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# Unusual chemical composition of a Mexican propolis collected in Quintana Roo, Mexico

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## SUMMARY

Propolis is a resinous natural substance collected by honeybees from buds and exudates of various trees and plants; it is widely accepted that the composition of propolis depends on the phytogeographic characteristics of the site of collection. In this study we have analyzed the chemical composition of a propolis collected in Quintana Roo, Mexico, and evaluated its antioxidant, antifungal and antibacterial activities. Unexpectedly, the chemical analysis showed that the main components of the ethanolic extract of a Mexican propolis appeared to be pentacyclic triterpenoids, such as  $\alpha$  and  $\beta$ -amyrin derivatives, and sterols. The crude extract did not show antioxidant activity when tested using the DPPH-reduction assay, and it also proved inactive when tested for antifungal and antibacterial activities using microdilution and agar diffusion assays, respectively. The fact that the presence of both  $\alpha$  and  $\beta$ -amyrins and their derivatives have been reported from the resin of *Bursera simaruba*, one of the plants used by the bees for propolis production in Quintana Roo, Mexico, confirms the relationship that exists between the flora available to bees in a given region and the chemical composition of the propolis that they produce.

## RESUMEN

El propóleo es un producto natural resinoso colectado por abejas a partir de brotes y exudados de diferentes plantas y árboles; es ampliamente aceptado que la composición del propóleo depende de las características fitogeográficas del sitio de colecta. En este estudio analizamos la composición química de un propóleo colectado en Quintana Roo, México, y evaluamos sus actividades antioxidante, antifúngica y antibacteriana. Inesperadamente, el análisis químico mostró que los principales componentes del extracto etanólico del propóleo mexicano parecían ser triterpenoides pentacíclicos, tales como derivados de  $\alpha$  y  $\beta$ -amyrina y esteroides. El extracto crudo no mostró actividad antioxidante al ser evaluado en el ensayo de reducción de DPPH, y también se mostró inactivo al ser evaluado en cuanto a su actividad antifúngica y antibacteriana utilizando los ensayos de microdilución y de difusión en agar, respectivamente. El hecho de que la presencia de  $\alpha$  y  $\beta$ -amyrina y sus derivados ya se ha reportado en la resina de *Bursera simaruba*, una de las plantas utilizadas por las abejas para su producción de propóleo en Quintana Roo, México, confirma la relación que existe entre la flora disponible para las abejas en una región en particular y la composición química del propóleo que producen.

**Keywords:** Mexican propolis, pentacyclic triterpenes, amyrins, sterols, *Bursera simaruba*

**Short title:** Chemical composition of a Mexican propolis

## INTRODUCTION

Propolis is a resinous natural substance collected by honeybees from buds and exudates of various trees and plants, mixed with beeswax and salivary enzymes. Bees generally use propolis to seal and smooth out the internal walls of the hive, as well as a protective barrier against fungal and bacterial infections. Propolis has been used in folk medicine since ancient times due to its pharmacological potential associated with antioxidant (Cottica *et al.* 2011; Gülçin *et al.* 2010; Miguel *et al.* 2010), antifungal (Ota *et al.* 2001; Sawaya *et al.* 2002), antibacterial (Kujumgiev *et al.* 1999; Popova *et al.* 2005; Raghukumar *et al.* 2010) and anti-inflammatory (Castaldo and Capasso, 2002) properties.

Propolis is generally composed of 50% resin and balm (including polyphenolic compounds), 30% wax and fatty acids, 10% essential oils, 5% pollen and 5% various organic and inorganic compounds. Presently it is widely accepted that the composition of propolis depends on the phytogeographic characteristics of the site of collection (Marcucci, 1995; Sforcin and Bankova, 2011) and a number of attempts have been made to define propolis types taking into account geographic origin, chemical composition, and plant source (Park *et al.* 2002; Salatino *et al.* 2005; Sforcin and Bankova, 2011; Trusheva *et al.* 2006); recently, five propolis types have been defined according to their chemogeographic patterns (Salatino *et al.* 2011). It is also generally accepted that propolis from temperate climatic zones, like Europe, North America and non-tropical regions of Asia, originate mainly from the bud exudates of *Populus* species and are rich in flavonoids, phenolic acids and their esters (Bankova *et al.* 2000), while propolis from tropical regions, where no poplars and birches exist, are rich in prenylated derivatives of *p*-coumaric acid and benzophenones (Bankova, 2005a; Kumazawa *et al.* 2004; Sforcin and Bankova, 2011); other examples include lignan-containing propolis from Chile, Spain and Kenya, and a propolis from Myanmar containing cycloartanes and prenylated flavanones (Salatino *et al.* 2011).

Mexico is considered as one of the three most important honey exporters in the world, with around 40% of its production located in and around the Yucatan peninsula (Pino *et al.* 2006). However, in spite of its potential importance, studies on the chemical composition and biological activity of Mexican propolis are limited. A chemical investigation of a red propolis sample collected in Campeche, in the southeast region of Mexico, reported the presence of flavanones, isoflavans and pterocarpanes, suggesting a possible relationship between the chemical composition of the Mexican red propolis and plants of the *Dalbergia* genus (Lotti *et al.* 2010). Other investigations report the antimicrobial activity against gram negative bacteria of several propolis samples collected in Campeche, Mexico (Tolosa and Canizares, 2002), the correlation between antimicrobial activity and composition of propolis and plants collected in central Mexico (Londono Orozco *et al.* 2010), and the identification by GC-MS of over 100 volatile constituents from two propolis samples collected in Yucatan, Mexico (Pino *et al.* 2006). Finally, a study on the chemical composition and biological activity of three propolis samples collected in different arid and semiarid regions of Sonora, Mexico showed cinnamic and phenylpropanoic acid derivatives and flavonoids as the main components, and antibacterial, free-radical scavenging, and cytotoxic activities (Li *et al.* 2010; Velazquez *et al.* 2007). We wish to report herein on the evaluation of the antioxidant, antifungal and antibacterial activities of a tropical propolis sample from Quintana Roo, a state in the Yucatan peninsula of Mexico, and the unexpected identification of pentacyclic triterpenoids and sterols as the only components present in its ethanolic extract.

## **MATERIALS AND METHODS**

**Chemicals.** 1,1-Diphenyl-2-picrylhydrazyl (DPPH), Folin-Ciocalteu reagent, formic acid, and gallic acid, all analytical grade, were purchased from Sigma-Aldrich (St Quentin Fallavier,

France). 6-Hydroxy-2,5,7,8-tetramethylchroman-2-carboxylic acid (Trolox<sup>®</sup>) and 5'-caffeoylquinic acid (chlorogenic acid) were obtained from Acros Organics (Geel, Belgium).

**Propolis sample.** The propolis sample (224 g), collected from apiaries located in the area of Macario Gómez in Solidaridad, Quintana Roo, Mexico, was obtained from the Dzidzilche honey-distributing store of the "Productores y Realizadores de Miel Maya" cooperative.

**Extractions.** Propolis was first pulverized in the presence of liquid nitrogen into a homogenous powder. A 1 g portion of propolis powder was extracted three times with ethanol (20 mL) at room temperature for 2 h; the combined filtrates were maintained at -18°C overnight, filtered to remove waxes, and evaporated under reduced pressure to dryness to give the ethanolic extract (**E1a**). A second 1 g portion of propolis was successively extracted with cyclohexane (**E2**, 3x20 mL, 2h), DCM (**E3**, 3x10 mL, 2h), EtOAc (**E4**, 3x10 mL, 2h), and MeOH (**E5**, 3x10mL, 2h) to produce the corresponding low, medium low, medium high, and high polarity extracts.

**Determination of total polyphenol content.** Total polyphenol content was determined according to the Folin-Ciocalteu colorimetric method (Rebiai *et al.* 2011); 20 µL of extract solution (2.5 mg/mL) in MeOH were mixed with 280 µL of distilled water and 100 µL of Folin-Ciocalteu's phenol reagent. After 3 min, 1200 µL of distilled water and 400 µL of 20% aqueous sodium carbonate solution were added and 200 µL of each solution were put into a 96-well microtiter plate. A blank was prepared in the same way by using MeOH instead of the extract solution. The absorbance was measured on a Tecan Infinite M200 microplate spectrophotometer, at 760 nm after 30 min in the dark, at room temperature. Gallic acid was used to calculate the calibration curve (0.4-1.2 mg/mL;  $y = 0.5800x$ ;  $r^2 = 0.9941$ ) and the total

polyphenol content was expressed as milligram of Gallic Acid Equivalent per gram of extract (mg GAE/g). The determination was performed in triplicate.

**HPLC-DAD procedure.** 10 mg of extracts were dissolved in 1 mL of MeOH and centrifuged at 13000 rpm for 10 min prior to injection (10  $\mu$ L). HPLC analyses were run on a 2695 Waters<sup>®</sup> separation module equipped with a diode array detector 2996 Waters<sup>®</sup>. Separations were carried out on a Lichrospher<sup>®</sup> column 100 RP-18 (125x4 mm i.d., 5  $\mu$ m) protected with a Lichrocart<sup>®</sup> 4-4 guard cartridge (4x4 mm i.d.), using a flow rate of 1 mL/min. The mobile phase consisted of 0.1% formic acid in water (solvent A) and methanol (solvent B) and the separation was performed by the following linear gradient: 25-100% B (0-40 min), 100% B (40-45 min). UV detection was achieved at three wavelengths: 254, 280 and 350 nm.

**Analytical TLC.** Analytical TLC was performed on TLC Alugram Xtra SIL G/UV<sub>254</sub>, using a mixture of cyclohexane:AcOEt 85:15 as eluant. Spots in the chromatogram were visualized by spraying with vanillin-sulfuric acid reagent (2 mL of concentrated sulfuric acid in 98 mL of a 1:99 w/v vanillin: 95% ethanol solution) and heating the chromatograms at 110°C for five minutes.

**Fractionation by Flash Chromatography.** 18 g of propolis powder were macerated in ethanol (6 x 200 mL, 2 h) to give, after solvent evaporation, 4.2 g of crude extract **E1b**. **E1b** was totally dissolved in the minimum volume of DCM and the solution was mixed with 8.4 g of silica gel. The solvent was allowed to evaporate until a fine dry powder was obtained. The fractionation was performed by using a CombiFlash Teledyne ISCO apparatus with a silica gel column (Redisep silica 80g), with a gradient elution of 100% cyclohexane to 100%

EtOAc, and a flow rate of 35 mL/min. 200 tubes of 20 mL were collected and combined into 10 fractions on the basis of their TLC chromatographic profiles (cyclohexane: EtOAc 85:15).

**<sup>1</sup>H and <sup>13</sup>C NMR analyses.** NMR spectra were recorded in CDCl<sub>3</sub> using a Bruker Avance spectrometer at 500 MHz for <sup>1</sup>H and 125 MHz for <sup>13</sup>C.

**GC-MS procedure.** The GC-MS analysis was performed on non-derivatized samples using an Agilent Technologies Gas Chromatograph 6890N connected to an Agilent Technologies 5975B mass detector system with an ionization voltage of 70eV. Separations were carried out using a 30 m long, 0.32 mm id, 0.5 mm film ultra 1 100% dimethylpolysiloxane column. The temperature was programmed as follows: 180°C (3 min), 180-280°C at the rate of 10°C/min and 280°C (30 min). Helium was used as a carrier gas at a flow rate 1.5 mL/min. Injector and detector temperatures were set at 250 and 280°C, respectively. Metabolites were identified by comparing their retention times and/or their fragmentation patterns with those of authentic samples and/or those contained in the fragmentation pattern library of the equipment (NIST Library in Chem Station G1701DA).

**Evaluation of antioxidant activity using the DPPH assay.** The assay was carried following a modified procedure on a reported methodology (Abdel-Lateff *et al.* 2002). Sample and standards were diluted in absolute ethanol at 0.02 mg/mL from stock solutions prepared at 1 mg/mL in DMSO; 100 µL aliquots of these solutions were placed in 96-well plates and the reaction was initiated by adding 25 µL of freshly prepared DPPH solution (1mM) and 75 µL of absolute ethanol using the microplate reader's injector (Infinite<sup>®</sup> 200, Tecan, France) to obtain a final volume of 200 µL per well. After 30 minutes in the dark and at room temperature, the absorbance was determined at 517 nm. Ethanol was used as a blank, whereas

10, 25, 50, and 75  $\mu\text{M}$  solutions of Trolox (hydrophilic  $\alpha$ -tocopherol analog) were used as calibration solutions. A sample of 0.02 mg/mL chlorogenic acid was used as a quality control. The evaluation was carried out in triplicate. The DPPH-scavenging activity was compared with that of Trolox using the calibration curve. Result was expressed in terms of Trolox equivalents (micromoles of Trolox equivalents per gram of extract).

**Evaluation of the antifungal activity.** Antifungal activity was assayed on human pathogenic fungi, including a yeast (*Candida albicans* ATCC 66396) and an opportunistic mould (*Aspergillus fumigatus* CBS 11326). The strains were obtained from the parasitology and mycology laboratory, University Hospital Center, Angers, France. Both microorganisms were cultivated at 37 °C on yeast extract–peptone–dextrose agar (YPDA) containing 0.5 g/L chloramphenicol for two (*C. albicans*) or three (*A. fumigatus*) days. Tests were performed according to a procedure described by Alomar *et al.* (Alomar *et al.* 2012) following the guidelines of the approved reference method of the National Committee for Clinical Laboratory Standards (NCCLS) for yeasts (NCCLS, 1997) and filamentous fungi (NCCLS, 2002). Briefly, the yeast suspensions were prepared in RPMI-1640 culture medium and adjusted spectrophotometrically at 630 nm to reach a final concentration of *ca.*  $0.5 \times 10^3$  to  $2.5 \times 10^3$  cells/mL. The tests were performed using sterile 96 flat shaped well microtiter plates. Serial two-fold sample dilutions were made in DMSO. Sample solutions (E1a and fractions A, B and C) were dispensed at a volume of 5  $\mu\text{L}$  in triplicate into the wells to obtain final concentrations from 250 to 1.95  $\mu\text{g/mL}$ . After 48 h at 37°C for *C. albicans*, and 72 h for *A. fumigatus*, the spectrophotometric MIC endpoint was calculated from the turbidimetric data as the lowest sample concentration causing a growth inhibition equal to or greater than 80% of the control (MIC<sub>80</sub>). Amphotericin B was used as a positive control.

**Evaluation of antibacterial activity.** Antibacterial activity was evaluated on 21 bacterial strains obtained from the laboratory of bacteriology, University Hospital Center, Angers, France: seven strains of *Acinetobacter baumannii* (RCH, SAN008, 12, AYE, CIP7034, CIP107292, CIP5377), five of *Staphylococcus aureus* (ATCC25923, two methicillin-sensitive clinical isolates, two methicillin-resistant clinical isolates), two of *Escherichia coli* (ATCC25922 and a clinical isolate), three of *Pseudomonas aeruginosa* (ATCC27853 and two clinical isolates), and one clinical isolate each of *Enterobacter cloacae*, *Enterobacter aerogenes*, *Klebsiella oxytoca*, and *Salmonella enteritidis* (phage type 4). Tests were performed using the methodology described in the guidelines of the Comité de l'Antibiogramme de la Société Française de Microbiologie (CA-SFM, [www.sfm.asso.fr](http://www.sfm.asso.fr)). Briefly, a stock solution of each sample was prepared at 20 mg/mL in DMSO under sterile conditions. Serial dilutions were prepared (sample concentrations: 1, 10, 20, 30, 40, 50, 60, 80 and 100 µg/mL) and 100 µL of each dilution were added to 19.9 mL Mueller Hinton agar (Merck Germany) and transferred to Petri plates. Bacterial strains ( $2 \times 10^4$ ) were suspended in sterile NaCl aqueous solution (0.15M) and inoculated on the different Petri plates using the multipoint inoculator (AQS, England). After 24h of incubation at 37°C, the minimum inhibitory concentration (MIC<sub>100</sub> µg/mL) of each sample against each bacterial strain was determined.

## RESULTS

The propolis EtOH extract (**E1a**) did not show a significant polyphenol content ( $18.2 \pm 2.9$  mg GAE/g); this unexpected finding was confirmed by the lack of phenolic components in the HPLC profile of **E1a** and by its lack of antioxidant activity when tested in the DPPH radical reduction assay.

The sequential gradient extraction of the propolis sample using cyclohexane (**E2**), DCM (**E3**), EtOAc (**E4**), and MeOH (**E5**) showed that the majority of the components were extracted into the most abundant (> 60%) low-polarity cyclohexane fraction (**E2**, Table 1), which exhibited a TLC profile very similar to that of **E1a**. None of the remaining fractions was obtained in more than 5% yield and none showed the presence of significantly important components by TLC. These results implied that the studied propolis was predominantly composed of non polar components in addition to beeswax. Since beeswax could be removed by cold filtration in EtOH, **E1a** was chosen to carry out the chemical characterization of the propolis.

Flash chromatography purification of **E1b** yielded three major fractions (**A**, **B** and **C**; Fig. 1), which accounted for more than 70% of the total crude extract (Table 2). <sup>1</sup>H NMR analyses of **E1a** and fractions **A**, **B** and **C** showed predominantly signals in the range of 0.8-3.0 ppm, with a few between 4.0-5.5 ppm, while the majority of the signals in the corresponding <sup>13</sup>C NMR spectra were located between 15 and 60 ppm, in addition to a few in the range of 110-150 ppm, that matches to triterpenes profiles. The GC/MS analysis of the most abundant fraction **A**, which showed a major low-polarity component by TLC ( $R_f = 0.70$ ), allowed its identification as a mixture of  $\alpha$  and  $\beta$ -amyrenone (**I** and **II**, respectively; structures of metabolites **I-VIII** are shown in Fig. 2), containing a small amount of  $\alpha$ -amyrin acetate (**III**). The TLC analysis of fraction **B** showed the presence of a single component ( $R_f = 0.33$ ) that proved to be inseparable under TLC conditions using a number of solvent systems; GC/MS analysis of **B** allowed the identification of the main component as a mixture of  $\alpha$ -amyrin (**IV**, major) and  $\beta$ -amyrin (**V**, minor). Finally, the GC/MS analysis of the least-abundant fraction **C**, which showed a major high-polarity component in its TLC profile ( $R_f = 0.18$ ), allowed its identification as a mixture of sterols, with fucosterol (**VI**) being the major component, and a mixture of  $\beta/\gamma$ -sitosterol (**VII/VIII**) being the minor one.

Evaluation of the antimicrobial activities of **E1b** and fractions **A**, **B** and **C** showed that they did not exhibit neither antifungal nor antibacterial activities when tested towards two fungal species ( $MIC_{80} > 250 \mu\text{g/mL}$ ) and 21 bacterial strains ( $MIC_{100} > 100 \mu\text{g/mL}$ ), respectively.

## DISCUSSION

The results obtained in this investigation were unexpected since, to date, there are only a limited number of reports about triterpenoids occurring as major components in propolis (de Castro Ishida *et al.* 2011; Kalogeropoulos *et al.*, 2009; Melliou *et al.*, 2007; Furukawa *et al.* 2002; Ito *et al.* 2001; Velikova *et al.*, 2000) and none of the propolis types that have been chemically described until now list triterpenes as their major components (Salatino *et al.* 2011; Sforcin and Bankova, 2011). Additionally, our results are not in agreement with reports stating that triterpenoids are rarely reported as propolis constituents because most chemical analyses of propolis samples are carried out with alcoholic extracts (Negri *et al.* 2003), or that triterpenoids are only occasionally collected by bees along with other classes of secondary metabolites such as phenolics (Negri *et al.* 2000).

The two plants used by the bees for their production of propolis in the region of Quintana Roo where the material was collected are commonly known with the Mayan names of “chakah” [*Bursera simaruba* (L.) Sarg.] and “tsalam” [*Lysiloma latisiliquum* (L.) Benth.] (Luis Ignacio Hernández-Chávez, personal communication), and both species are found as part of the flora of what is known as a mature medium-statured semideciduous forest (*selva mediana*) (Schultz, 2009). While no phytochemical knowledge exists about *L. latisiliquum*, the bark and resin of *B. simaruba* have been reported to contain lignans and other phenolic metabolites (Maldini *et al.*, 2009; Peraza-Sánchez and Peña-Rodríguez, 1992), with the resin also reported to contain pentacyclic triterpenes such as the ubiquitous  $\alpha$ -amyrin (IV) and  $\beta$ -amyrin (V) (Peraza-Sánchez *et al.* 1995). Although no lignans could be detected in the analyzed samples, these findings could confirm the reported relationship that exists between the flora available to bees in a given region and the chemical composition of the propolis that they produce (Adelmann *et al.* 2007; Bankova, 2005a; de Castro Ishida *et al.* 2011; Dausch *et al.* 2008; Salatino *et al.* 2011; Sforcin and Bankova, 2011) and the fact that in tropical regions, the

chemical composition of the propolis can be highly variable because of the richness and versatility of the local flora (Trusheva *et al.* 2004).

Finally, it is well known that propolis is mainly used by bees as a defense material against parasites (Salatino *et al.* 2011) and to prevent bacterial and fungal infections and that often, while the biological activity of different propolis samples is similar, the secondary metabolites responsible for the activity can be different (Bankova, 2005b; Trusheva *et al.* 2006). Although **E1a** did not show significant antioxidant or antimicrobial activity, pentacyclic triterpenoids which include  $\alpha$ -amyrin (**IV**) and  $\beta$ -amyrin (**V**) and their various derivatives have been reported as antibacterials, antifungals and with cytotoxic activity (Choi *et al.* 2012; Chung *et al.* 2013; Cota *et al.* 2011; Irshad *et al.* 2013; Sharma *et al.* 2010); the lack of activity of both the propolis extract **E1a** and the purified fractions **A-C** in the assays for antioxidant and antimicrobial activity, suggests that the chemical composition of the propolis collected in Quintana Roo is of particular importance in the ecological interaction between the bees and the parasites and microorganisms that occur specifically in that region. Therefore additional research works will be needed to establish the mode of protective action of this Mexican propolis.

#### ABBREVIATIONS USED

**ATCC** American Type Culture Collection; **CBS** Central Bureau voor Schimmelcultures; **CDCl<sub>3</sub>** Deuterated chloroform; **DCM** Dichloromethane; **DMSO** Dimethylsulfoxide; **DPPH** Diphenylpicrylhydrazyl; **EtOAc** Ethyl Acetate; **GC/MS** Gas Chromatography coupled with Mass Spectrometry; **HPLC/DAD** High Performance Liquid Chromatography coupled with Diode Array Detector; **MIC** Minimum Inhibitory Concentration; **NCCLS** National Committee for Clinical Laboratory Standards; **NMR** Nuclear Magnetic Resonance; **RPMI**

Roswell Park Memorial Institute; **TLC** Thin Layer Chromatography; **UV** Ultraviolet; **YPDA** Yeast Peptone Dextrose Agar.

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#### SUPPORTING INFORMATION

HPLC profile of EtOH extract (E1a) and GC-MS data of E1b and fractions A-C are available in Supporting Information.

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## ILLUSTRATIONS AND GRAPHICS

Figure 1. TLC profiles of E1b and fractions A, B and C.

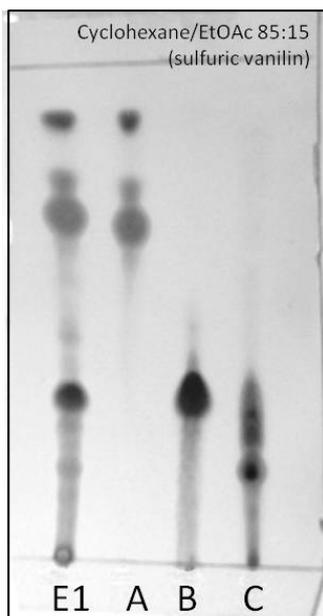
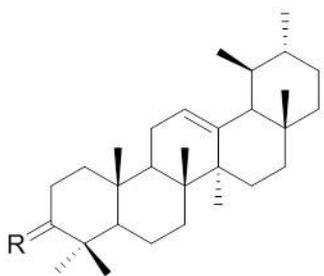
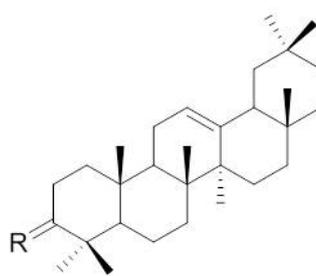


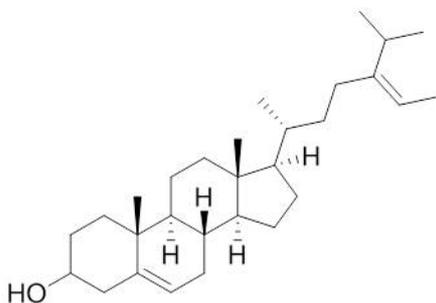
Figure 2. Structures of major constituents identified in E1b and fractions A, B and C by GC-MS analyses



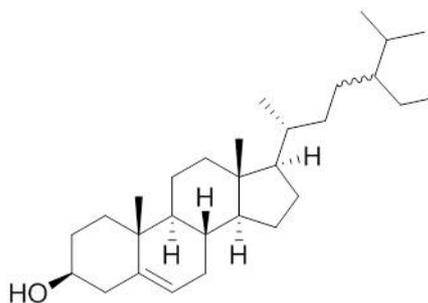
R: O,  $\alpha$ -amyrenone **I** (A)  
 R:  $\alpha$ =H,  $\beta$ =OAc,  $\alpha$ -amyrin acetate **III** (A)  
 R:  $\alpha$ =H,  $\beta$ =OH,  $\alpha$ -amyrin **IV** (B)



R: O,  $\beta$ -amyrenone **II** (A)  
 R:  $\alpha$ =H,  $\beta$ =OH,  $\beta$ -amyrin **V** (B)



Fucosterol **VI** (C)



$\beta$ / $\gamma$ -sitosterol **VII/VIII** (C)

TABLES

Table 1. Yields of propolis extracts

Extract	Solvent	Extraction yield (%)
<b>E1a</b>	EtOH	34.2
<b>E2</b>	Cyclohexane	61.6
<b>E3</b>	DCM	2.5
<b>E4</b>	EtOAc	1
<b>E5</b>	MeOH	4.6

Table 2. Yields of fractions **A**, **B** and **C** and  $R_f$  values of their major components on TLC

Fraction	Yield (%)	$R_f^*$
<b>A</b>	47.7	0.89, 0.77, <b>0.70</b>
<b>B</b>	19.9	<b>0.33</b>
<b>C</b>	4.1	<b>0.18</b>
Total	71.7	

\*Eluant system: cyclohexane:EtOAc 85:15

# Unusual chemical composition of a Mexican propolis collected in Quintana Roo, Mexico

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## SUPPORTING INFORMATION

List of supporting information:

Fig. S1. HPLC chromatographic profile of extract **E1a** at 254 (a), 280 (b) and 350 nm (c).

Fig. S2a. GC chromatographic profile of fraction **A**

Fig. S2b. MS spectra of components in fraction **A**

Fig. S3a. GC chromatographic profile of fraction **B**

Fig. S3b. MS spectra of components in fraction **B**

Fig. S4a. GC chromatographic profile of fraction **C**

Fig. S4b. MS spectra of components in fraction **C**

Fig. S5a. GC chromatographic profile of extract **E1b**

Fig. S5b. MS spectra of components in extract **E1b**

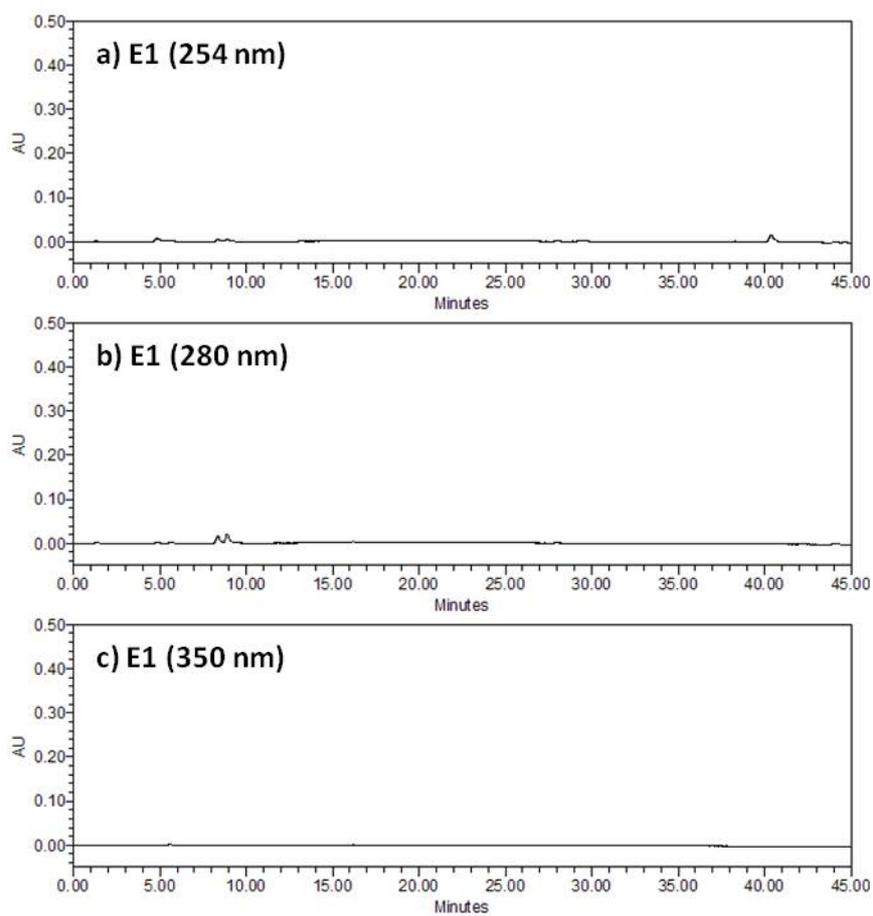


Fig. S1. HPLC chromatographic profile of EtOH extract (**E1a**) at 254 (a), 280 (b) and 350 nm (c).

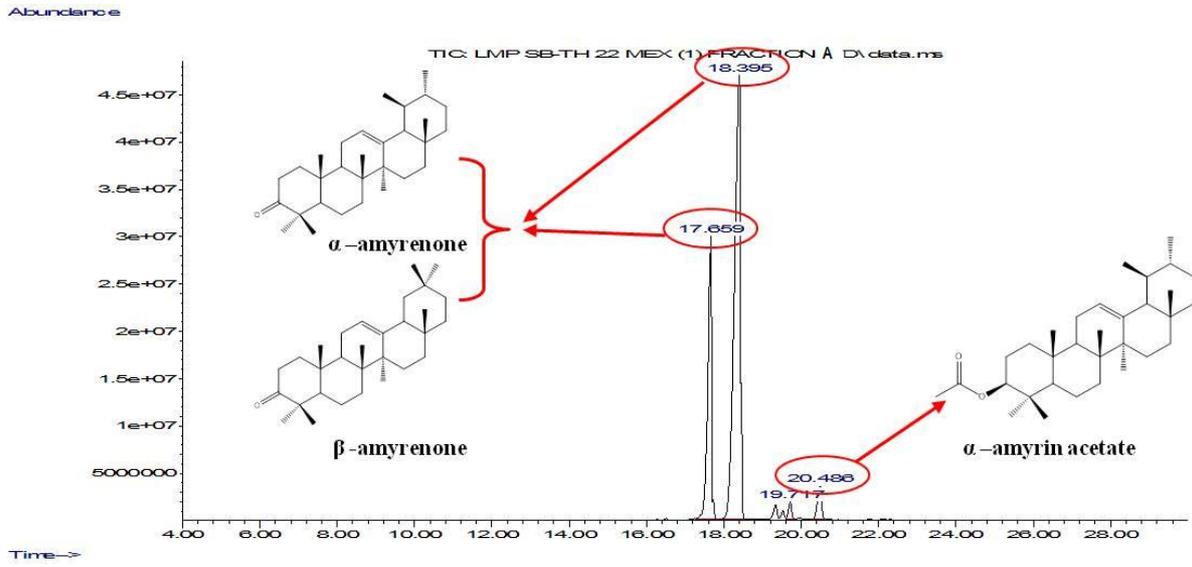


Fig. S2a. GC chromatographic profile of fraction A

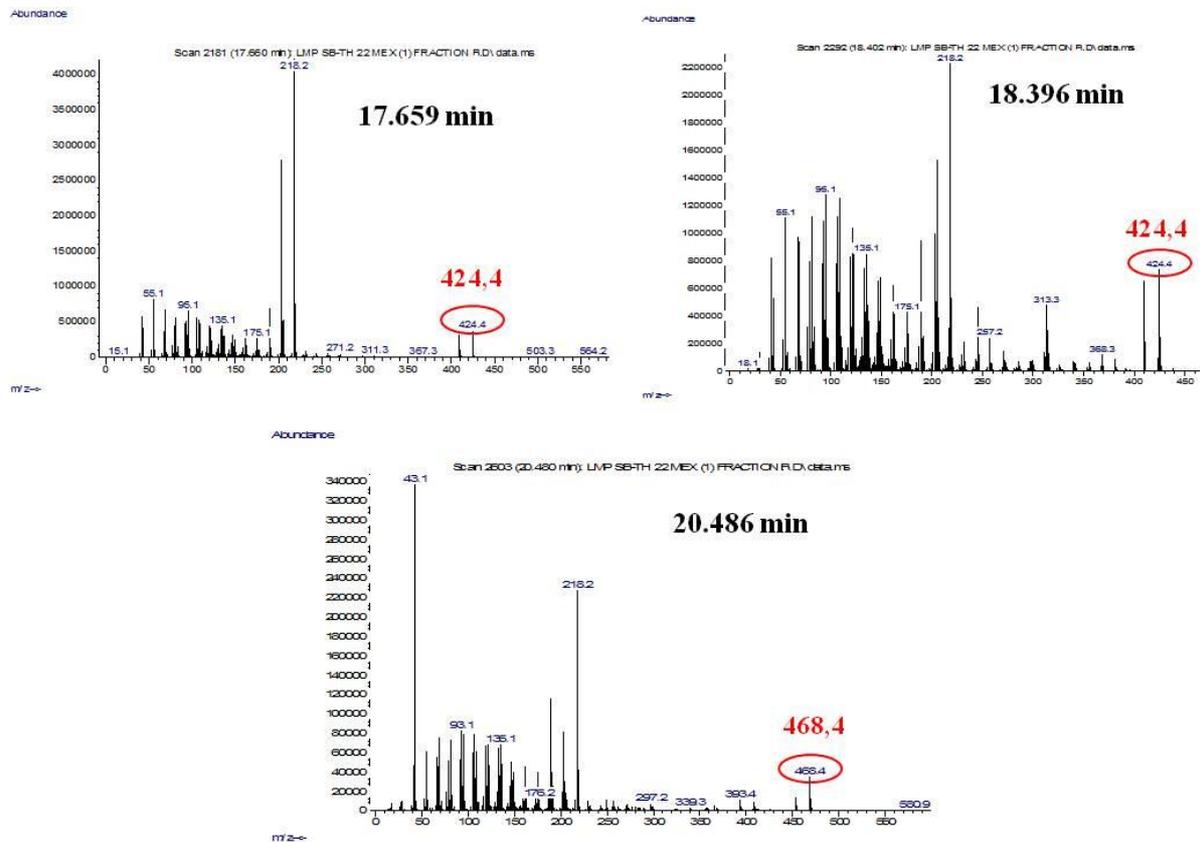


Fig. S2b. MS spectra of components in fraction A

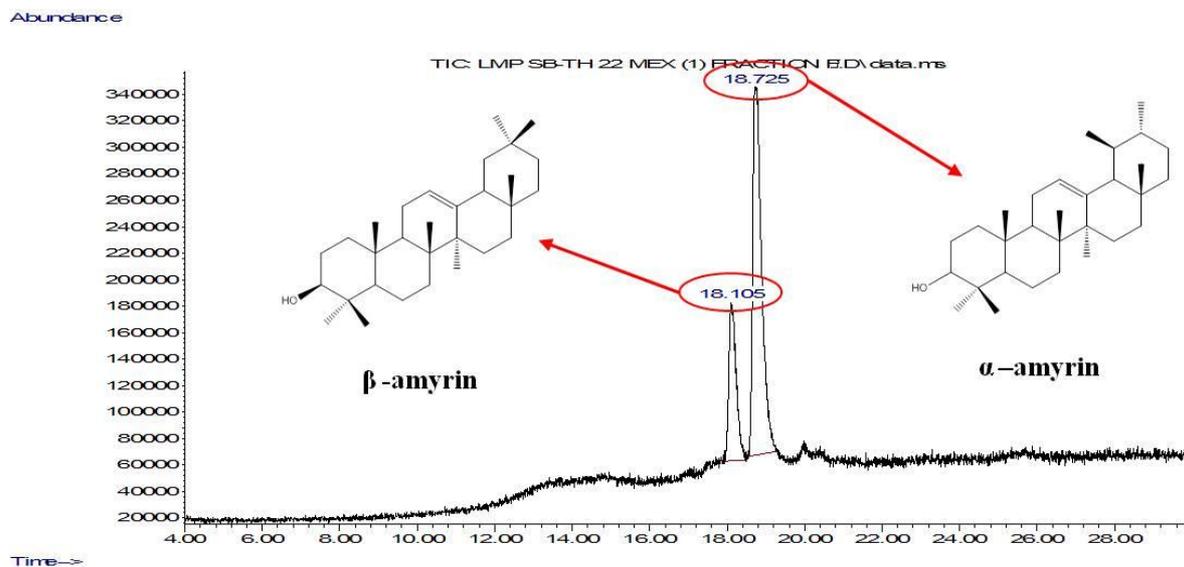


Fig. S3a. GC chromatographic profile of fraction B

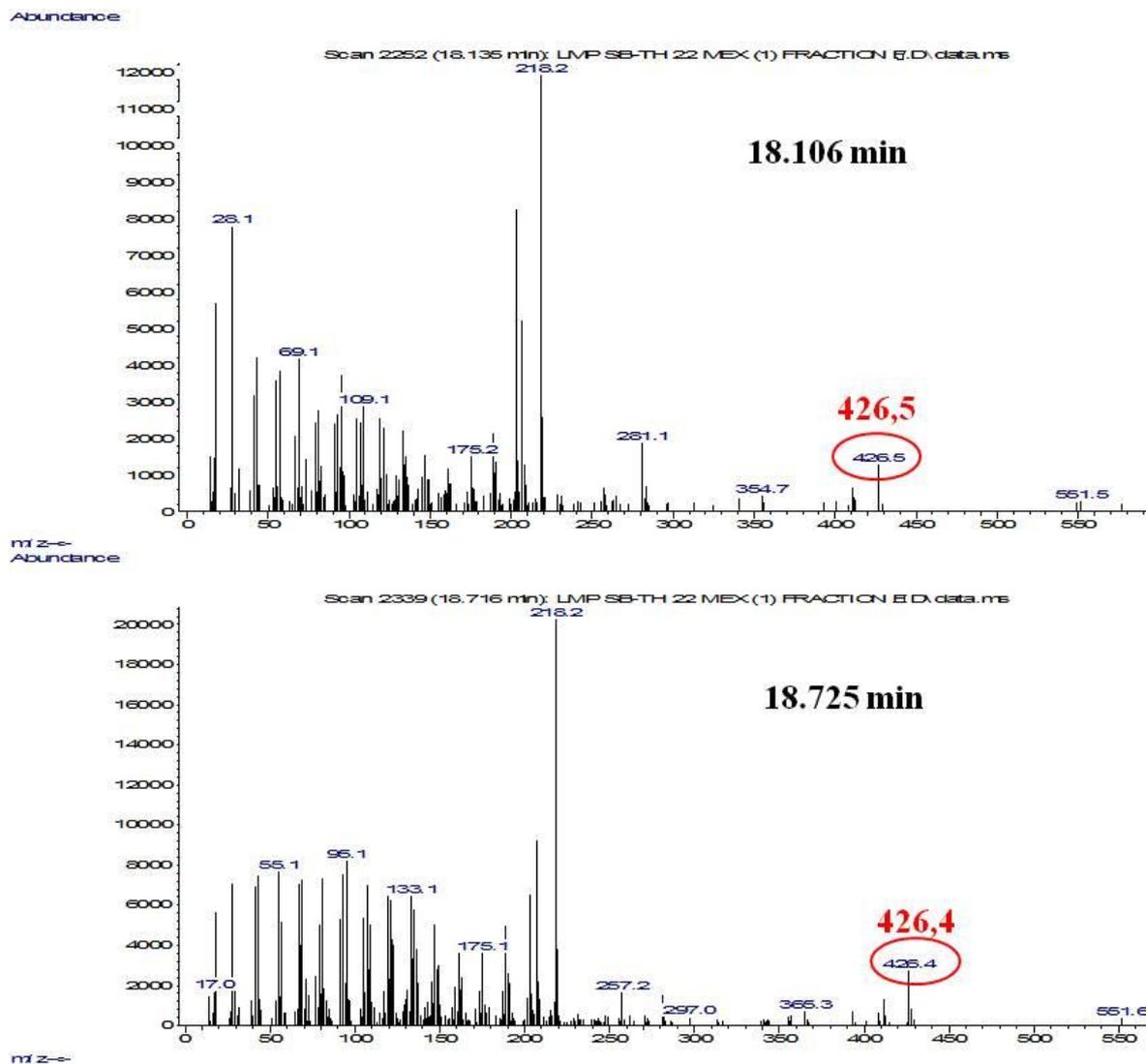


Fig. S3b. MS spectra of components in fraction B

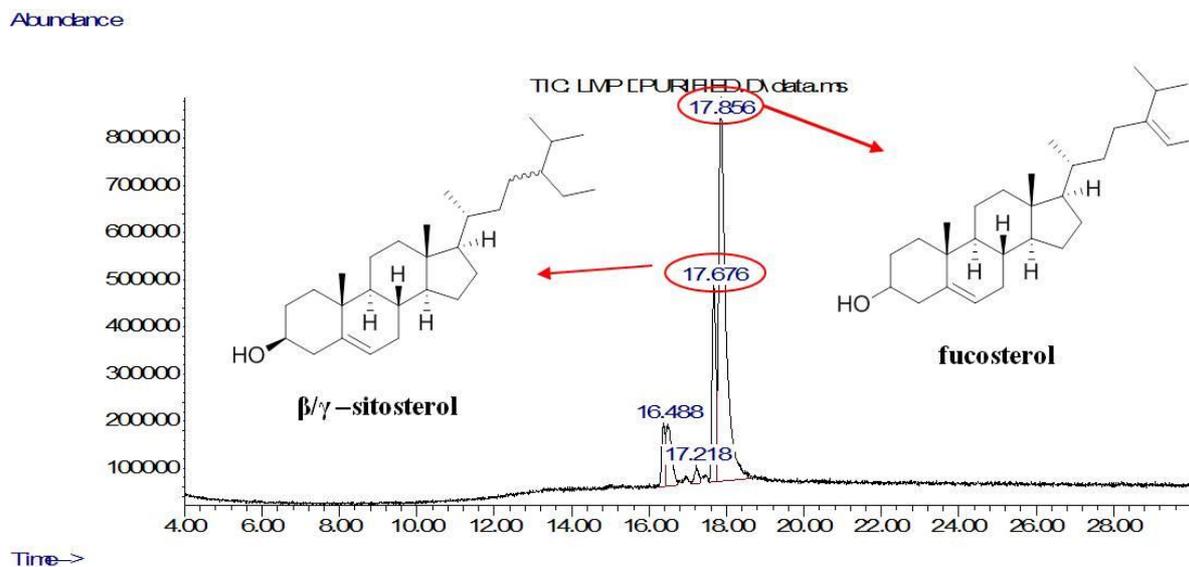


Fig. S4a. GC chromatographic profile of fraction C

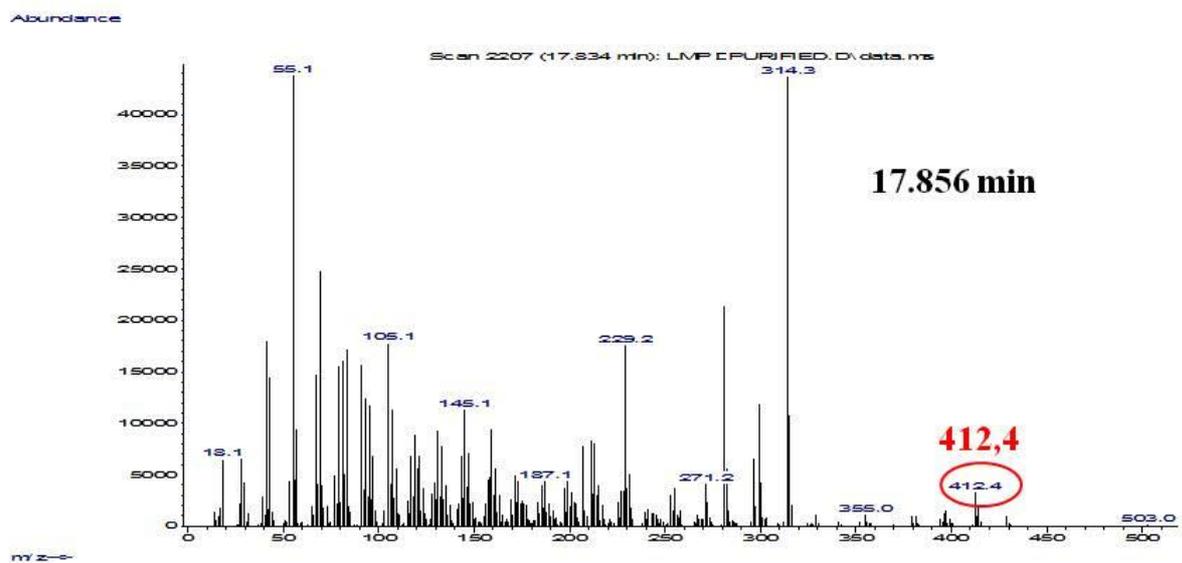
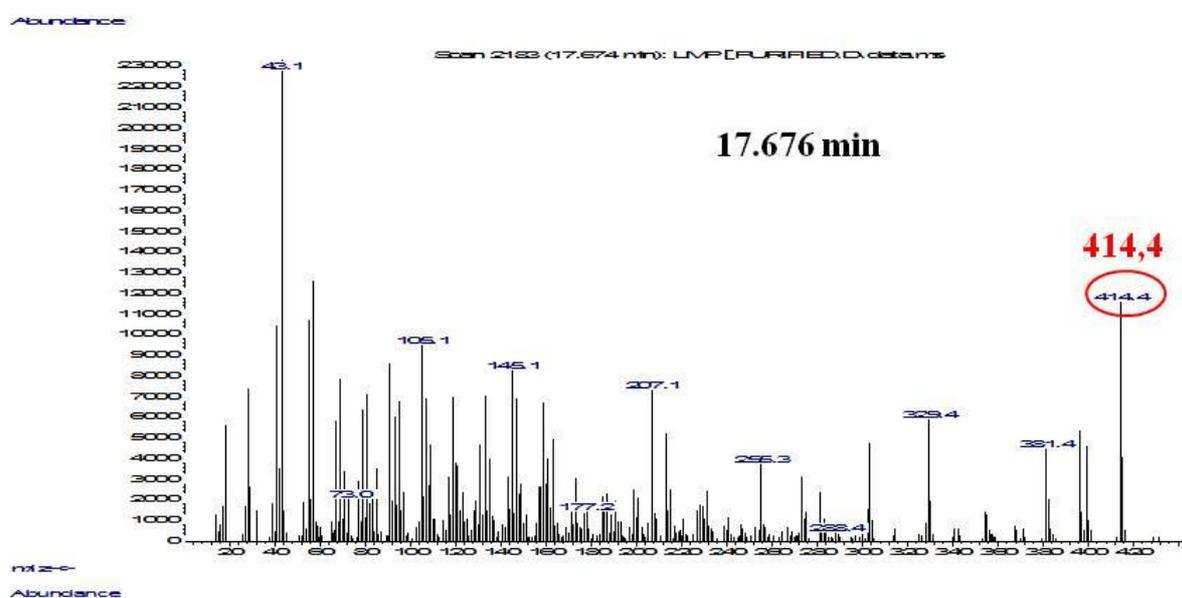


Fig. S4b. MS spectra of components in fraction C

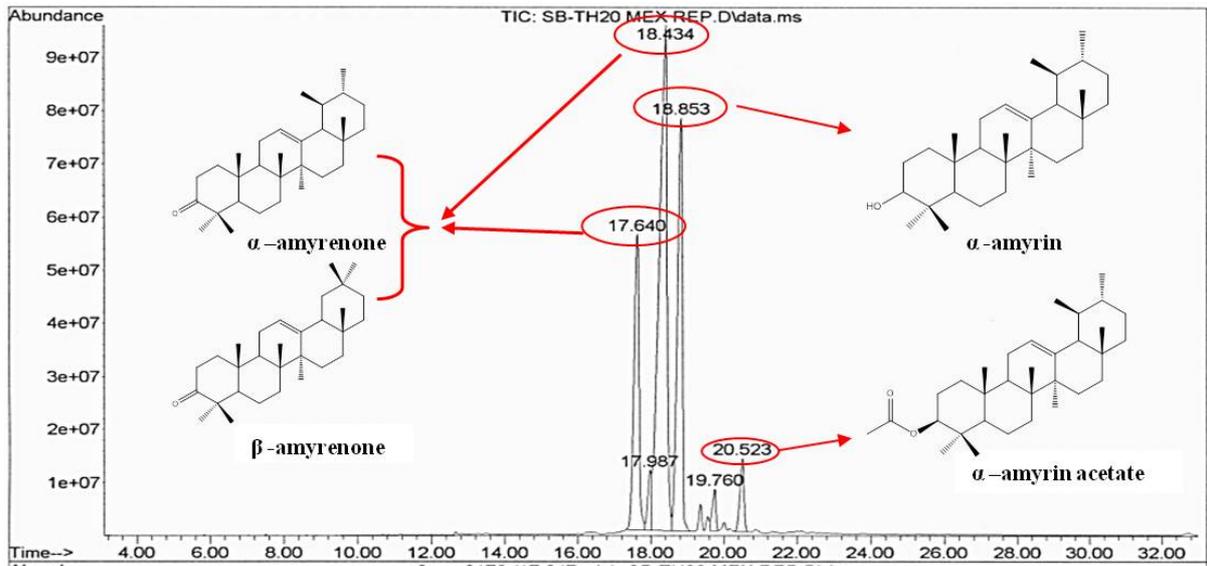


Fig. S5a. GC chromatographic profile of extract E1b

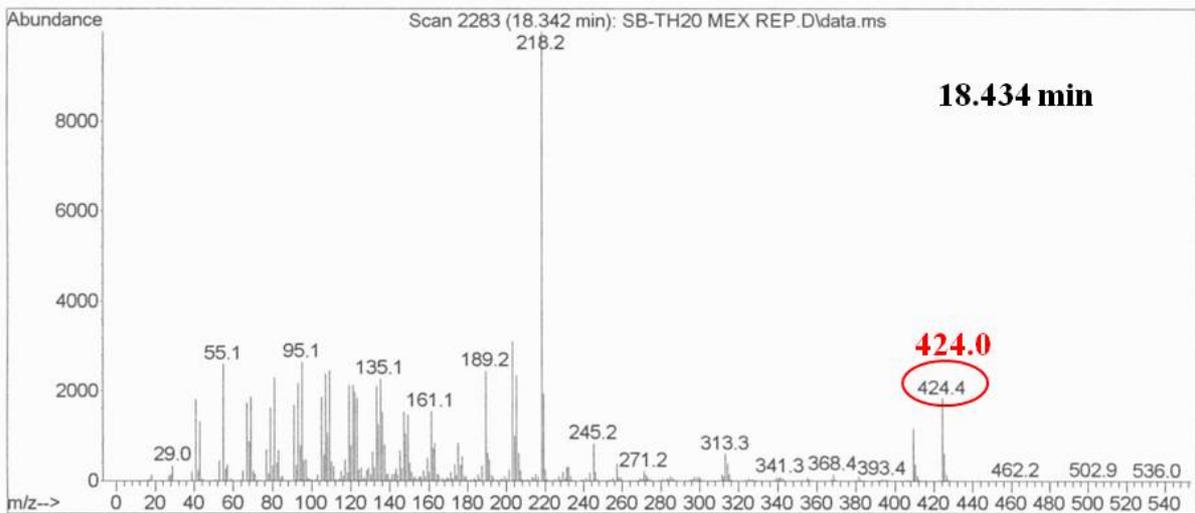
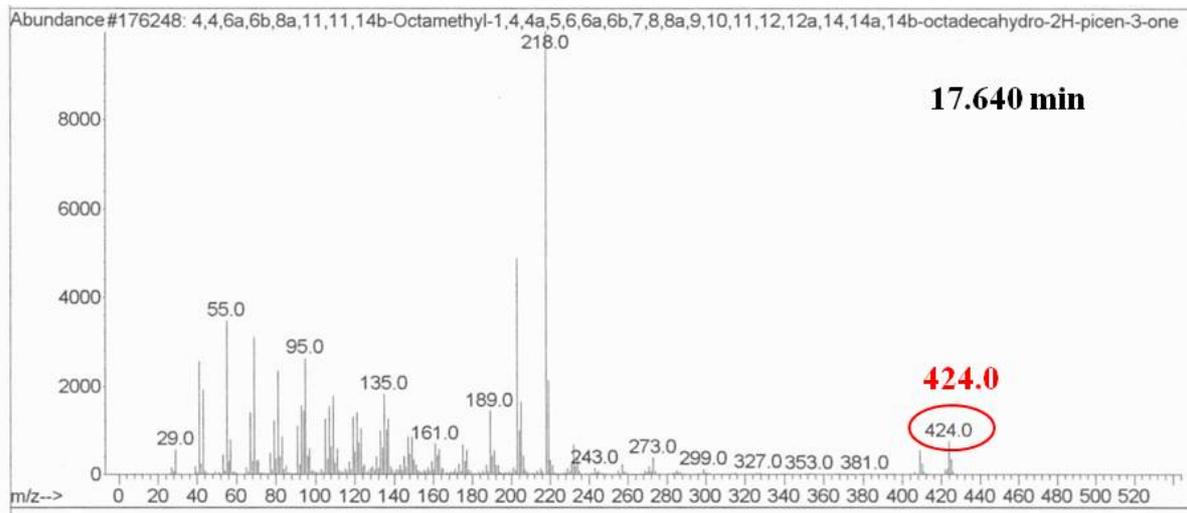


Fig. S5b-i). MS spectra of components in extract E1a (first part)

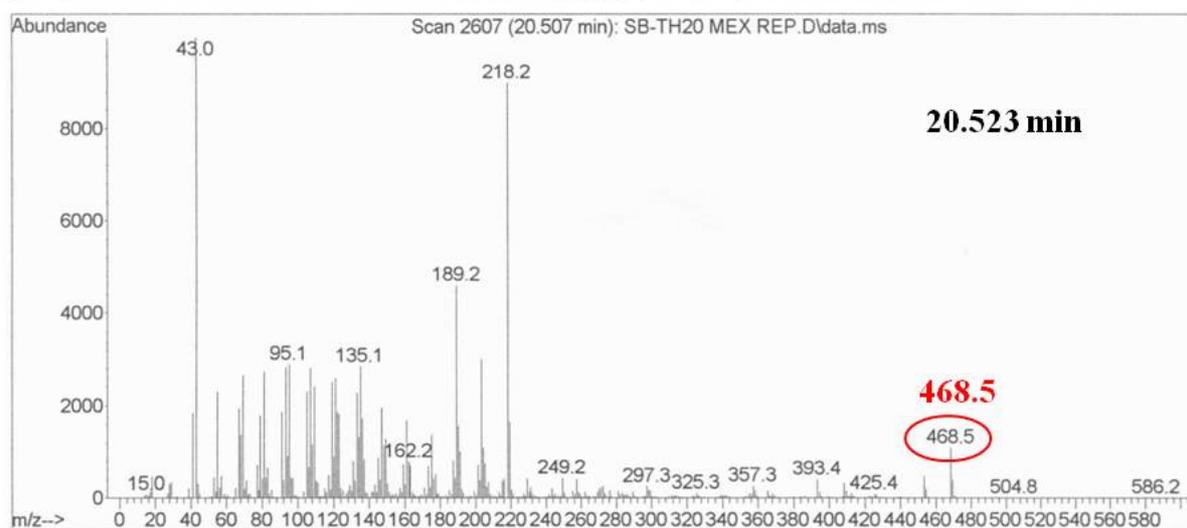
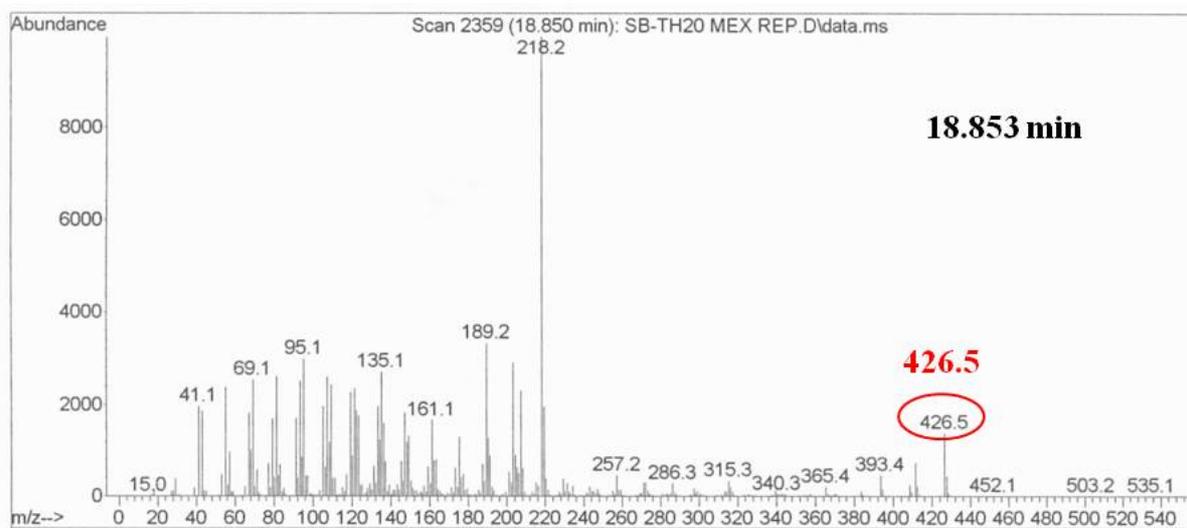


Fig. S5b-ii). MS spectra of components in extract E1a (second part)