

New frontiers in calcium stable isotope geochemistry: Perspectives in present and past vertebrate biology

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Abstract

 Beyond their established uses in Earth and Planetary sciences, calcium (Ca) isotopes have a promising future in the study of the biology of present and past vertebrates, including humans. Early work paved the way to the ongoing research on the potential of Ca isotopes as relevant tools to disciplines other than geology, including palaeobiology, bioarchaeology and biomedical research. In this article, we first review the rationale behind the cycling of Ca isotopes in vertebrate organisms. We then summarize and discuss the use of Ca isotopes as dietary tracers from trophic reconstructions in past vertebrate ecosystems to the tracking of early life dietary transitions. Next, we review and examine the research outcomes on the potential of Ca isotopes as biomarkers of bone loss in physiological and pathological conditions such as bone cancers and osteoporosis. While emphasizing the needs of future research in each of these applications, we suggest new potential uses of Ca isotopes in vertebrate biology. Finally, we identify challenges and barriers faced when developing such interdisciplinary projects and suggest how these can be overcome.

Keywords:

- Calcium isotopes,
- Vertebrate biology,
- Palaeobiology,
- Bioarchaeology,
- Biomedical research,
- Bone loss

Preliminary remarks

 Here, we shortly introduce the concepts of stable isotope geochemistry used in this review. *Isotopes* of a given element have nuclei made of a given number of protons but a varying number of neutrons. Depending on the number of neutrons, the nucleus of a given isotope is either stable over geological timescales or disintegrates into daughter nucleus or nuclei by radioactive decay. Isotopes are commonly discussed in scientific literature in connection to radiometric dating for instance in geology, archaeology or palaeontology, or as artificially prepared stable or radioactive nuclides used as tracers or as radiation sources in biomedical contexts for example. This paper is not concerned with any of these widely known applications, but with the study of variations in natural abundances of isotopes – referred to as *isotope compositions* – of elements with more than one naturally occurring isotope. The 52 isotopes of interest are either stable (e.g., here 40 Ca, 42 Ca, 43 Ca, 44 Ca and 46 Ca) or have half-53 lives so long that their instability can be ignored in most contexts (e.g., 48 Ca). The stable 54 isotope compositions are measured as abundance ratios (e.g., $44Ca^{42}Ca$ or $44Ca^{40}Ca$) with mass-spectrometers that allow separation and "counting" of accelerated ions according to their atomic masses (and charge). Stable isotope compositions are often expressed as deviations relative to isotope ratios measured in a reference material, by means of the 58 "**delta" notation** defined as follows (here for ⁴⁴Ca/⁴²Ca ratios):

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\delta^{44/42}Ca = \left(\frac{\left(\frac{44Ca}{A^2Ca}\right)_{sple}}{\left(\frac{44Ca}{A^2Ca}\right)_{std}} - 1\right) \times 1000
$$

59 where ⁴⁴Ca/⁴²Ca refer to abundance ratios of sample (*sple*) or standard (*std. i.e.* SRM915a in 60 this article), and $\delta^{44/42}$ Ca values are expressed in per mil (‰). Variations in the **stable** *isotope compositions* of an element result from differences in their mass and nuclear structure. Physical and chemical processes acting on these differences can "*fractionate*" isotopes, *i.e.* selectively partition isotopes between pools, for example between reactants

 and products in a chemical reaction. Isotope fractionation based on differences in mass between isotopes is referred to as "*mass dependent fractionation*". This implies that the amplitude of a mass dependent fractionation is a function of the mass difference between the considered isotopes. Isotope fractionation based on other differences, such as nuclear magnetic effects, is called "*mass independent fractionation*" (*e.g.,* Dauphas and Schauble 2016). All of the isotope effects discussed in this paper are mass dependent. This notably 70 implies that the $\delta^{44/40}$ Ca value of a given material is approximately twice its $\delta^{44/42}$ Ca value. Mass dependent isotope fractionation between freely exchanging pools at equilibrium, such as reactants and products in a reversible chemical reaction, is known as "*equilibrium isotope fractionation*" and are notably a function of temperature at Earth's surface. "*Kinetic isotope fractionation*" is associated with unidirectional incomplete processes, such as rapid change in phase. This paper deals with both equilibrium and kinetic effects. Readers wanting to further familiarize with concepts of metal stable isotope (bio-)geochemistry can also refer to several introductory reviews and book sections (*e.g.,* Albarède, 2015, Albarède et al., 2017, Martin et al., 2017a, Jaouen and Pons 2016, Wombacher et al., 2016).

1. Introduction

 81 Stable isotope geochemistry has a rich history of fruitful interdisciplinary collaboration, 82 resulting in the adoption of isotope geochemical techniques and concepts in fields such as 83 ecology, forensics and archaeology. Calcium (Ca) stable isotopes have been found useful in the earth and planetary sciences, for example in reconstructing secular variations in seawater composition (e.g., De La Rocha and DePaolo, 2000; Fantle and DePaolo, 2005, 2007; Farkaš et al., 2007a, 2007b; Hinojosa et al., 2012; Jost et al., 2014; Le Houedec et al., 2017; A. D. Schmitt et al., 2003). However, promising potential applications of calcium isotopes also lie outside of geology. Calcium isotopes have the potential to become an important tool in vertebrate biology and biomedicine, notably if challenges to their widespread application can be overcome.

20 100 The abundance and widely shared vital functions of Ca in vertebrate organisms makes it a crucial stable isotope system in the evolution and biology of vertebrates, including humans. The appearance in Cambrian marine vertebrates of calcium phosphate mineral tissues, i.e. bioapatite, marks the development of revolutionary strategies in the physiological regulation of Ca and inorganic phosphate, that are now widely shared among vertebrates (e.g., Bouillon and Suda, 2014; Doherty et al., 2015). Calcium phosphate minerals have a high preservation potential and are globally more resistant to diagenesis than carbonates. This has resulted in a rich vertebrate fossil record (e.g., Armstrong et al., 2001; Barham et al., 2012; Hinojosa et al., 2012; Joachimski et al., 2004; Kohn and Cerling, 2002; Luz et al., 1984; Pucéat et al., 2004; Wenzel et al., 2000).

22 101 102 $\frac{26}{27}$ 103 $\frac{28}{29}$ 104 31 105 33 106 $\frac{35}{26}$ 107 38 108 40 109 42 110 $\frac{46}{47}$ 112 49 113 51 114 115 $\frac{55}{56}$ 116 58 117 The pioneering work on Ca isotopes in biology was done by J.L. Skulan in the late 1990s (Skulan et al., 1997; Skulan and DePaolo, 1999; Skulan, 1999). Their key finding was that mineralized tissues, including bone, are significantly and constantly depleted in heavy isotopes of Ca when compared to Ca dietary sources. Following this early work, an increasing number of investigators have explored biological and palaeobiological applications of Ca stable isotopes. These explorations have proceeded in two main directions. The first aims at using Ca isotopes to reconstruct the dietary habits of present and extinct vertebrates in order to assess the trophic structures of past ecosystems, or the dietary behaviours of past human individuals and populations. This field of research (that could be referred to as Metal Stable Isotope Palaeobiology or Bioarchaeology) tackles questions relevant to palaeontology, (palaeo-)ecology, archaeology and anthropology. The second topic of ongoing research aims at developing the use of Ca isotopes to better understand and diagnose human metabolic bone disorders, such as bone loss induced by inactivity, microgravity or disease, including osteoporosis and cancers affecting bone. This field of research (that could be referred to as Stable Isotope Metallomics as proposed by Albarède (2015)) is at the crossroads between isotope geochemistry, fundamental physiopathology and applied biomedical research.

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2 120 4 121 $\frac{6}{9}$ 122 $\frac{8}{9}$ 123 11 124 13 125 $15/126$ 119 However, beyond the challenges inherent in extending fundamental understanding and developing practical applications, these projects also face difficulties that are common to all interdisciplinary research. More than 15 years have passed by since the first recognition of the potential of Ca isotopes in vertebrate biology, and yet only about 30 articles have been 123 published on the topic during this period (Figure 1). Calcium isotopes remain poorly utilized in scientific communities outside earth and planetary sciences. This raises the following question: what obstacles stand in the way of more widespread applications of Ca stable isotope analysis?

17 18 20 128 22 129 24 130 $\frac{26}{27}$ 131 In this article, we first briefly summarize the current understanding of the Ca isotope cycling in vertebrates before reviewing and discussing the potential and future of Ca isotopes in palaeobiology and biomedical research. Finally, we identify challenges and barriers related to the collaborative research necessary for the adoption of Ca isotope tools by the wider scientific community.

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133 **2. The cycling of calcium and its isotopes in vertebrates**

33 134 $\frac{35}{26}$ 135 38 136 40 137 42 138 The biological processing of Ca by vertebrates induces mass dependent isotope fractionations responsible for maintained and observable heterogeneous distributions of Ca isotopes amongst the different compartments of the organisms. Hence, the distribution and biological functions of Ca in vertebrates are central features of the potential of Ca stable isotopes as tools to study their biology.

- 44 $\frac{44}{45}$ 139
- $\frac{46}{47}$ 140

140 **2.1. Distribution, functions and regulation of Ca in vertebrates**

49 141 51 142 ⁵³ 143 $\frac{55}{56}$ 144 58 145 60 146 In vertebrate organisms, the distribution of $Ca²⁺$ is highly compartmentalized at all anatomical levels (e.g., Del Valle et al., 2011; Doherty et al., 2015; Peterson and Riggs, 2010). Briefly, most body Ca is stored in bone mineral as a major constituent of bioapatite $(Ca_{10}(PO_4)_6(OH)_2)$. Mineralized tissues account for ca. 99 % of body Ca in vertebrates, while the remaining fraction is found in non-mineralized or so-called soft tissues. The majority of soft tissue Ca (around 90 %) is found inside cells (mainly in organelles), while extracellular 2 148 4149 $\frac{6}{9}$ 150 147 fluids including blood contain about 10 % of the soft tissue Ca. However, the free Ca²⁺ of extracellular fluids is 10,000 to 20,000 times more concentrated than intracellular cytosolic $Ca²⁺$ (Case et al., 2007). This constitutes a steep and actively maintained chemical gradient between extracellular and intracellular milieus.

 $\frac{8}{9}$ 151 11 152 13 153 $15/154$ 17 18 20 156 22 157 24 158 $\frac{26}{27}$ 159 $\frac{28}{29}$ 160 The heterogeneous and tightly regulated distribution of Ca reflects the adaptation to the vital biological functions to which it contributes (see Berridge et al., 2000; Bootman, 2012; 153 Brini et al., 2013; Brown and MacLeod, 2001; Carafoli, 2002; Clapham, 1995). Calcium interacts with numerous proteins, either for storage and transport or for the regulation of protein activity. In multicellular organisms, $Ca²⁺$ is a ubiquitous intracellular second messenger regulating numerous electrical, chemical, mechanical and genetic cellular responses. From the cell to the whole organism, $Ca²⁺$ is a critical component in many vital functions, including cell differentiation or death, muscle contraction or propagation of electrical signals in some nerve cells. The preservation of these vital functions primarily depends on the fine regulation of Ca^{2+} cycle and on the blood Ca^{2+} concentrations (Figure 2).

31 161 32 33 162 34 $\frac{35}{26}$ 163 36 37 38 164 39 40 165 41 42 166 43 $^{44}_{45}$ 167 45 $\frac{46}{47}$ 168 47 48 49 169 50 51 170 52 53 171 54 $\frac{55}{56}$ 172 56 57 58 173 59 60 174 61 In terrestrial tetrapods, the principal source of Ca is the intestinal absorption of dietary Ca, while it is also exchanged with ambient water through gills in bony and cartilaginous fishes and through skin to a certain extent in amphibians (Bouillon and Suda, 2014; Doherty et al., 2015; Stiffler, 1993). Blood and extracellular fluids exchange Ca^{2+} with various nonexcreting reservoirs, such as muscle and more importantly with bone, which acts as a 166 dynamic storage organ, especially in terrestrial vertebrates (Bouillon and Suda, 2014). Various sinks balance the incoming $Ca²⁺$ fluxes, with contrasting contributions depending on species and life stages. In mammals, skin, hair and sweat account for some of the $Ca²⁺$ losses but the main loss occurs via urinary excretions and digestive secretions through faeces in most vertebrates. Tissues playing the roles of Ca^{2+} dynamic stores (e.g., bone) and interfaces exchanging with the environment (e.g., gills, intestine, kidneys) participate to the 172 regulation of the whole organism Ca cycle, called Ca homeostasis (Bouillon and Suda, 2014; de Matos, 2008; Doherty et al., 2015; Flik and Verbost, 1993; Peacock, 2010; Stiffler, 1993). In humans, many physiopathological conditions are associated with disrupted Ca cycles.

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2 176 175 Chronic perturbation of Ca and phosphate homeostasis may lead to metabolic bone disease 176 (e.g., osteoporosis) or result from disease (e.g., cancers affecting bone, c.f. Peacock, 2010).

 4 177 $\frac{6}{9}$ 178 $\frac{8}{9}$ 179 11 180 13 181 $15/182$ In reproductive contexts, Ca^{2+} cycles are modified especially in female organisms. In mammals, pregnancy and lactation are associated with modified Ca homeostasis (Del Valle et al., 2011; Doherty et al., 2015; Kalkwarf, 1999; Kovacs and Fuleihan, 2006). For instance, a significant Ca amount is lost to the foetus skeleton during pregnancy, while breastfeeding involves significant Ca daily output. Vertebrates that lay eggs with heavily mineralized shells 182 may also lose significant fractions of Ca via secretion and formation of shell Ca carbonate as well as yolk and albumen.

20 184 22 185 24 186 To conclude, as a major bio-essential metal in vertebrates, calcium lies at the crossroads of important scientific questions relevant to the evolution of biology and ecology of present and past vertebrates as well as to crucial modern health issues in humans.

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 $\frac{28}{29}$ 188

188 **2.2. Cycling of Ca isotopes in vertebrates**

31 189 33 190 $\frac{35}{26}$ 191 38 192 40 193 42 194 $^{44}_{.5}$ 195 $\frac{46}{47}$ 196 49 197 51 198 ⁵³ 199 $\frac{55}{56}$ 200 58 201 The main and first described feature of the Ca isotopes cycle in bony vertebrates is the significantly 44 Ca-depleted isotope composition of bone when compared to dietary Ca (Skulan and DePaolo, 1999). Compilation of literature data in 6 species of mammals and one bird (Figure 3) shows that Ca of bone systematically displays lower $\delta^{44/42}$ Ca values than dietary Ca, by about -0.57 ± 0.10 ‰ (2SE, ~ -1.14‰ in $\delta^{44/40}$ Ca) (Chu et al., 2006; Heuser et 194 al., 2016; Hirata et al., 2008; Skulan and DePaolo, 1999; Tacail et al., 2014). Despite the various represented species in literature, no significant relationship between the extent of this isotope effect and the organism physiology has been identified yet. A diet-bone isotopic offset is well conserved within amniotes, suggesting a phylogenetically shared mechanism resulting in Ca isotope fractionation during biological processing. In teleost and elasmobranch fishes, the amplitude of the offset between diet and mineralized tissues remains to be more thoroughly explored in controlled conditions but is possibly not as marked. Seawater ingestion and osmoregulation involving seawater filtering through gills

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2 2 0 3 202 could possibly attenuate or buffer these effects, as suggested by studies of marine ecosystems (Clementz et al., 2003; Martin et al., 2015).

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 4204 $\frac{6}{9}$ 205 $\frac{8}{9}$ 206 11 207 13 208 $15/209$ 17 18 20 211 22 212 24 213 According to the comparison of soft tissue and blood Ca with bone of the same animals (Skulan and DePaolo, 1999), this 44 Ca-depleted composition of bone primarily results from a major isotope fractionation occurring during mineralisation of bone. On the contrary, Ca loss from bone, during bone remodelling for instance, does not fractionate Ca isotopes. The amplitude of the mineralization isotope effect is thought to be the same as the diet-bone difference (Channon et al., 2015; Heuser and Eisenhauer, 2010; Morgan et al., 2012; Reynard et al., 2010; Skulan and DePaolo, 1999; Skulan et al., 2007), i.e. -0.57‰, although more recent comparison of bone and blood of sheep (Tacail et al., 2014) and pig (Heuser et al., 2016) suggest that fractionation during mineralisation could be less pronounced.

 $\frac{26}{27}$ 214 27 28 29 215 30 31 216 32 33217 34 $\frac{35}{36}$ 218 36 37 38 219 39 40 220 41 42 221 43 $^{44}_{45}$ 222 45 The processing of Ca by kidneys also induces a significant isotope effect, as observed in various mammals. The Ca excreted in urine is systematically enriched in heavy isotopes when compared to blood Ca, by ca. +1.2‰ in $\delta^{44/42}$ Ca (ca. +2.4‰ in $\delta^{44/40}$ Ca), in 217 human (Channon et al., 2015; Eisenhauer et al., 2019; Heuser and Eisenhauer, 2010; Skulan et al., 2007), sheep (Tacail et al., 2014) and pig (Heuser et al., 2016; Morgan et al., 219 2012). The relationship between the amount of excreted urinary Ca and its isotope composition in human suggests a Rayleigh isotope distillation process resulting from the preferential reabsorption of Ca light isotopes from primary urine to blood along the nephron (Heuser et al., 2019; Heuser and Eisenhauer, 2010; Morgan et al., 2012).

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49 224 51 225 $\frac{53}{1}$ 226 56 227 58 228 60 229 Vertebrate reproduction is also associated with significant Ca isotope effects. In mammals, the isotope composition of milk was found to be remarkably ⁴⁴ Ca-depleted in comparison with that of the mother's diet (Chu et al., 2006). The amplitude of this difference in $\delta^{44/42}$ Ca values averages at around -0.6‰ from diet to milk as observed in milk of ewes, cows and humans (Chu et al., 2006; Gussone and Heuser, 2016; Heuser, 2016; Tacail, 2017). Prolonged milk production by lactating females could also result in a 44 Ca-enriched

 1 2 2 3 1 3 4232 5 $\frac{6}{9}$ 233 7 $\frac{8}{9}$ 234 9 10 11 235 12 13 236 14 $^{15}_{16}$ 237 16 $\frac{17}{18}$ 238 18 19 20 239 230 composition of their bone when compared to males, as suggested by observations in modern 231 sheep (Reynard et al., 2010). In birds, despite limited datasets, Ca isotope compositions of eggs inner parts and carbonate shells were found to display remarkably extreme isotopic patterns (see Skulan and DePaolo, 1999 and Figure 5). While the isotopic composition of eggshell appears to be comparable to diet and significantly enriched in 44 Ca in comparison with bone of egg-laying hens, white albumen, and yolk to a lesser extent, display compositions that are even more enriched in heavy isotopes than for eggshell and diet. Such a 44 Ca-enriched composition of eggshells likely relates to the high demand in Ca for its formation, resulting in high fluxes of poorly fractionated diet-like Ca to the eggshell (Skulan et al., 1997).

22 240 23 24 241 25 This understanding of the Ca isotope cycle led to two main hypotheses that have been since then further tested and exploited:

 $^{26}_{27}$ 242 $\frac{28}{29}$ 243 29 31 244 33 245 $\frac{35}{26}$ 246 36 37 38 242 (i) as diet directly influences the systematically shifted isotope compositions of mineralized tissues, Ca isotopes can be used to reconstruct dietary behaviours in past vertebrates (ii) as Ca isotopes are fractionated during bone mineralisation, Ca isotope compositions of 245 blood and urine vary with the balance between bone formation and resorption. Calcium isotopes could be used to monitor variations of bone mineral balance in human subjects suffering from bone disorders.

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249 **2.3. Future research**

 $^{44}_{45}$ 250 $\frac{46}{47}$ 251 49 252 51 253 New research is needed in order to i) further characterize the distribution of Ca isotopes in the various reservoirs of vertebrate organisms, ii) identify factors of variability (e.g., species-specific physiological effects) and iii) improve our understanding of the mechanisms responsible for the major biologically induced isotope fractionations.

 53254 $\frac{55}{56}$ 255 58 256 60 257 These remaining aspects would benefit from various approaches. At the scale of the whole organism, the results of controlled feeding experiments done on species from various clades of vertebrates could help further identify physiological factors of variability. This type 257 of approach would help refine the identification of Ca fluxes associated with isotope effects

2 2 5 9 4260 $\frac{6}{2}$ 261 $\frac{8}{9}$ 262 11 263 13 264 $15/265$ $\frac{17}{18}$ 266 20 267 22 268 24 269 $^{26}_{27}$ 270 258 potentially affecting the whole organism Ca isotope distribution (e.g., Balter et al., 2013). Isotope box modelling of whole organism at steady-state and in dynamic conditions will then allow us to test the proposed mechanisms and investigate the potential of Ca isotopes for identified and future applications (e.g., Jaouen et al., 2019). Various physiological conditions could notably be explored, such as the age-related changes in the balance between bone anabolism and catabolism or the cycling of Ca isotopes between pregnant mammals and 264 their foetus. In order to better understand the cellular and molecular mechanisms at play, it will also be possible to compare the results of experimental approaches on cellular in vitro models with controlled expression of cross-membrane protein transporters (e.g., Cadiou et 267 al., 2017) and estimations of molecular equilibrium fractionation with numerical *ab initio* calculations (e.g., Moynier and Fujii, 2017). All these approaches will greatly benefit both palaeobiology and modern biomedical applications and potentially further reveal other yet to be identified applications.

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272 **3. Calcium isotopes in palaeobiology and bioarchaeology**

33 273 34 $\frac{35}{26}$ 274 36 37 38 275 39 40 276 41 42 277 43 $^{44}_{45}$ 278 45 $\frac{46}{47}$ 279 47 48 49 280 50 51 281 52 53 282 54 $\frac{55}{56}$ 283 56 57 58 284 59 60 285 61 Stable isotopes and trace elements enable the development of quantitative approaches to reconstruct ecological, physiological and environmental characteristics that 275 otherwise remain unseen. Unravelling the partitioning of resources notably helps to better 276 understand past and present ecosystem dynamics and biodiversity. Geochemical proxies are useful means of complementing inferences of morphofunctional and microwear analyses of osteological remains (e.g., Martin et al., 2017a). The same applies to the study of past human societies, where such tools are valuable to complement the conclusions of archaeology and anthropology (Jaouen, 2018; Jaouen and Pons, 2016). Following the example of other more established proxies, such as collagen carbon and nitrogen stable isotopes or Sr/Ca and Ba/Ca elemental ratios (Balter and Lécuyer, 2004; Koch, 2007; 283 Newsome et al., 2010; Peek and Clementz, 2012), Ca stable isotopes of fossil bioapatite have a high potential for the study of past vertebrate ecosystems and human communities for three main reasons.

2 2 8 7 4288 6289 $\frac{8}{9}$ 290 11 291 13 292 $15/293$ 17 18 286 First, as discussed above, the Ca isotope composition of diet primarily determines the 287 composition of vertebrate mineralized tissues. This early finding came along with the hypothesis that such a trophic effect could propagate along food chains. The trophic level 289 index, ranging from primary producer to apex predators and decomposers, describes the 290 position of species within trophic chains and thus allows synthetizing the energetic pathways within ecosystems (Polis and Strong, 1996). Early datasets from both marine and terrestrial environments supported this trophic level effect (Clementz et al., 2003; DePaolo, 2004; Skulan et al., 1997; Skulan and DePaolo, 1999). The biological processing of Ca from a trophic level to another would thus lead to a stepwise decrease of the $\delta^{44/42}$ Ca values.

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29 299 29 30 31 300 32 333 301 34 $\frac{35}{36}$ 302 36 37 38 303 39 40 304 41 42305 43 $^{44}_{45}$ 306 45 46 47 307 48 49 308 50 Second, as a major constituent of a stable mineralogical phase (\sim 40 wt.% Ca of bioapatite), the post-mortem diagenetic processes tend not to alter skeletal bioapatite Ca and its isotope compositions especially in the denser and less porous tooth enamel (Heuser et 298 al., 2011; Melin et al., 2014; Martin et al., 2017; Martin et al., 2018). The determination of Ca isotope compositions in osteological remains makes it possible to explore deep past ecosystems. So far, studies of vertebrate fossils ecology with Ca isotopes focused on assemblages dating back to Late Devonian (Balter et al., 2019), Triassic to Cretaceous 302 (Hassler et al., 2018; Heuser et al., 2011; Martin et al., 2017b) or Pliocene and Pleistocene 303 (Martin et al., 2018, 2015, Tacail et al., 2019). Contrastingly, the conservation of pristine nitrogen isotope compositions of bone collagen primarily depends on the preservation of the 305 organic phase, the degradation of which rarely allows investigating on trophic relationships beyond Holocene or Late Pleistocene (Koch, 2007). On the other hand, the preservation of biogenic Sr/Ca and Ba/Ca trace element ratios strongly depends on burial conditions (e.g., Martin et al., 2018; Reynard and Balter, 2014).

51 309 52 $\frac{53}{54}$ 310 54 55 56 311 57 58 312 59 60 313 61 Finally, the abundance of Ca in osteological remains makes it possible to sample minute amounts of material (down to 20 µg or less). Such sample sizes allow studying precious fossils, without significantly altering their physical and structural integrity. It also motivates the development of high resolution sampling techniques involving mechanical sampling of tooth enamel by micro-drilling (Tacail et al., 2016, 2017) and laser cutting (Li et

 1 2 3 1 5 3 4316 5 $\frac{6}{9}$ 317 7 $\frac{8}{9}$ 318 9 314 al., 2016). *In situ* laser ablation isotope analyses were shown to be possible in tooth enamel 315 (Tacail et al., 2016; Zhang et al., 2019), although these approaches could greatly benefit from the implementation of technological innovations such as the currently developing use of 317 collision-cell multi-collector plasma mass spectrometry (*e.g.,* Lewis et al., 2018, Zhao and Simon 2019), with potential interference removal.

11 319 13 320 $\frac{15}{16}$ 321 $\frac{17}{18}$ 322 After the first assessments of the utility of Ca isotopes for reconstructions of vertebrate ecology, several studies further explored and developed strategies to reconstruct biological, ecological and behavioural characteristics of past vertebrates and human populations.

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324 **3.1. Calcium isotopes and vertebrate ecosystem reconstructions**

 24 325 $^{26}_{27}$ 326 $\frac{28}{29}$ 327 The sensitivity of skeletal Ca isotope compositions to trophic levels was explored and exploited in a series of modern and fossil ecosystems, both in marine and continental environments.

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329 **3.1.1. Marine ecosystems**

 $\frac{35}{36}$ 330 38 331 40 332 42 333 Seawater is the primary source of Ca in most marine ecosystems (\sim 420 mg/L, Elderfield and Schultz, 1996). The Ca isotope composition of seawater is particularly enriched in heavy isotopes (Figure 5) and relatively homogeneous at the scale of the oceans (e.g., Fantle and Tipper, 2014).

 $^{44}_{45}$ 334 45 $\frac{46}{47}$ 335 47 48 49 336 50 51 337 52 53 338 54 $\frac{55}{56}$ 339 56 58 340 59 60 341 61 Following the first description of decreasing $\delta^{44/42}$ Ca values with increasing trophic levels (Skulan et al., 1997), Clementz et al. (2003) reported consistent patterns in bones and teeth of both modern and fossil marine mammals. Species of low trophic levels (feeding on marine vegetation or algae and invertebrates) display ⁴⁴ Ca-enriched compositions when compared to higher trophic level mammals (feeding on fishes and other marine mammals). This pattern was also later reported in elasmobranch species of diverse ecologies (i.e. cartilaginous fishes, namely sharks and rays, see Martin et al., 2015). These findings were finally applied to a fossil marine ecosystem dating back to late Cretaceous (Martin et al.,

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2 343 4344 $6/345$ $\frac{8}{9}$ 346 11 347 342 2017b), composed of samples from actinopterygian and elasmobranch fishes, as well as Mosasaurid and Plesiosaurid large marine reptiles. This assemblage shortly predates the end-Cretaceous extinction and sheds light on the vulnerability of this ecosystem, where high trophic level large reptiles were relying on abundant yet poorly diversified resources. The use 346 of Ca isotopes also yielded new constraints onto the ecology and trophic level of extinct Devonian conodonts organisms for which no modern analogue is known (Balter et al., 2019).

13 348 14 15349 16 $\frac{17}{18}$ 350 18 19 20 351 21 22 352 23 24353 25 $^{26}_{27}$ 354 27 $\frac{28}{29}$ 355 29 30 31 356 32 33 357 34 $\frac{35}{26}$ 358 36 37 38 359 39 40 360 41 42 361 43 These studies identified features that require further dedicated investigation in order to improve the use of Ca isotopes as an indicator of marine ecosystem structures. For instance, data indicates that trophic ecology primarily determines tooth and bone isotope 351 compositions of marine vertebrates, but mammals and fishes (cartilaginous or ray-finned) do not appear to display the same average isotope shifts from one trophic level to another 353 (Clementz et al., 2003; Martin et al., 2017b, 2015). The ingestion of seawater and processing of Ca via gill-mediated osmoregulation in actinopterygian and elasmobranch species is suspected to buffer the diet-skeleton isotope offset. On the other hand, marine mammal organisms are rather closed systems with respect to seawater. The differences in Ca isotope 357 physiology of fishes and mammals should thus motivate dedicated research in order to decipher the physiological and ecological controls on the Ca isotope compositions of their tissues. A better understanding of the isotope systematics of marine mammals would notably be of importance as their ecology makes them useful sentinels of ecosystem changes (e.g., Moore, 2008).

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363 **3.1.2. Continental ecosystems**

49 364 51 365 ⁵³ 366 $\frac{55}{56}$ 367 58 368 60 369 The nature and isotope compositions of primary Ca dietary sources of most terrestrial vertebrate trophic chains are more diversified than in marine ecosystems (see Figure 5). Indeed, in addition to the possible variability of isotope compositions of environmental Ca 367 (e.g., interacting bedrock, soil and water), the plant Ca isotope compositions are highly variable between types, species and organs (e.g., Schmitt, 2016 and Figure 5). Depending on the ecological niches of primary consumers (e.g., in vertebrate herbivores), such 2 371 370 variability can propagate throughout the trophic chain and make the interpretation of Ca isotope compositions more complex for trophic relationships reconstructions.

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4 372 5 $\frac{6}{9}$ 373 7 $\frac{8}{9}$ 374 9 10 11 375 12 13 376 14 $\frac{15}{16}$ 377 16 $\frac{17}{18}$ 378 18 19 20 379 21 22 380 23 24381 25 $^{26}_{27}$ 382 27 $\frac{28}{29}$ 383 29 30 31 384 32 33 385 34 $\frac{35}{26}$ 386 36 37 387 39 40 388 41 ⁴² 389 43 $^{44}_{45}$ 390 45 $\frac{46}{47}$ 391 47 48 49 392 50 $\frac{51}{2}$ 393 52 53 54 394 55 56 395 57 58 396 59 Melin et al. (2014) explored the bone Ca isotope compositions of various mammals including primates from two tropical forested ecosystems. The authors confirmed the decrease of Ca isotope ratios in large carnivores, but found no significant differences between lower trophic levels, notably small faunivores and insectivores. They however suggest an encouraging complementary use of carbon (C) and Ca stable isotopes. Martin et al. (2017a) reported encouraging bone and tooth enamel Ca isotope compositions of two Western European Pleistocene mammal assemblages. The stable isotope compositions of each taxonomic group (up to 6 individuals) are rather clustered and carnivores tend to display lower $\delta^{44/42}$ Ca values than herbivores. However, the outlying compositions of some species remain difficult to interpret in terms of trophic inferences, suggesting either speciesdependent physiological effects or peculiar dietary niches effects. Recently, Martin et al. (2018) reported the results of the systematic study of mammal tooth enamel of modern and fossil Pliocene-Pleistocene ecosystems from East Africa. This study further documents the potential of Ca isotopes as helpful markers for trophic inferences. The combination of C and Ca stable isotopes helps to distinguish between C_3 and C_4 trophic chains structures, in extant and extinct ecosystems. Indeed, the carbon stable isotope compositions of the enamel of mammals (δ^{13} C) are characteristic of the photosynthetic pathways of the plants at the base of the trophic chain (C_3 or C_4 carbon fixing sugars, c.f. Bender 1971), while Ca isotopes appear to allow discriminating between herbivores and carnivores. This is exemplified by the figure 4A, where the carnivores appear depleted in 44 Ca isotopes with respect to their respective inferred preys, being either predominantly C_3 herbivores with low $\delta^{13}C$ values (namely browsers) or a mix between the latter and C_4 herbivores with higher δ^{13} C values (namely grazers). Furthermore, when taking into account all available geochemical proxies relevant to feeding and habitat ecology (namely Sr/Ca and Ba/Ca ratios and $δ¹³C$, $δ¹⁸O$, $δ^{44/42}Ca$, see Martin et al., 2018), a principal component analysis (Figure 4B) reveals an inferred

2 398 4 399 397 ecosystem structure that is very similar to what δ^{13} C and $\delta^{44/42}$ Ca alone suggest (Figure 4A). In other words, the coupling of δ^{13} C and $\delta^{44/42}$ Ca appears to efficiently summarize the ecosystem structure.

 6^{6} 400 $\frac{8}{9}$ 401 11 402 13 403 15 404 Finally, in the Mesozoic fossil record, despite the fact that rather inconclusive results were obtained from dinosaur faunas on large geographical and chronological scales (Heuser et al., 2011), the systematic study of skeletal remains Ca isotope compositions from a fossil assemblage at regional scale brings new constraints to the resource partitioning between 404 mid-Cretaceous predatory dinosaurs and more precisely to the ecology of spinosaurids (Hassler et al., 2018).

20 406 22 407 24 408 $^{26}_{27}$ 409 29 410 31 411 33 412 $\frac{35}{26}$ 413 38 414 40 415 42 416 44 45 46 47 49 419 51 420 53 $\frac{55}{56}$ 422 58 423 406 To conclude, the ongoing exploration at regional scale of the ecological controls over vertebrate Ca isotope compositions in the continental environment is encouraging. Calcium isotopes show a very good potential for the reconstruction of dietary relationships in past vertebrate ecosystems, from Palaeozoic to Quaternary, and allow constraining local ecosystem structures and resource partitioning, especially between herbivores and carnivores. These results motivate the development of new strategies that notably include i) the combination of $\delta^{44/42}$ Ca with complementary proxies, such as δ^{13} C, ii) the study of assemblages composed of taxa with diverse ecologies, iii) at a regional scale, iv) with taxonomic groups comprising multiple individuals. Further research needs to be carried out, notably because some taxa do not entirely fit within this interpretative framework. This could 416 relate to peculiar physiological adaptations or ecological niches (e.g., Martin et al., 2018, 2017a). For instance, the observed isotopic offsets between bone of most secondary or tertiary consumers and their vertebrate preys are likely to primarily rely on the partial consumption of mineralized tissues from the latter. The intake of only 1 wt. % bone in total diet is sufficient to cause a significant trophic effect in Ca isotope compositions (-0.50‰ on the $\delta^{44/42}$ Ca scale) of consumer's tissues in terrestrial vertebrates (Heuser et al., 2011). Although little is known about muscle and bulk soft tissues isotope compositions, the consumption of energy-rich but Ca-poor soft tissues could result in different isotope offsets

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2 4 2 5 424 between secondary consumers and their vertebrate preys, making trophic level inferences less straightforward.

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427 **3.1.3. Future directions**

(i.) The development of statistical approaches for diet reconstruction with Ca isotopes could help to refine and make the most of trophic ecosystem reconstructions (such as Bayesian statistical inferences, see for instance Fernandes et al., 2014). The inter-individual variability of Ca isotope compositions within a given taxonomic group could also relate to the degrees of specialisation for various Ca resources (e.g., Martin et al., 2018). By means of statistical methods, Ca isotopes could allow characterizing "isotope niches" and help to document 434 breadths of ecological niches or the specialisation of taxonomic groups (*e.g.,* Yeakel et al., 2015).

 $^{26}_{27}$ 436 $\frac{28}{29}$ 437 31 438 33 439 $\frac{35}{26}$ 440 38 441 40 442 42 443 44 444 *(ii.)* Profound changes in the biological management of Ca occurred in the course of vertebrate evolution. A series of major evolutionary steps could potentially be studied with the help of Ca isotopes: the appearance of calcium phosphate mineralized endoskeletons, the ecological and physiological adaptations related to fish-to-tetrapod transitions followed by terrestrialisation, the development of egg-laying and carbonated shells in vertebrates or finally the development of pregnancy and lactation in first mammalians (Bouillon and Suda, 2014; Doherty et al., 2015). Each of these evolutions of vertebrate physiology and ecology likely affected their Ca isotopes cycling, which could in turn be exploited in order to document the evolutionary mechanisms at play.

49 446 51 447 ⁵³ 448 *(iii.)* Calcium isotope might be useful in other practical applications such as in food forensics. 446 For instance, combined with other stable isotopes, Ca isotopes could help in tracking the consumption of animal meat and bone meals by farmed animals (e.g., cattle, poultry; see Carrijo et al., 2006).

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450 **3.2. Calcium isotopes and dietary reconstructions in hominids and human**

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452 **3.2.1. From ecosystems to human societies**

2 453 4454 $\frac{6}{9}$ 455 $\frac{8}{9}$ 456 11 457 13 458 $15/15$ 459 17 18 20 461 22 462 24 463 Major changes in dietary practices in the course of human evolution, being ecologically and/or culturally driven, likely came along with significant changes of their use of Ca 455 resources (e.g., Eaton and Nelson, 1991). The changing dietary habits of early *Homo* or the development of agriculture and livestock farming in the course of Neolithic mark significant modifications of the use of environmental Ca and were associated with major ecological, 458 physiological, genetic or cultural evolutions (e.g., Aiello and Wheeler, 1995; Hublin et al., 2015; Ségurel and Bon, 2017; Vigne, 2011). The utility of Ca isotopes to track these changes has been however poorly explored. While the previously described ecosystem approach using Ca isotopes could help to interpret the evolution of hominid ecologies, the study of bone Ca isotopes in human societies could improve our understanding of their contrasting dietary habits influenced by varied cultural practices.

 $^{26}_{27}$ 464 27 $\frac{28}{29}$ 465 29 30 31 466 32 ³³ 467 34 35 36468 As shown in Figure 5, the major dietary sources of Ca for human have significantly contrasting isotope compositions. Therefore, the culturally driven practices of consumption of particular Ca dietary sources could lead to distinctive distributions of bone $\delta^{44/42}$ Ca values in human populations. This could be of use to reconstruct dietary practices of past human societies and characterize their subsistence modes.

38 469 39 40 470 41 42 471 43 $^{44}_{45}$ 472 45 46 473 48 49 474 50 51 475 52 $\frac{53}{1}$ 476 54 55 56 477 57 58 478 59 ⁶⁰ 479 61 For instance, dairy products constitute the most 44 Ca-depleted end-member of this isotopic landscape (with modern human milk being the lightest), while eggs and marine resources display the most ⁴⁴ Ca-enriched compositions. Following these observations, Chu et al. (2006) proposed that Ca isotopes could be used to reconstruct the consumption of dairy products in past human groups, especially during the processes of Neolithization with domestication of dairy producing animals. Indeed, the consumption of significant proportions of Ca from dairy products should induce a global decrease in $\delta^{44/42}$ Ca values of dietary Ca and thus in bone of human consumers. Reynard et al. (2010, 2011) tested the potential of Ca isotopes in faunal and human bones as a biomarker of dairy product consumption in several archaeological populations before and after domestication of dairy producing animals. These authors found however no significant change in the $\delta^{44/42}$ Ca offset between fauna and human

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2 481 480 of the studied archaeological sites after the domestication of milk producing animals. They suggested that controls other than animal milk consumption were likely at play.

Nevertheless, further research including actualistic calibrations in human osteological remains could help refine our understanding of the controls of dietary practices over bone Ca isotope compositions.

486 **3.2.2. Life history reconstructions in individuals**

Throughout their development and life, humans experience major adaptations of their Ca cycle both in terms of nutrition (e.g., birth or weaning in offspring) or Ca homeostasis (e.g., pregnancy or breastfeeding in females) (Humphrey, 2010; Kovacs and Fuleihan, 2006). The characteristics of these events (e.g., timing, frequency) are crucial and informative traits of the life history of individuals, relating to the ecological strategies of species and to cultural behaviours (Kramer and Otarola-Castillo, 2015; Lee, 2012; Robson and Wood, 2008). Such changes can in turn influence the Ca isotope distributions, which could be exploited if archived in human remains.

33 495 $\frac{35}{26}$ 496 38 497 40 498 42 499 $^{44}_{45}$ 500 $\frac{46}{47}$ 501 49 502 51 503 53 504 $\frac{55}{56}$ 505 58 506 60 507 So far, the major life history traits explored with Ca isotopes relate to mother's milk consumption by children. The biologically and culturally driven weaning behaviours are central traits of the evolution of mammals and hominins (Humphrey, 2010; Sellen, 2007; Van Noordwijk et al., 2013) but these practices remain fairly undocumented in the deep past because biomarkers are lacking. As milk was early recognized as a ⁴⁴Ca-depleted material in comparison with diet, Reynard et al. (2013) first tested this hypothesis by analysing bones of past human populations as a function of age of individuals but no significant trends were observed. This hypothesis was later tested on temporary tooth enamel of modern human individuals with known early life histories (Tacail et al., 2017). Contrary to bone which 504 continuously remodels, tooth enamel forms incrementally. Provided the sampling techniques allow sufficient spatial resolution, the enamel structure permits the reconstruction of the evolution of Ca isotope compositions over time of formation of tooth crowns. This study demonstrates that weaning practices are recorded within human tooth enamel by Ca

2 509 4510 $\frac{6}{9}$ 511 $\frac{8}{9}$ 512 508 isotopes and therefore could allow documenting weaning patterns in past humans and hominids. This method was recently applied to the tooth enamel of Pleistocene early 510 hominins of South Africa (Tacail et al., 2019). The results support that early *Homo* infants 511 were breastfed in significant proportions for longer periods than *A. africanus* and *P. robustus* and opens discussion on the evolution of weaning practices in human lineage.

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514 **3.2.3. Future directions**

 $\frac{15}{16}$ 515 20 517 22 518 $^{24}519$ $^{26}_{27}$ 520 29 521 515 (i) Given the reported systematic variability of Ca dietary sources, and provided this variability is further documented, Ca isotopes could be a successful marker of human dietary practices in relation with human cultures and subsistence modes. A more actualistic approach with a focus on modern and historic human populations with documented and contrasting diets would for instance open the way to new archaeological applications, the same way Zn isotopes were shown to be sensitive to consumption of fish and meat (Jaouen et al., 2018).

31 522 32 33 523 34 $\frac{35}{26}$ 524 36 37 38 525 (ii) As biomarkers of weaning practices, Ca isotopes in tooth enamel would benefit from the investigation of its complementarity with other biomarkers of early life dietary transitions such as Ba/Ca ratios in dentin or enamel (e.g., Austin et al., 2013) or $\delta^{15}N$ in bone and dentin collagen (Tsutaya and Yoneda, 2015).

40 526 42 527 $^{44}_{45}$ 528 $\frac{46}{47}$ 529 49 530 (iii) Beyond tooth enamel, other incrementally growing tissues could be explored for their potential as time resolved archives. Dentin, hair or nail could record Ca isotopic variations relating to weaning practices but also to other physiopathological modifications of Ca homeostasis such as puberty as proposed by Li et al, 2016, the same way other isotope systems can be affected (e.g., Ohno et al., 2005; Tsutaya and Yoneda, 2015).

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532 4. **Calcium isotopes in biomedical research**

 $\frac{55}{56}$ 533 58 534 60 535 533 *Stable Isotope Metallomics* is the study of normal and disrupted cycles of regulated and essential metals from the perspective of their natural stable isotope compositions. This field of research has developed in the last two decades following seminal works on Ca, Fe,

2 537 4538 6 539 $\frac{8}{9}$ 540 11 541 536 Cu or Zn, aided by the development and widespread application of multicollection inductively coupled plasma mass spectrometry (MC-ICPMS) (see reviews such as Albarède et al., 2017; Costas-Rodríguez et al., 2016). This research has served two main purposes: i) to shed new light on the biological processing of these elements, beyond quantitative description of their 540 elemental distributions (*e.g.,* identify pathways or biochemical mechanisms), ii) to develop new markers for rapid and specific diagnosis or prognosis of diseases affecting these cycles.

13 542 155 543 $\frac{17}{10}$ 544 20 545 22 546 24547 $^{26}_{27}$ 548 $\frac{28}{29}$ 549 In this context, the potential of Ca isotopes is important. Because of the significant fractionation occurring during bone mineralisation, Ca stable isotopes are directly indicative of the short-term variations of net bone mineral balance (BMB), i.e. the mass balance between bone resorption and formation. Individuals with positive BMB should thus display blood and urine Ca isotope compositions enriched in heavy isotopes with respect to neutral BMB. On the other hand, individuals with a negative BMB should display ⁴⁴ Ca-depleted blood and urine. This recognition sparked a vivid interest for the study of Ca isotopes as a marker of disruptions of BMB.

31 550 33 551 $\frac{35}{26}$ 552 36 38 553 40 554 42 555 $^{44}_{45}$ 556 $\frac{46}{47}$ 557 47 49 558 51 559 53 560 $\frac{55}{56}$ 561 56 Disruptions of bone mineral balance are widespread in modern societies, mainly because of the high incidence of metabolic bone disease, such as osteoporosis in ageing populations (Becker et al., 2010; Peacock, 2010). Another related issue is the bone loss induced by microgravity, as experienced by crew members in space missions, for which 554 effective countermeasures are still being investigated (Grimm et al., 2016). Clinical tools for predicting and monitoring the evolution of these physiological and pathological conditions resulting from disruptions of Ca metabolism are currently limited to: (i) molecular markers with poor sensitivity or quantitative insight into BMB changes, (ii) X-ray densitometry techniques that only provide a 6-months to yearly resolution of the changes in bone mineral density (e.g., Kuo and Chen, 2017). The potential of Ca isotopes as a quantitative, radiationfree and non-invasive biomarker for the measurement of BMB has been investigated in a series of studies over the last twelve years.

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563 **4.1. The effects of induced bone loss in bed rest studies**

2 565 4566 $\frac{6}{9}$ 567 $\frac{8}{9}$ 568 11 569 13 570 564 Several studies presented evidence of variations in human blood and urine Ca isotope compositions caused by bone loss induced by bed rest (Channon et al., 2015; 566 Heuser et al., 2019; Morgan et al., 2012; Skulan et al., 2007) (see Figure 6). Bed rest provokes significant loss of muscle as well as bone mineral mass in response to inactivity, and mimics the effects of microgravity in spaceflight (Pavy-Le Traon et al., 2007). Bed rest experiments are helpful in identifying the physiopathological causes of bone mineral loss, and also permit the assessment of drug and physical exercise countermeasures to bone mineral loss.

 $18\,572$ 20 573 22 574 $^{24}_{25}$ 575 $\frac{26}{27}$ 576 29 577 31578 $\frac{33}{34}$ 579 36 580 38 581 40582 $\frac{42}{43}$ 583 45 584 47 585 49 586 $\frac{51}{52}$ 587 54 588 56 589 58 590 Skulan et al. (2007) first published a report of significant variations of $\delta^{44/40}$ Ca values in weekly-pooled urine of individuals submitted to bed rest experiments. The authors reported significant negative departure from baseline in 44 Ca 40° Ca ratios during bed rest in the group with no countermeasure to bone loss followed by a return to baseline after the experiment. By contrast, subjects treated with pharmacological or exercise countermeasures to bone loss displayed either no change or positive change of their urine $\delta^{44/40}$ Ca values. 578 These results provided a first proof of concept. This work was later augmented by Morgan *et* al. (2012) with a report of changes in urine Ca isotope compositions during a bed rest experiment conducted with a finer chronological sampling. This study demonstrated that the decrease in $\delta^{44/42}$ Ca values of urine occur after one week, indicating a loss of bone in agreement with markers of osteolytic activity (NATX) but long before any changes in bone mineral density could be detected by means of X-ray methods. These results were further confirmed in the blood of the same subjects (Channon et al., 2015). The recognition of the isotopic fractionation induced by renal function lead the authors to adjust the mathematical 586 models used to link changes in bone mass and urine Ca isotope compositions (*e.g.,* Heuser and Eisenhauer 2010, Morgan et al., 2012). The observed negative offset in urine $\delta^{44/42}$ Ca results from loss of Ca from bone but also from an increase in Ca urinary losses in kidneys 589 (*i.e.* a decrease in Ca reabsorption). Taking this into account, Morgan et al. (2012) calculated the relative bone loss during the bed rest experiment. The estimated relative bone loss

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2 592 4593 591 agreed with bone loss predicted on the basis of previous experiments in which bed rest was of a duration long enough to produce changes in BMD resolvable by X-ray densitometry (around 0.25 % of total bone mass, for -0.30% decrease of $\delta^{44/42}$ Ca values).

 $\frac{6}{7}$ 594 $\frac{8}{9}$ 595 11 596 13597 $\frac{15}{16}$ 598 18 599 20 600 22601 More recently, in connection with another bed rest experiment, Heuser et al. (2019) proposed a methodology to deconvolute the combined effects of changes in renal Ca excretion and bone loss. Using both the urine Ca isotope compositions and the measured 597 daily Ca urinary excretion rates, the authors distinguish the contribution of bone loss to the variations in urinary Ca isotope compositions while cancelling out the effects of kidney Rayleigh-type isotope distillation. These results stress the need for the development a more thorough understanding of influence of renal function in the variability of urine Ca isotope compositions while emphasizing the potential of $\delta^{44/42}$ Ca as a diagnostic biomarker.

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603 **4.2. Calcium isotopes as biomarkers of metabolic bone diseases**

29 604 31605 Calcium isotopes are sensitive to other bone related disorders. Their efficiency as a biomarker (i.e. sensitivity and specificity) was tested in two published clinical studies so far.

 $\frac{33}{24}$ 606 36 607 38 608 40 609 $^{42}_{42}$ 610 45 611 47 612 49 613 51614 Gordon et al. (2014) reported significant ⁴⁴ Ca-depletion of blood serum Ca in subjects affected by active forms of multiple myeloma when compared to subjects with non-active disease. Multiple myeloma is a form of cancer affecting the plasma cells and inducing osteolytic lesions, leading to osteoporosis and bone fracture. While current methods can 610 detect osteolytic lesions only after they have caused significant damage, Ca isotopes could reveal abnormal bone loss, and thus disease activity, long before it can be detected by other means. This preliminary study reported good performances for Ca isotopes as a disease predictor, with significant specificity and sensitivity, regardless of other factors, such as age or gender.

 $\frac{53}{54}$ 615 56 616 58 617 60 618 Based on those preliminary yet promising first results, Eisenhauer et al. (2019) reported the first clinical trial conducted to assess the potential of Ca isotopes as a diagnosis biomarker for osteoporosis in 80 postmenopausal women. Compared with the outcomes of the gold-standard densitometry diagnosis method, the blood and urine Ca isotope

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 $\frac{6}{9}$ 622 619 compositions were significantly 44 Ca-depleted in the 14 subjects effectively affected by 620 osteoporosis, when compared to subjects without osteoporosis. Calcium isotopes appear to display very strong sensitivity and good specificity to the osteoporosis condition, and are in good agreement with various classical clinical parameters and chemical biomarkers of bone loss. These results suggest that Ca isotopes could be developed into a stand-alone clinical test for osteoporosis.

626 **4.3. Future directions**

All the work to date has demonstrated that calcium isotopes have characteristics of an efficient isotopic biomarker for bone loss. The clinical applications of Ca isotopes remain in their infancy, but these promising results should stimulate more research to improve diagnostic protocols and develop broad clinical reference databases.

 $\frac{28}{29}$ 632 31 633 33 634 $\frac{35}{26}$ 635 631 *(i.)* Despite considerable progress, the understanding of how Ca isotopes correlate with epidemiology of metabolic bone diseases could be improved. Continued efforts will be required to advance our understanding of the Ca isotopes cycle in the whole organism both in health and disease. Further characterization of the mechanisms of isotope fractionation at the scale of organs and cells will also greatly benefit to this field of research.

38 636 40 637 42 638 $^{44}_{45}$ 639 $\frac{46}{47}$ 640 49 641 51 642 53643 $\frac{55}{56}$ 644 (ii.) The reported intra- and inter-individual variability of blood or urine Ca isotope compositions highlights the need for identifying factors of variability other than bone loss. Besides dietary practices varying through time in a given individual and/or from one individual to another, several nutrients and molecules are known to affect Ca homeostasis, and thus potentially intra- and inter-individual variability of their Ca isotope compositions. For instance, dietary Vitamin D has notably been shown to result in changes of urine Ca isotope compositions (Rangarajan et al., 2018). Clearly, strategies need to be implemented in order to take these sources of intra- and inter-individual variability into account and/or cancel them out.

60 646 (iii.) Other physiopathological conditions affecting Ca homeostasis could be successfully explored with the help of Ca isotopes including diseases or deficiencies that strongly affect

2 648 4649 $⁶$ 650</sup> $\frac{8}{9}$ 651 11 652 647 the Ca cycle and have high incidence in modern societies (Peacock, 2010; Shroff et al., 2013). For instance, the progressive disruption of the kidney function in individuals suffering from chronic kidney diseases is likely to affect the distribution of Ca isotopes. Other disturbances of the Ca homeostasis being either induced by drugs or in some digestive disorders could also be explored with Ca isotopes. Dedicated research could also potentially lead to the development of new biomarkers for such conditions of disrupted Ca homeostasis.

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654 5. **Challenges of vertebrate Ca isotope geochemistry**

20 656 22 657 The potential of Ca isotopes is high in the field of vertebrate biology. However the development of new applications and their possible adoption by scientific communities other than Earth scientists faces barriers, and depend on the communities involved.

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659 **5.1. Developing the use of Ca isotopes in palaeobiology and bioarchaeology**

 $\frac{28}{29}$ 660 31 661 33 662 $\frac{35}{26}$ 663 38 664 The use of stable isotopes in palaeobiology and bioarchaeology is not new. From the 661 1970's onwards, palaeontologists, paleoanthropologists and archaeologists (as well as 662 modern ecologists and forensic scientists) used *traditional* stable isotopes (C, H, O, N) to assess trophic ecology, habitat use, physiology or migration of past and present vertebrates including humans (Koch, 2007; Newsome et al., 2010).

40 665 42 666 $^{44}_{45}$ 667 $\frac{46}{47}$ 668 49 669 51 670 53671 $^{55}_{56}$ 672 58 673 60 674 Beyond the currently investigated scientific questions regarding Ca isotopes, the main leap one could identify lies in the practical and possibly poorly identified differences between 667 these so-called *traditional* stable isotopes and the developing *non-traditional* metal stable isotopes. First, while *traditional* stable isotopes are routinely analysed by means of gas 669 source mass spectrometers, *non-traditional* stable isotopes, including Ca, require the use of different mass spectrometers (mainly MC-ICP-MS but also possible with TIMS) that are so far predominantly hosted by Earth sciences laboratories and require different analytical skills and laboratory set-ups. These analytical challenges can be overcome, as exemplified by the growing use of other isotope systems requiring these analytical set-ups, such as 87 Sr $/86$ Sr 674 ratios in archaeology. Clear communication on differences between *traditional* and *non-*

2 676 4677 $\frac{6}{9}$ 678 $\frac{8}{9}$ 679 11 680 13 681 $15/15$ 682 17 18 20 684 22 685 24686 675 *traditional* stable isotopes is also a key to successful collaborations with communities of palaeobiology and bioarchaeology scientists. For instance, as a major element, Ca allows favourable sampling strategies of osteological remains (small sample sizes, less destructive analyses, high spatial resolution) and grants access to geological periods so far poorly 679 explored with *traditional* stable isotopes. Also, the elements in the focus of *traditional* stable isotope systems do not share the same biological cycles and functions in vertebrate organisms as do metal stable isotopes. While the former constitutes the very structure of a variety of soft tissues building blocks, the more specific functional roles of bio-essential metals allow focussing on new aspects of the biology of vertebrates. The developing use of Ca isotopes by these communities can thus count on pre-existing fruitful exchanges and collaborations between isotope geochemists and specialists of these disciplines but also require a communication effort regarding Ca isotopes and their peculiarities.

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 $\frac{28}{29}$ 688

688 **5.2. Using Ca isotopes from fundamental to applied biomedical research**

31 689 33 690 $\frac{35}{26}$ 691 38 692 40 693 42 694 $^{44}_{45}$ 695 $\frac{46}{47}$ 696 49 697 51 698 ⁵³ 699 $\frac{55}{56}$ 700 58 701 When taking the plunge into biomedical research and aiming at developing diagnosis 690 biomarkers, the challenges are more acute. The natural variations of *traditional* stable isotope ratios are not routinely used by biomedical research scientists or medical doctors in order to understand element cycling, characterize aetiology of diseases, nor as biomarkers, notably because of their limited specificity. A few groups of isotope scientists and biomedical 694 researchers currently investigate the potential of metal stable isotope systems (*e.g.,* Ca, Fe, Cu or Zn, see reviews such as Costas-Rodríguez et al., 2016; Albarède et al., 2017). However, this field of research is currently in its infancy. More generally, all involved communities (stable isotope scientists, biomedical scientists and medical doctors) do not have a shared scientific or technical background and may have different goals. Beyond purely scientific questions, there are potential intellectual and cultural barriers that must be 700 overcome in order to implement the truly interdisciplinary collaboration that biomedical stable isotope research demands.

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703 **5.2.1. Initiating successful interdisciplinary research projects**

2 704 4705 $\frac{6}{9}$ 706 $\frac{8}{9}$ 707 11 708 13 709 15 710 As recently reported by Sauzéat et al. (2019), interdisciplinary collaboration requires reciprocal curiosity and communication between fields, which in turn requires scientists to understand each other's scientific and technic cultures well enough to identify scientific questions that could be addressed collaboratively. Scientists must endeavour to make their work accessible to people in other fields. For isotope scientists, this implies for instance 709 avoiding unnecessary jargon and preparing clearly written documents, or *vade-mecum,* that explain key concepts of stable isotope metallomics to non-specialists.

17 18 20 712 22 713 24714 $\frac{26}{27}$ 715 29 716 31 717 Also, the fates of such new frontiers research projects are unknown at first and can be seen as risky. It is important to encourage and support the researchers that develop interdisciplinary careers. While the risk is that such profiles are potentially hard to fit in job 714 descriptions, researchers with multiple backgrounds (*e.g.,* medical doctors or biomedical researcher skilled as isotope scientist) pave the way to the future of this research. In this matter, the support of institutions is particularly valuable, when for example implementing interdisciplinary grant schemes and recruitment campaigns.

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719 **5.2.2. Analytical and methodological challenges**

37 38 720 39 40 721 41 42 722 43 $^{44}_{45}$ 723 45 $\frac{46}{47}$ 724 47 48 49 725 50 51 726 52 53 727 54 $\frac{55}{56}$ 728 56 57 58 729 59 60 730 61 Another challenge is to identify the methodological constraints imposed by the various disciplines involved in the project, and to accommodate these constraints in the study design. For example, the extreme care typically required to maintain sterility and integrity of biological samples for molecular biological assays is generally unnecessary for samples destined for isotope analysis. On the other hand, substances commonly used as preservatives in blood and urine samples can be unacceptable contaminants for isotope analysis. Disciplines also have different requirements for experimental controls. In human experiments involving Ca isotopes, this typically requires sampling and analysis of all dietary and pharmacological Ca sources, as well as multiple pre-experimental sampling of all study subjects in order to establish individual baselines. Finally, isotope geochemistry laboratories are not biomedical laboratories. Ethical and health regulations often require formal

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2 732 4 733 $\frac{6}{9}$ 734 $\frac{8}{9}$ 735 11 736 13 737 $15/738$ 17 18 731 authorization for the handling of human tissues, as well as adaptation of laboratory practices and proper training of isotope scientists, engineers, technicians or students in the hazards posed by these materials. Also, isotope analyses in isotope geochemistry laboratories tend to be slow when compared to routine biomedical analyses, even when machine time is not limited. For Ca isotopes to become a widely used clinical tool, sample throughputs must be increased by several orders of magnitude, while ensuring a sufficient precision and accuracy 737 of the isotope analysis. This is technically feasible (with approaches such as automated sample preparation, spiking methods, collision-cell instruments, e.g., Romaniello et al., 2015, Lewis et al., 2018) but will not happen without being spurred by demand.

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741 **5.2.3. Developing a medical biomarker**

 24 742 $^{26}_{27}$ 743 $\frac{28}{29}$ 744 31 745 33 746 $\frac{35}{26}$ 747 In general, the development of medical biomarkers is codified and standardized, for instance in terms of clinical trials methodology and statistics or ethical and legal aspects. These are constraints for which isotope geochemists, and their host laboratories and institutions, are neither prepared nor able to address on their own. The first challenge is thus to recruit the assistance of medical scientists in the regulatory and administrative aspects of study design.

38 748 39 40 749 42 750 43 $^{44}_{45}$ 751 45 $\frac{46}{47}$ 752 47 48 49 753 50 51 754 52 53755 54 $\frac{55}{56}$ 756 56 In addition to aforementioned challenges, we suggest that another lies in the differences in technical and scientific cultures in which stable isotope geochemists and medical scientists have evolved. For example, stable isotope geochemists tend to direct their research toward a fundamental understanding of processes while medical scientists and hospital doctors may feel more constrained by the need for practical and immediate tools to achieve clinical goals. This urgency is something geochemists do not often encounter on their home turf. As a result, because geochemists tend to assume that practical application proceeds from theoretical understanding, they could tend to underestimate the current clinical utility of their techniques and undersell them.

58 757 60 758 Finally, medical science is immersed in a biological paradigm that views life as an interaction between complex organic macromolecules (including genes) at the cellular level.

 1 2 760 4761 5 $\frac{6}{7}$ 762 $\frac{8}{9}$ 763 9 10 11 764 13 765 14 759 Metal stable isotope analysis, focused as it is on inorganic chemistry, does not mesh well with this paradigm. Apart from the practical clinical tools they can provide, isotope chemists also are in a position to advance a long overdue appreciation of the central role purely inorganic processes play in biology. It is possible to say a great deal about the state of an 763 organism by looking at the distribution of simple ions and isotopes, without reference to cells or macromolecules, in the same way that it is possible to say a great deal about an 765 environmental or geological system without reference to the details of its component parts.

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63 64 65

1157 **Captions:**

Figure 1: Number of publications per year reporting new research on Ca isotopes in vertebrates (Single column)

Figure 2: Typical Ca cycle in vertebrate organisms. The Ca fluxes for which Ca isotope fractionation is documented, possible or not suspected are indicated with orange dashed arrows, black dashed arrows and black solid line respectively. (2-column)

Figure 3: δ^{44/42}Ca of bone of 7 species of vertebrates as a function of their diet $δ^{44/42}$ Ca (in ‰, relative to SRM915a reference material) shows the shared physiological isotope effect. The red dashed line is the average bone-diet offset, the blue full line is the linear regression and the red shaded area delimits its 95% confidence interval. Error bars are 2sd. ¹ Skulan and DePaolo, 1999; ² Chu et al., 2006; ³ Heuser et al., 2016; ⁴ Tacail et al., 2014; ⁵ Hirata et al., 2008. (Single column)

Figure 4: Ecological reconstructions of a modern mammal ecosystem (Tsavo National Park, Kenya, dataset from Martin et a., 2018): A. Tooth enamel $\delta^{44/42}$ Ca (in ‰, relative to SRM915a reference material) as a function of $\delta^{13}C$ (in ‰, relative to VPDB reference material) 1176 (Modified from Martin et al., 2018). B. Principal Component Analysis (PCA) of various chemical proxies in tooth enamel (grouped by main ecological groups and Hippos, as described in Martin et al., 2018, displayed with 95% distribution ellipses). The δ^{13} C- $\delta^{44/42}$ Ca space summarizes well the ecosystem structure clearly depicted by the PCA, while hippos $\frac{51}{52}$ 1180 stand as outliers that remain to be explained. (2-column)

Figure 5: Compilation of Ca isotope compositions (in ‰, relative to SRM915a reference material) of a series of Ca sources in diets of vertebrates including human. This compilation

 6^{4} ا و $15₁$ $17₄$ $18¹$ includes continental river water considered as fresh water (Tipper et al., 2016), marine resources (fish, mollusc and crustaceans) (Skulan et al., 1997; Skulan and DePaolo, 1999), dicotyledon roots, stems and leaves or fruits and monocotyledon including cereals (Christensen et al., 2018; Chu et al., 2006; Farkaš et al., 2011; Gussone and Heuser, 2016; $\frac{8}{9}$ 1188 Heuser, 2016; Hindshaw et al., 2013; Holmden and Bélanger, 2010; Huang et al., 2012; Moore et al., 2013; Page et al., 2008; A. Schmitt et al., 2003; Skulan and DePaolo, 1999; Tacail et al., 2014; Wiegand et al., 2005), hen and quails' eggs (Skulan and DePaolo, 1999; Tacail, 2017), herbivore's soft tissues and blood (Heuser, 2016; Morgan et al., 2012; Skulan and DePaolo, 1999; Tacail et al., 2014), animal milk and dairy products (Chu et al., 2006; Gussone and Heuser, 2016; Heuser, 2016; Tacail et al., 2017) and modern human milk (Chu et al., 2006; Tacail et al., 2017). (2-column)

 $27'$ $33₁$ $36¹$ 1203 $44.$ $45''$ Figure 6: Bedrest experiments induce bone loss and a decrease of urine and blood $\delta^{44/42}$ Ca values. Changes in Ca isotope compositions $(\Delta^{44/42}$ Ca) of blood or urine at each date of experiment are calculated as the difference with average baseline (pre-bedrest) composition in all subjects of three studies: (A.) $\Delta^{44/42}$ Ca in urine of 4 individuals over 17 weeks of bedrest (Skulan et al., 2007), (B.) $\Delta^{44/42}$ Ca in urine of 6 individuals over 21 days of bedrest (Heuser et al., 2019), (C.) $\Delta^{44/42}$ Ca in urine (top) and blood (bottom) of 12 individuals over 30 days of bedrest (Morgan et al., 2012, Channon et al., 2015). Graphs display boxplots of $\Delta^{44/42}$ Ca values, indicating 5, 25, 50, 75 and 95% quantiles. Red diamonds and red dashed lines show the evolution of average $\Delta^{44/42}$ Ca values with time. (2-column)

 1205

 $^{24}_{\circ}$ 1195

Figure 1

Figure 4

Figure 6

