.3(1)					
<i>Uvigerina mediterranea</i>		1.4(18)	0.8(180)	0.7 (67)	0.9 (96)	0.2 (3)				
<i>Uvigerina peregrina</i>	1.2 (43)	1.0(2)	0.8 (54)	1.0 (2)	0.9 (94)	0.9(41)	0.6 (22)	0.7 (44)	0.9 (94)	
<i>Valvulineria bradyana</i>	0.9(210)									
<i>Porcellaneous</i>										
<i>Pyrgo elongata</i>		1.5 (9)					0.6(14)			
<i>Quinqueloculina</i> sp. 1	1.2(15)		0.3 (2)		0.5(1)	0.3 (7)	0.3 (2)	0.9(1)		0.6 (9)
<i>Fossilizing agglutinates</i>										
<i>Bigenerina nodosaria</i>		0.7(95)	0.5 (7)							
<i>Eggerella bradyi</i>					0.8 (3)		0.7 (6)		0.6 (6)	0.5 (29)
<i>Non-fossilizing agglutinates</i>										
<i>Ammobaculites agglutinans</i>					0.7(15)				0.3(11)	0.4(3)
<i>Clavulina cylindrica</i>	1.0(105)	1.5(18)	1.3(18)							
<i>Cribrostomoides</i> spp.	0.9 (2)		0.8 (27)	0.7(10)	0.8 (8)	2.1 (4)	1.9(12)	2.1 (4)	0.4(3)	0.4(3)
<i>Lagenammina pseudodifflugiformis</i>					1.0 (27)					
<i>Psammosphaera</i> spp.		0.3 (4)	0.5 (2)	0.3 (9)	1.4 (4)					
<i>Reophax dentaliniformis</i>		2.6 (2)	1.6 (2)				0.9(3)		0.9(9)	1.3 (2)
ALD ₁₀ of the entire fauna	1.6	2.3	1.21	0.8	1.2	1.2	1.3	1	0.5	0.5

high densities (more than 300 ind./50 cm² except at station FP12) and the oxygen penetration depth is varying from 1.9 to 2.8 cm. This zone seems to present eutrophic conditions, but less pronounced than at station D1;

- at the mid to lower slope stations, between 1200 and 2000 m depth (F, FP11, H), the oxygen penetration depth varies from 4.5 to 6.3 cm. Live foraminiferal faunas present densities around 100 ind./50 cm². We consider this zone as mesotrophic;
- at the stations deeper than 2000 m (FP1 and FP2), the oxygen penetration depth is 8 to 9 cm. Live foraminifera are present with very low densities (< 50 ind./50 cm²). We consider this zone as oligotrophic.

These observations agree with several earlier studies (e.g., Douglas, 1981; Altenbach and Sarthein, 1989; Gooday, 1993; Jorissen et al., 1998). However, many authors assume that each

single species has specific requirements with respect to the quantity and quality of organic supplies (Rathburn and Corliss, 1994; Altenbach et al., 1999; Fontanier et al., 2002).

4.3. Trophic dependencies of individual taxa

Several authors have suggested a critical threshold of total organic flux under which the majority of “eutrophic” species disappear and the assemblages become dominated by other species, more adapted to oligotrophic conditions (Altenbach et al., 1999; Jian et al., 1999; Morigi et al., 2001). In Fig. 7, we plotted the distribution of the main species along our bathymetric transect, together with curves of faunal density and oxygen penetration, which we use as qualitative indicators of the trophic level of the environment. It should of course be kept in mind that other environmental parameters than organic input alone, could have an impact on the depth distribution of the various species.

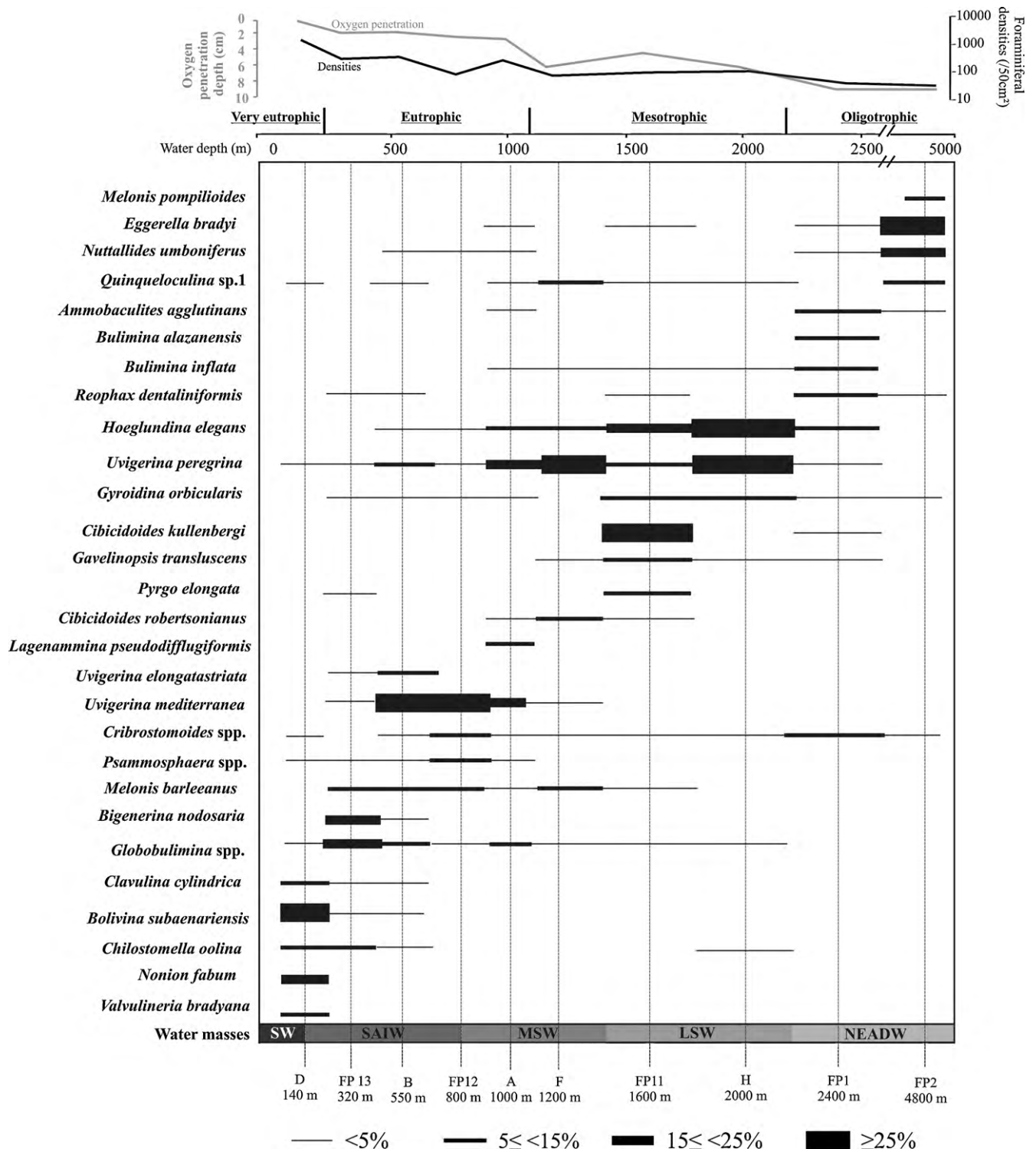


Fig. 7. Relative densities of the important live foraminiferal taxa (present with percentage higher than 5% in at least one station) along the 10-stations bathymetric transect.

Densités relatives des espèces majoritaires vivantes (présentes à $\geq 5\%$ dans au moins une station) au niveau des 10 stations du transect bathymétrique.

For instance, until the early 1980s, the benthic foraminiferal assemblages were thought to mainly reflect the water mass characteristics where they are found (e.g., Streeter, 1973; Lutze, 1979; Caralp, 1987). Nowadays, many scientists doubt the validity of the so-called “water mass concept” (Jorissen et al., 2007). Our data clearly suggest that in our study area, species occur-

rences are not related to water masses. In fact, none of the dominant species is restricted to a single water mass (Fig. 7). The main species appear to be distributed according to different trophic conditions. On the basis of the information summarised in Fig. 7, we can separate the species (see the taxonomic reference list in Appendix F) into the following categories:

- the first group of species appears to be associated with highly eutrophic conditions: *Bolivina subaenariensis* is strongly dominant at the shallowest station D (140 m) and is nearly absent or absent at the other stations. In a previous study on the seasonal variability of foraminiferal faunas at this 140 m deep station (Langezaal et al., 2006), this species has been qualified as a shallow infaunal species, fairly tolerant of low oxygen values and able to react to high food concentrations with rapid growth. *Valvulineria bradyana* and *Nonion fabum* are only present at station D (140 m). In the Adriatic Sea, Jorissen (1987) describes a minimum water depth of 40 m for *V. bradyana*, at the outer part of the organic carbon enriched clay belt. In a more recent study, this taxon seems to combine a preference for high quality resources, a high competitiveness and perhaps a high tolerance for anoxic conditions (Fontanier et al., 2002). It settles together with *N. fabum* (= *N. scaphum* in Fontanier et al., 2002) in strongly suboxic and anoxic layers slightly deeper in the sediment. *Chilostomella oolina* is well represented in the live faunal assemblages at stations D (140 m) and FP13 (320 m); it is present with very low proportions at stations B (550 m) and H (2000 m) and is totally absent at the other stations. In the literature, *C. oolina* has been described as an intermediate or deep infaunal species which is well adapted to suboxic conditions (Corliss and Emerson, 1990; Bernhard, 1992; Rathburn and Corliss, 1994; Jorissen et al., 1998; Licari et al., 2003). In these stressed environments, this taxon appears to have a competitive advantage over more superficially living taxa (Rathburn and Corliss, 1994). *Clavulina cylindrica* is present only between stations D (140 m) and B (550 m) in the study area with very rather low relative frequencies;
- the second group of species appears to be associated with eutrophic conditions: *Uvigerina mediterranea* dominates the live assemblages between stations B (550 m) and A (1000 m) with a maximal relative frequency at station FP12 (800 m). Its proportion diminishes at station FP13 (320 m) and station F (1200 m). This species is totally absent at all stations below 1200 m. De Rijk et al. (2000) suggest an inferior limit of about 2.5 g labile organic C. m⁻². yr⁻¹ whereas Altenbach et al. (1999) observe a maximal abundance around 10 g total organic C. m⁻². yr⁻¹. *Uvigerina elongatastriata* is present only at stations FP13 (320 m) and B (550 m), with much lower percentages than *U. mediterranea*. *Melonis barleeanus* is found between stations FP13 (320 m) and FP11 (1600 m). Its maximal relative abundance is recorded at station FP13 (320 m). In the literature, this species is generally associated with important amounts of organic matter in deep sea environments (e.g., Lutze and Coulbourn, 1984; Mackensen et al., 1985; Corliss and Chen, 1988; Caralp, 1989; Jorissen et al., 1995; Fontanier et al., 2002). *Globobulimina* spp. is present with very low proportions between stations FP13 (320 m) and H (2000 m). Percentages above 5.0% are only recorded between stations FP13 (320 m) and A (1000 m). This species has been described in various high productivity areas (e.g., Altenbach, 1985; Fariduddin and Loubere, 1997). A recent study (Risgaard-Petersen et al., 2006) indicates that *Globobulimina pseudospinescens*, and possibly many other infaunal taxa, are able to store, and to respire nitrates, allowing them to live in anoxic sediments for prolonged periods of time. It is very probable that *G. pyrula* has a similar adaptation (Larkin and Gooday, 2009). Also the agglutinated species *Lagenammina pseudodifflugiformis*, *Bigenerina nodosaria* and *Psammosphaera* spp. seem to prefer eutrophic areas in the Bay of Biscay. In fact, *L. pseudodifflugiformis* is only present alive at station A (1000 m) with relatively low percentages. *B. nodosaria* is dominant at station FP13 (320 m) and nearly disappears at station B (550 m). *Psammosphaera* spp. is present with percentages greater than 5.0% only at station FP12 (800 m). At the other stations, this species accounts for less than 1.0% of the total fauna. *Cribrostomoides* spp. is present at nearly all stations with very low percentages. Percentages above 5.0% are only recorded at stations FP12 (800 m) and FP1 (2400 m). This species does not show a very clear trophic preference;
- the third group of species is more or less limited to sites with conditions which we consider as mesotrophic: *Uvigerina peregrina* has a wide distribution, between stations FP13 (320 m) and FP1 (2400 m). This species is abundant at stations A (1000 m), F (1200 m) and H (2000 m) and present in fair numbers at stations B (550 m), FP11 (1600 m) and FP1 (2400 m). In the literature, this species has been described in areas with high organic fluxes (Lutze, 1980; Altenbach and Sarthein, 1989; Licari et al., 2003; Schönfeld, 2006). Altenbach et al. (1999) describe it at sites with estimated carbon organic fluxes from 2 to 20 gC. m⁻². yr⁻¹ (Altenbach et al., 1999), with a maximum between 5 and 9 gC. m⁻². yr⁻¹. The studies of Lutze (1986), Van Leeuwen (1986) and Schönfeld (2006) underline the existence of a large morphological variability in this taxon, with the different morphotypes having different dependencies on organic flux levels. These different morphotypes, which very probably have different ecological requirements, can be distinguished as distinct species or subspecies, or rather be differentiated as morphotypes within a single species. In our study area, we find a gradual transition from typical *peregrina*-types (pl. 1, Figs. 1–6 in Lutze, 1986) at the deeper sides, to *parva*-types (pl. 3, Figs. 1–5 in Lutze, 1986) at the shallower sites where *U. peregrina* occurs. Because of the very gradual transition between the two morphotypes, without a sharp break, we decided not to count them separately. *Hoeglundina elegans* is very abundant at stations FP11 (1600 m) and H (2000 m) and occurs with fair proportions at stations A (1000 m), F (1200 m) and FP1 (2400 m). In the literature, this species is often described as typical of low organic carbon areas (Lutze and Coulbourn, 1984; Corliss, 1985; Corliss and Emerson, 1990; Corliss, 1991; Fontanier et al., 2002; Morigi et al., 2001). *Cibicidoides kullenbergi* is particularly abundant at station FP11 (1600 m) and is present with a low percentage at station FP1 (2400 m). Altenbach et al. (1999) record it in an area with an estimated total carbon organic flux of 1.8 to 3.8 gC. m². yr⁻¹. Fariduddin and Loubere (1997) and Schmiedl et al. (1997) suggested that the ecological requirements of this species are very similar to those of *Hoeglundina elegans*. *Gavelinopsis translucens* is relatively well represented

between stations F (1200 m) and FP1 (2400 m), *Cibicidoides robertsonianus* is only present at stations F (1200 m) and FP11 (1600 m), *Gyroidina orbicularis* is present with significant percentages only between stations FP11 (1600 m) and H (2000 m), whereas *Pyrgo elongata* is present with 5.0% of the live assemblages at station FP11 (1600 m). In the Bay of Biscay, all these species seems to prefer mesotrophic settings;

- the fourth group contains species that are associated with the most oligotrophic conditions found along our transect: *Bulimina alazanensis* is only found at station FP1 (2400 m) where it dominates the live fauna. In the south Atlantic, this species is found in significant numbers between 1600 and 3700 m (Schmiedl et al., 1997). *Nuttallides umboniferus* dominates the fauna at station FP2 (4800 m) is present with very low percentages at stations FP1 (2400 m), A (1000 m) and B (550 m). Schmiedl et al. (1997) ascribed its abundance in the abyssal part of the South Atlantic Ocean to a preference for oligotrophic settings. *Eggerella bradyi* is an agglutinated species which is particularly abundant at station FP2 (4800 m). At the other stations where it is found (FP1, 2400 m; FP11, 1600 m and A, 1000 m), its relative proportion does not exceed 4.0%. *Ammobaculites agglutinans* and *Reophax dentaliniformis* present both maximal abundances at FP1 (2400 m). *R. dentaliniformis* has a wider distributional area. It is found between stations FP13 (320 m) and FP2 (4800 m) but with proportions less than 2.0%. In general, a dominance of agglutinated species reflects the oligotrophic character of the benthic ecosystem (Jorissen et al., 1998). *Melonis pompilioides* is only present at station FP2 at 4800 m water depth. This species is always found at great water depths (Lutze, 1980; Harloff and Mackensen, 1997; Jorissen et al., 1998). *Quinqueloculina* sp. 1 is present at nearly all stations, generally with very low abundances. However, its maximal density is recorded at station FP2 (4800 m). *Bulimina inflata* is present between stations B (550 m) and FP2 (4800 m) with a maximum at station FP1 (2400 m). *B. inflata* is a shallow infaunal surface dweller (Corliss, 1985, 1991; Mackensen and Douglas, 1989; Corliss and Emerson, 1990; Rathburn et al., 1996; Fontanier et al., 2002) which may reflect well oxygenated microenvironments and/or the presence of labile organic carbon deeper in the sediment. According to other studies, *B. inflata* appears to prefer slightly more eutrophic conditions, such as those encountered in the mid-slope environments under the very productive surface waters off Cape Blanc, NW Africa (Jorissen et al., 1998; Morigi et al., 2001). It is possible that this apparently contradictory evidence is caused by the fact that forms which are genetically different, but morphologically similar, have been lumped within the same species.

The evolution of the species composition of the foraminiferal faunas along the bathymetric transect confirms the general picture of decreasing organic fluxes from the continental shelf to the abyssal area in the Bay of Biscay; we observe high densities of species typical of eutrophicated environments at the shallower stations, that are gradually replaced by species typical of oligotrophic settings in the abyssal area. It appears that most species

have indeed a pronounced preference for a certain trophic level, or a specific organic flux range. Within the range of optimal trophic conditions, the species is highly competitive, and may attain high densities, and consequently, elevated percentages. Beyond critical upper and/or lower threshold values, species rapidly become less competitive. They are therefore “replaced” by other taxa, which are better adapted and more competitive in the trophic conditions in this domain. Koho et al. (2008) observe relatively similar trend in the distribution of abundant taxa in the Portuguese margin.

Although the quantity of organic carbon appears to be important to explain the foraminiferal distributional patterns in the Bay of Biscay, many studies suggest also a strong linkage between faunal characteristics and the quality (reactivity) of the organic matter arriving at the sea floor (e.g., De Rijk et al., 2000; Koho et al., 2007; Mojtahid et al., 2009). It is the relatively more labile organic carbon which is remineralised. Unfortunately, due to a lack of information on organic matter quality, this factor could not be considered in this study. Also, our study does not integrate the short term variability of the standing stocks and of the composition of the foraminiferal faunas in response to interannual and seasonal primary production oscillations and the ensuing organic fluxes to the sea floor. The densities of some very reactive opportunistic species, such as *Uvigerina peregrina* and *Epistominella exigua*, appear to depend strongly on the supplies of highly reactive organic carbon (e.g., Gooday, 1993; Fontanier et al., 2003, 2006). Their density is probably not representative for an average annual organic flux level, but is rather determined by the appearance of pulsed organic matter flux events in the weeks to month before sampling. This appears to be due to their capability to rapidly adapt to changing trophic conditions (Gooday and Lambshead, 1989; Ohga and Kitazato, 1997; Kitazato et al., 2000; Jorissen et al., 2007).

4.4. Microhabitats

According to the TROX-model (Jorissen et al., 1995), proposed for open marine benthic foraminiferal faunas, shallow infaunal foraminifera profit from labile organic matter supplies in the well oxygenated niches around the sediment–water interface, whereas deep infaunal foraminifera may survive in strongly hypoxic, or even anoxic sediments (e.g., Bernhard and Reimers, 1991; Bernhard, 1992; Moodley and Hess, 1992; Bernhard et al., 1997), where labile organic particles are scarce (Koho et al., 2008). Most species are able to change their microhabitat in response to food supply and to dissolved oxygen concentration in the bottom and pore waters (Linke and Lutze, 1993; Jorissen et al., 1995). In food-limited environments (oligotrophic environments), commonly characterised by well oxygenated bottom waters, both foraminiferal standing stock and diversity are rather low and the fauna mainly comprises shallow infaunal taxa. In mesotrophic environments, standing stocks are moderately high and the faunal assemblages comprise a variety of shallow, intermediate and deep infaunal species, which show a vertical succession in topmost 5 cm of the sediment. Finally, in eutrophic and oxygen limited environments, low diversity faunas with high standing stocks prevail, often mainly composed of infaunal

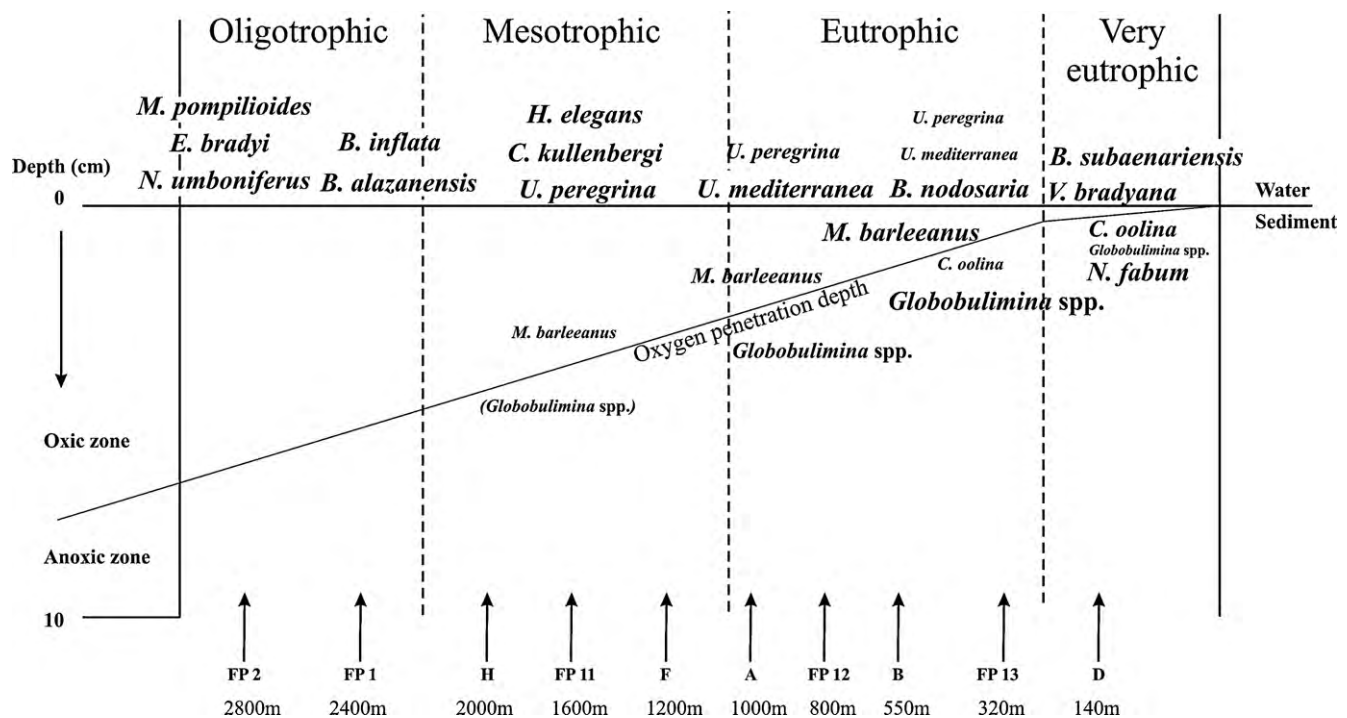


Fig. 8. Microhabitat distribution and specific foraminiferal composition along the bathymetric transect in the Bay of Biscay; the approximate position of our 10 stations is indicated (see text for full explanation). The species written in large font character represent the taxa that are very abundant and largely dominate the faunas. The species written in smaller font are still dominant but occur in lower proportions.

Microhabitat et composition des foraminifères vivants le long du transect bathymétrique dans le Golfe de Gascogne; la position approximative des 10 stations est indiquée (voir texte pour l'explication complète). Les espèces figurées en grand caractère sont les espèces abondantes et largement dominantes; celles figurant en petit caractère restent dominantes mais sont présentes avec des plus faibles proportions.

species that are adapted to strongly dysoxic conditions (Jorissen et al., 1995), which are found close to the sediment–water interface here. In such environments, the vertical species succession is often strongly compressed, and taxa which elsewhere occupy deep infaunal niches may be found close to, or at the sediment surface.

At all our stations, the live faunas are strongly concentrated in the first cm of the sediment, suggesting a strong dependence on the flux of labile, easily consumable organic matter. Despite the strong faunal concentration in the first cm, the ALD₁₀ of the total faunas is not uniform: 1.6 and 2.3 cm for the relatively shallow stations D and FP13, between 0.8 and 1.3 cm for the intermediate stations (B, FP12, A, F, FP11 and H) and 0.5 cm for the deepest stations FP1 and FP2. The deepening of the ALD₁₀ for the total fauna at station FP13 is mainly the result of the high abundance of intermediate (e.g., *Melonis barleeanus*) and deep infaunal species (*Globobulimina* spp. and *Chilostomella oolina*). The very shallow ALD₁₀ of the deepest stations (FP1 and FP2) is mainly caused by the scarcity of deep infaunal species.

Fig. 8 synthesises the spatial and microhabitat distribution of live benthic foraminifera along the bathymetric/trophic transect: The shallower station D, described in detail by Langezaal et al. (2006), is characterised by very high foraminiferal densities that are largely restricted to the upper 3 cm of the sediment. Apparently this fauna reflects a very high labile organic carbon flux. *Valvulineria bradyana* and *Bolivina subaenariensis* dominate the first half cm. *Chilostomella oolina* and *Globobulimina*

spp. settle together with *Nonion fabum* in strongly suboxic and anoxic sediments down to 3 cm depth. A major faunal change takes place at station FP13 (320 m), i.e. the intermediate between D (140 m) and B (550 m). This station is characterized by the disappearance of *N. fabum* and *V. bradyana* and the appearance of the uvigerinids (*U. mediterranea*, *U. elongastriata* and *U. peregrina*). At this station, the deepest ALD₁₀ (2.3 cm) is recorded. The intermediate (*M. barleeanus*) and deep infaunal (*Globobulimina* spp.) taxa, found fairly close to the sediment surface at station D, are living deeper in the sediment here. The almost complete separation between superficial and deeper infaunal species (Fig. 5) is probably due to a deeper penetration of oxygen into the sediment and an expanded succession of the successive redox zones. Towards deeper sites, until about 2000 m water depth, the surface faunas are dominated by uvigerinids. The proportion of deep infaunal species strongly decreases whereas their microhabitat deepens. This appears to reflect a progressive decrease of organic carbon fluxes and a progressively deeper position of the oxygen penetration limit. The appearance of more oligotrophic shallow infaunal taxa, such as *H. elegans*, *N. umboniferus* and *C. kullenbergi* shows the influence of the gradually decreasing organic flux towards deeper areas. The combination of the progressive disappearance of deep infaunal taxa and the increasing proportion of shallow infaunal species leads to a lower ALD₁₀ of the total fauna at these stations. A critical threshold of organic flux is attained between station H (2000 m) and FP1 (2400 m). In fact, between those two sta-

Appendix A (Continued)

Station FP13																
Sediment level (cm)	0–0.5	0.5–1	1–1.5	1.5–2	2–2.5	2.5–3	3–3.5	3.5–4	4–5	5–6	6–7	7–8	8–9	9–10	Total	%
<i>Porcellaneous</i>																
<i>Biloculinella</i> spp.	1		2												3	0.62
<i>Miliolinella subrotunda</i>		2													2	0.41
<i>Pyrgo elongata</i>		3	2	3	1										9	1.86
<i>Pyrgoella sphaera</i>		1													1	0.21
<i>Fossilizing agglutinates</i>																
<i>Bigenerina nodosaria</i>	36	38	14	5	2					1					96	19.79
<i>Clavulina cylindrica</i>	1	5	4	6	1		1								18	3.71
<i>Sigmoilopsis schlumbergeri</i>		1													1	0.21
<i>Siphotextularia concava</i>	2	2	2					1		1		1			9	1.86
<i>Textularia agglutinans</i>	5	11	1	1	1			1						1	21	4.33
<i>Non-fossilizing agglutinates</i>																
<i>Haplophragmoides</i> spp.			1												1	0.21
<i>Psammosphaera</i> spp.	3	1													4	0.82
<i>Pseudoclavulina crustata</i>	1														1	0.21
<i>Recurvoides</i> spp.		5	3		1										9	1.86
<i>Reophax dentaliniformis</i>			1		1										2	0.41
<i>Reophax spiculifer</i>		1	3												4	0.82
<i>Total living specimens</i>	74	100	67	43	43	20	19	6	50	38	17	5	1	2	485	100
<i>Species number</i>	19	26	19	16	16	9	11	5	6	6	4	4	1	2		

Appendix B. Census data (non-standardized) of live benthic foraminifera at station FP12. All numbers are based on the study of one interface core with a $\sim 71 \text{ cm}^2$ surface area

Données non standardisées des foraminifères vivants à la station FP12 basées sur l'étude d'une carotte de $\sim 71 \text{ cm}^2$ de surface

[illegible]

Appendix B (Continued)

Station FP12															
Sediment level (cm)	0-0.5	0.5-1.5	1.5-2	2-2.5	2.5-3	3-3.5	3.5-4	4-5	5-6	6-7	7-8	8-9	9-10	Total	%
<i>Non-fossilizing agglutinates</i>															
<i>Ammolagena</i> spp.		1												1	0.71
<i>Cribrostomoides</i> spp.	4	6												10	7.14
<i>Glomospira charoides</i>	3													3	2.14
<i>Psammosphaera</i> spp.	9													9	6.43
<i>Recurvoides</i> spp.	1	1												2	1.43
<i>Tritaxis challengerii</i>			2											2	1.43
Total living specimens	79	42	7	4	5			1		1	1			140	1
Species number	15	14	6	2	2			1		1	1				

Appendix C. Census data (non-standardized) of live benthic foraminifera at station FP11. All numbers are totals for 2 investigated cores, each with a ~71 cm² surface area

Données non standardisées des foraminifères vivants à la station FP11. Les nombres sont les totaux basés sur l'étude de 2 carottes de ~71 cm² de surface chacune

Station FP11																
Sediment level (cm)	0-0.5	0.5-1	1-1.5	1.5-2	2-2.5	2.5-3	3-3.5	3.5-4	4-5	5-6	6-7	7-8	8-9	9-10	Total	%
<i>Perforates</i>																
<i>Bulimina inflata</i>	3	4				1	1							1	10	3.1
<i>Cibicidoides kullenbergi</i>	36	42	2			1									81	25.4
<i>Cibicidoides robertsonianus</i>	1	8	2												11	3.4
<i>Cyclogyra involvens</i>		1			1										2	0.6
<i>Fissurina</i> sp.					1		1								2	0.6
<i>Gavelinopsis translucens</i>	10	2				2					1		1		16	5.0
<i>Globobulimina</i> spp.											2	1			3	0.9
<i>Gyroidina orbicularis</i>	2	6			3	4			1		1				17	5.3
<i>Hoeglundina elegans</i>	53	11				1	3				1	1			70	21.9
<i>Melonis barleeanus</i>					1	1		1	2		1				6	1.9
<i>Nonion</i> sp.		1													1	0.3
<i>Oridorsalis umbonatus</i>	2					1									3	0.9
<i>Pullenia bulloides</i>									1						1	0.3
<i>Pullenia quinqueloba</i>	1		1	1						4					7	2.2
<i>Uvigerina peregrina</i>	10	11		1			1						1		24	7.5
<i>Porcellaneous</i>																
<i>Miliolinella</i> spp.	3						1								4	1.3
<i>Pyrgo elongata</i>	7	4	2	1			1		1						16	5.0
<i>Pyrgo subsphaerica</i>	5	1													6	1.9
<i>Quinqueloculina</i> sp. 1	2														2	0.6
<i>Spiroloculina</i> sp.	1														1	0.3
<i>Fossilizing agglutinates</i>																
<i>Egerella bradyi</i>	1	5						1							7	2.2
<i>Textularia</i> sp.	1														1	0.3
<i>Non-fossilizing agglutinates</i>																
<i>Cribrostomoides</i> spp.	1	4	2	2		1				2		1			13	4.1
<i>Glomospira cordialis</i>			1												1	0.3
<i>Pseudonosaria</i> sp.	1														1	0.3
<i>Reophax dentaliniformis</i>		2	1												3	0.9
<i>Trochammina</i> spp.			2								8				10	3.1
Total living specimens	140	102	13	5	6	12	8	2	5	6	14	3	2	1	319	100
Species number	18	14	8	4	4	8	6	2	4	2	6	3	2	1		

Appendix D. Census data (non-standardized) of live benthic foraminifera at station FP1. All numbers are totals for 6 investigated cores, each with a $\sim 31 \text{ cm}^2$ surface area

Données non standardisées des foraminifères vivants à la station FP1. Les nombres sont les totaux basés sur l'étude de 6 carottes de ~31 cm² de surface chacune

[illegible]

Appendix E. Census data (non-standardized) of live benthic foraminifera at station FP2. All numbers are totals for 6 investigated cores, each with a ~31 cm² surface area

Données non standardisées des foraminifères vivants à la station FP2. Les nombres sont les totaux basés sur l'étude de 6 carottes de ~31 cm² de surface chacune

Station FP2																
Sediment level (cm)	0-0.5	0.5-1	1-1.5	1.5-2	2-2.5	2.5-3	3-3.5	3.5-4	4-5	5-6	6-7	7-8	8-9	9-10	Total	%
Perforates																
<i>Cibicides wuellerstorfi</i>	4														4	3.8
<i>Epistominella exigua</i>	2	1	1												4	3.8
<i>Gyroidina orbicularis</i>			1												1	1.0
<i>Melonis pompilioides</i>	6	1													7	6.7
<i>Melonis zaandamae</i>		1													1	1.0
<i>Nutallides umboniferus</i>	10	7	1												18	17.3
<i>Oridorsalis umbonatus</i>	1	2	1												4	3.8
Porcellaneous																
<i>Biloculina</i> spp.	2	2													4	3.8
<i>Pyrgo subsphaerica</i>	1	1	1												3	2.9
<i>Pyrgoella sphaera</i>		1													1	1.0
<i>Quinqueloculina</i> sp. 1	5	3		1											9	8.7
Fossilizing agglutinates																
<i>Eggerella bradyi</i>	20	7	1	1											29	27.9
Non-fossilizing agglutinates																
<i>Ammobaculites agglutinans</i>	2	1													3	2.9
<i>Cribrostomoides</i> spp.	2	1													3	2.9
<i>Eggerella</i> spp.	1														1	1.0
<i>Haplophragmoides</i> spp.	2														2	1.9
<i>Reophax dentaliniformis</i>		1		1											2	1.9
<i>Reophax piluliferis</i>	1														1	1.0
<i>Reophax scorpiurus</i>		1													1	1.0
<i>Robertinoides bradyi</i>			1	1											2	1.9
<i>Saccamina</i> spp.		1													1	1.0
<i>Scutulloris</i> sp.	1	1	1												3	2.9
<i>Total living specimens</i>	60	32	8	4											104	100
<i>Species number</i>	15	16	8	4												

Appendix F. Taxonomic reference list. For all taxa determined at a species level, a modern reference is given, in which the species is correctly figured

Liste des références taxonomiques. Pour chaque taxon déterminé au niveau spécifique, une référence récente, dans laquelle l'espèce est correctement figurée, est donnée

Species	References
<i>Ammobaculites agglutinans</i> (d'Orbigny, 1846)	Hess (1998), pl. 4, fig. 4
<i>Amphicoryna scalaris</i> (Batsch, 1791)	Jones (1994), pl. 63, figs. 28–31
<i>Bigenerina nodosaria</i> (d'Orbigny, 1826)	Jones (1994), pl. 44, figs. 19–24
<i>Bolivina alata</i> (Seguenza, 1862)	Wright (1978), pl. 1, fig. 13
<i>Bolivina striatula</i> (Cushman, 1922)	Cimernan and Langer (1991), pl. 62, figs. 6–9
<i>Bolivina subaenariensis</i> (Cushman, 1922)	Phleger et al. (1953), pl. 7, figs. 24, 25
<i>Bolivinata quadrilatera</i> (Schwager, 1866)	Jones (1994), pl. 42, figs. 8–12
<i>Bulimina aculeata</i> (d'Orbigny, 1826)	Jones (1994), pl. 51, figs. 7–9
<i>Bulimina alazanensis</i> (Cushman, 1927)	Hess (1998), pl. 10, fig. 10
<i>Bulimina costata</i> (d'Orbigny, 1852)	Van Leeuwen (1989), pl. 8, figs. 2, 3
<i>Bulimina inflata</i> (Seguenza, 1862)	Van Leeuwen (1989), pl. 8, fig. 4
<i>Bulimina marginata</i> (d'Orbigny, 1826)	Hess (1998), pl. 10, fig. 7
<i>Cancris auriculus</i> (Fichtel and Moll, 1942)	Jones (1994), pl. 106, fig. 4
<i>Cassidulina carinata</i> (Silvestri, 1896)	Phleger et al. (1953), pl. 9, figs. 32–37
<i>Cassidulina crassa</i> (d'Orbigny, 1839)	Jones (1994), pl. 54, fig. 4
<i>Chilostomella oolina</i> (Schwager, 1878)	Jones (1994), pl. 55, figs. 12–14

Appendix F (Continued)

Species	References
<i>Cibicides wuellerstorfi</i> (Schwager, 1866)	Van Leeuwen (1989), pl. 10, figs. 1–9
<i>Cibicidoides kullenbergi</i> (Parker, 1953)	Wright (1978), pl. 4, figs. 5–7
<i>Cibicidoides pachydermus</i> (Rzehac, 1886)	Jones (1994), pl. 94, fig. 9
<i>Cibicidoides robertsonianus</i> (Brady, 1881)	Van Leeuwen (1989), pl. 9, figs. 1–3
<i>Cibicidoides ungerianus</i> (d'Orbigny, 1846)	Marks (1951), pl. 8, figs. 2a, b
<i>Clavulina cylindrica</i> (Cushman, 1922)	Jones (1994), pl. 44, figs. 19–24
<i>Cyclogyra involvens</i> (Reuss, 1849)	Jones (1994), pl. 11, figs. 1–3
<i>Dentalina bradyensis</i> (Dervieux, 1894)	Jones (1994), pl. 62, figs. 19, 20
<i>Dentalina subemaciata</i> (Parr, 1950)	Jones (1994), pl. 62, figs. 25, 26
<i>Eggerella bradyi</i> (Cushman, 1911)	Jones (1994), pl. 47, figs. 4–7
<i>Epistominella exigua</i> (Brady, 1884)	Jones (1994), pl. 103, figs. 13, 14
<i>Gavelinopsis translucens</i> (Phleger and Parker, 1951)	Schiebel (1992), pl. 4, fig. 5
<i>Glomospira charoides</i> (Jones and Parker, 1860)	Phleger et al. (1953), pl. 5, fig. 1
<i>Glomospira cordialis</i> (Jones and Parker, 1860)	Phleger et al. (1953), pl. 5, fig. 2
<i>Gyroidina orbicularis</i> (d'Orbigny, 1826)	Jorissen (1987), pl. 1, fig. 13
<i>Gyroidina soldanii</i> (d'Orbigny, 1846)	Longinelli (1956), pl. 14, fig. 16
<i>Hanzawaia boueana</i> (d'Orbigny, 1846)	Jorissen (1987), pl. 3, fig. 10
<i>Hoeglundina elegans</i> (d'Orbigny, 1826)	Phleger et al. (1953), pl. 9, figs. 24, 25
<i>Hyalinea balthica</i> (Schroeter, 1783)	Jones (1994), pl. 112, figs. 1, 2
<i>Lagena hispidula</i> (Cushman, 1913)	Jones (1994), pl. 56, figs. 10, 11
<i>Lagenammina difflugiformis</i> (Brady, 1879)	Mackensen et al. (1990), pl. 6, fig. 9
<i>Lenticulina gibba</i> (d'Orbigny, 1839)	Jones (1994), pl. 69, figs. 8, 9
<i>Lenticulina peregrina</i> (Schwager, 1866)	Cushman and McCulloch (1950), pl. 39, fig. 5
<i>Lenticulina vortex</i> (Fitchell and Moll, 1798)	Jones (1994), pl. 69, figs. 14–16
<i>Marginula obesa</i> (Cushman, 1923)	Jones (1994), pl. 65, figs. 5, 6
<i>Melonis barleeanus</i> (Williamson, 1858)	Van Leeuwen (1989), pl. 13, figs. 1, 2
<i>Melonis pompilioides</i> (Fichtel and Moll, 1798)	Van Marle (1991), pl. 20, figs. 4–6
<i>Melonis zaandamae</i> (Van Voorthuysen, 1952)	Schmiedl et al. (1997), pl. 2, figs. 12, 13
<i>Miliolinella subrotunda</i> (Montagu, 1803)	Jones (1994), pl. 4, fig. 3
<i>Nonion fabum</i> (Fichtel and Moll, 1798)	Loeblich and Tappan (1988), pl. 690, figs. 1–7
<i>Nonionella turgida</i> (Williamson, 1858)	Jones (1994), pl. 109, figs. 17–19
<i>Nuttallides umboniferus</i> (Cushman, 1933)	Van Leeuwen (1989), pl. 15, figs. 11–13; pl. 16, figs. 1–7
<i>Oridorsalis umbonatus</i> (Reuss, 1851)	Van Leeuwen (1989), pl. 17, figs. 1–13
<i>Pseudoclavulina crustata</i> (Cushman, 1936)	Jorissen (1987), pl. 1, fig. 1
<i>Pullenia bulloides</i> (d'Orbigny, 1826)	Phleger et al. (1953), pl. 10, fig. 19
<i>Pullenia quinqueloba</i> (Reuss, 1851)	Marks (1951), pl. 7, fig. 19
<i>Pyrgo depressa</i> (d'Orbigny, 1826)	Jones (1994), pl. 2, figs. 12, 16, 17
<i>Pyrgo elongata</i> (d'Orbigny, 1826)	Jones (1994), pl. 2, fig. 9
<i>Pyrgo lucernula</i> (Schwager, 1891)	Jones (1994), pl. 2, figs. 5, 6
<i>Pyrgo murrhina</i> (Schwager, 1866)	Hess (1998), pl. 9, fig. 1
<i>Pyrgo subsphaerica</i> (d'Orbigny, 1839)	Cushman (1929), pl. 18, figs. 1, 2
<i>Pyrgoella sphaera</i> (d'Orbigny, 1839)	Jones (1994), pl. 2, fig. 4
<i>Reophax bilocularis</i> (Flint, 1899)	Schmiedl et al. (1997), pl. 1, figs. 3, 4
<i>Reophax dentaliniformis</i> (Brady, 1881)	Jones (1994), pl. 30, figs. 21, 22
<i>Reophax guttiferus</i> (Brady, 1881)	Jones (1994), pl. 30, figs. 10–15
<i>Reophax scorpiurus</i> (Montfort, 1808)	Loeblich and Tappan (1988), pl. 44, figs. 1–3
<i>Reophax spiculifer</i> (Brady, 1879)	Jones (1994), pl. 31, figs. 16, 17
<i>Reophax piluliferis</i> (Brady, 1884)	Jones (1994), pl. 30, figs. 18–20
<i>Robertinoides bradyi</i> (Cushman and Parker, 1936)	Jones (1994), pl. 50, fig. 18
<i>Sigmoilopsis schlumbergeri</i> (Silvestri, 1904)	Jones (1994), pl. 8, figs. 1–4
<i>Siphogenerina columellaris</i> (Brady, 1881)	Jones (1994), pl. 75, figs. 15–17
<i>Siphotextularia concava</i> (Karrer, 1868)	Jones (1994), pl. 42, figs. 13, 14
<i>Sphaeroidina bulloides</i> (d'Orbigny, 1826)	Longinelli (1956), pl. 10, fig. 1
<i>Spirillina decorata</i> (Brady, 1884)	Jones (1994), pl. 85, figs. 22–25
<i>Spiroplectammina wrightii</i> (Silvestri, 1903)	Jones (1994), pl. 42, figs. 17, 18
<i>Techinitella melo</i> (Norman, 1978)	Jones (1994), pl. 25, fig. 7
<i>Textularia agglutinans</i> (d'Orbigny, 1839)	Jones (1994), pl. 43, figs. 1–3
<i>Textularia conica</i> (d'Orbigny, 1839)	Le Calvez (1977), pl. 18, figs. 1, 2
<i>Trifarina bradyi</i> (Cushman, 1923)	Jones (1994), pl. 67, figs. 1–3
<i>Tritaxis challengerii</i> (Hedley, Hurdle and Burdett, 1964)	Jones (1994), pl. 41, fig. 3
<i>Uvigerina elongatastriata</i> (Colom, 1952)	Van der Zwaan et al. (1986), pl. 6, figs. 1–8
<i>Uvigerina mediterranea</i> (Hofker, 1932)	Van der Zwaan et al. (1986), pl. 5, figs. 1–7
<i>Uvigerina peregrina</i> (Cushman, 1923)	Van Der Zwaan et al. (1986), pl. 1, figs. 1–6
<i>Valvulineria bradyana</i> (Fornasini, 1900)	Jorissen (1987), pl. 4, figs. 1, 2

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